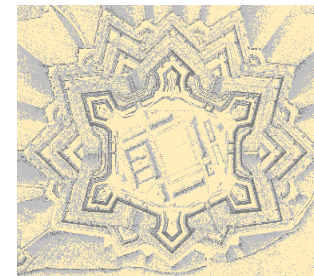


Università del Piemonte Orientale
"Amedeo Avogadro"



Alessandria: La Cittadella

Intensive Programme

"Design, Synthesis and Validation of Imaging Probes"

Torino – September 19-30, 2011

**Gd^{III} complexes: mechanisms of
action and optimization strategies**

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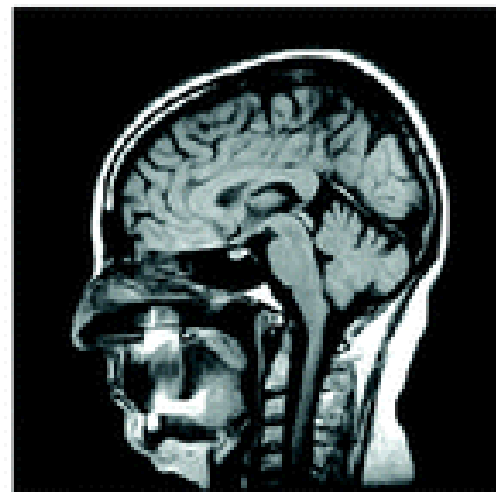
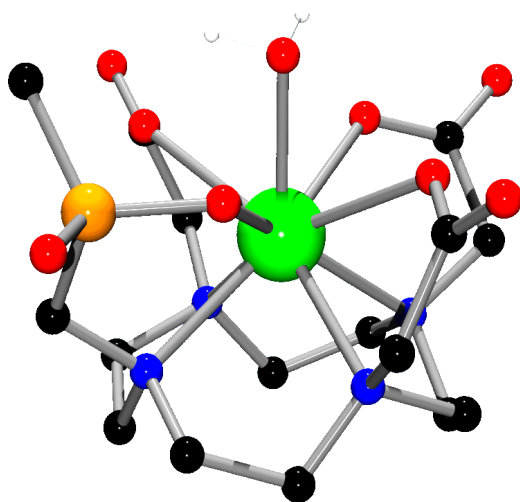
mauro.botta@mfn.unipmn.it



Lanthanides in MRI

“More than 10 million MRI studies are performed with Gd each year”

(P. Caravan, *Chem. Soc. Rev.*, 2006, 35, 512)

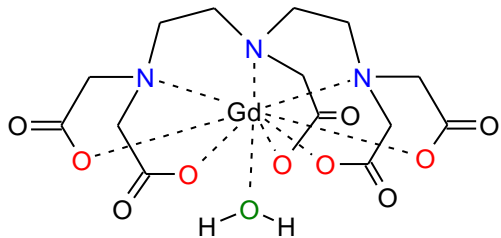


Gd-based T_1 contrast agents:

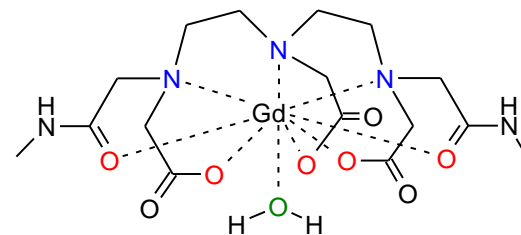
- Increase signal intensity on T_1 -weighted images
- Shorten acquisition time
- Improve diagnostic confidence

P. Caravan, *Contrast Media Mol. Imaging.*, 2009, 4, 89

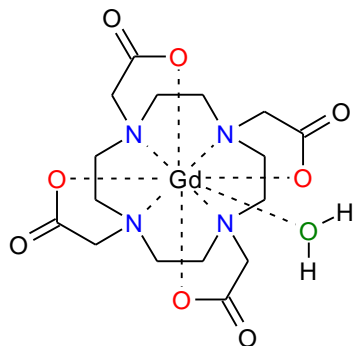
Commercial Agents



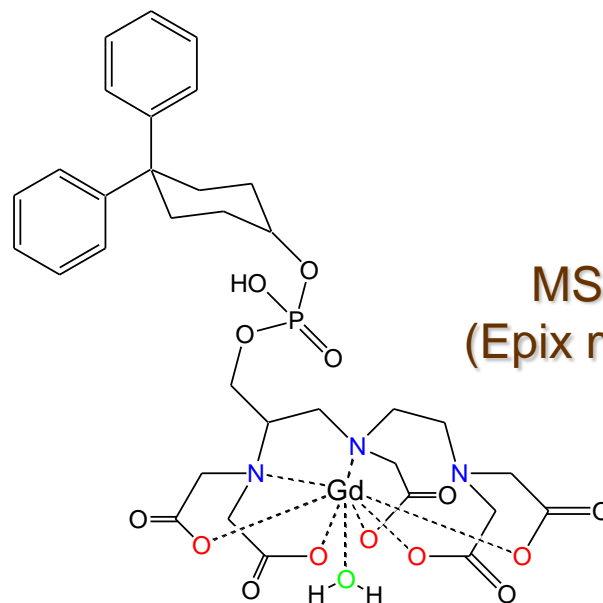
[Gd(DTPA)(H₂O)]²⁻
(Magnevist: Berlex/Schering)



[Gd(DTPA-BMA)(H₂O)]
(Omniscan: Nycomed-Amersham)



[Gd(DOTA)(H₂O)]⁻
(Dotarem: Guerbet)

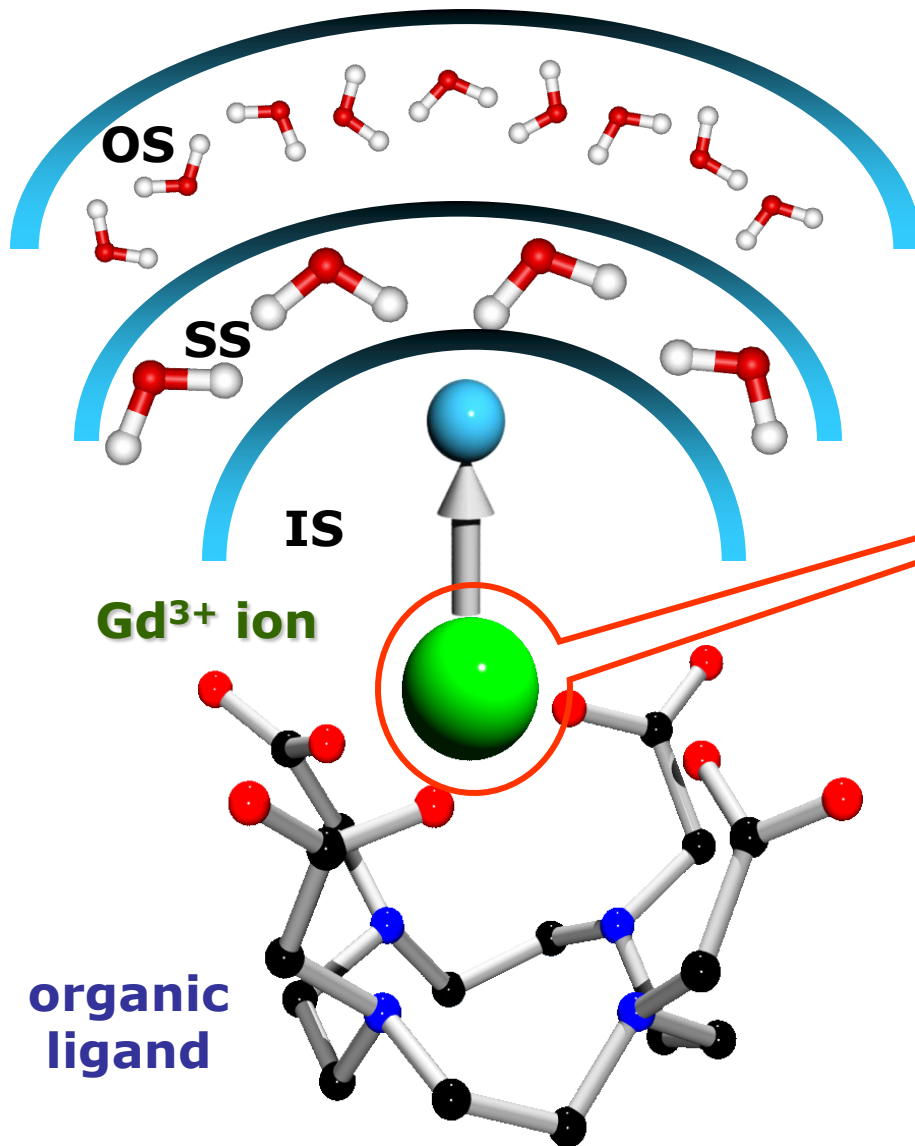


MS-325
(Epix medical)

Clinically approved Gd^{III}-based contrast agents

Complex	Brand name	Company	Log K_{GdL}
GdDTPA	Magnevist[®]	Schering	22.5
GdDOTA	Dotarem[®]	Guerbet	24.7
GdHPDO3A	ProHance[®]	Bracco	23.8
GdDO3A-butrol	Gadovist[®]	Schering	20.8
GdDTPA-BMA	Omniscan[®]	Nycomed- Amersham	16.8
GdDTPA-BMEA	OptiMARK[®]	Mallinckrodt	16.8
GdBOPTA	MultiHance[®]	Bracco	22.6
GdEOB-DTPA	Eovist[®]	Schering	23.5

"Anatomy" of a MRI contrast agent



H																			He
Li	Be											B	C	N	O	F		Ne	
Na	Mg											Al	Si	P	S	Cl		Ar	
K	Ca	Sc	Ti	V	Cr	Mn	Fe	Co	Ni	Cu	Zn	Ga	Ge	As	Se	Br		Kr	
Rb	Sr	Y	Zr	Nb	Mo	Tc	Ru	Rh	Pd	Ag	Cd	In	Sn	Sb	Te	I		Xe	
Cs	Ba		Hf	Ta	W	Re	Os	Ir	Pt	Au	Hg	Tl	Pb	Bi	Po	At		Rn	
Fr	Ra		Rf	Db	Sg	Bh	Hs	Mt	Uun	Uuu	Uub								
			La	Ce	Pr	Nd	Pm	Sm	Eu	Gd	Tb	Dy	Ho	Er	Tm	Yb		Lu	
			Ac	Th	Pa	U	Np	Pu	Am	Cm	Bk	Cf	Es	Fm	Md	No		Lr	

- 7 unpaired e⁻
- Slow electron spin relaxation
- High magnetic moment

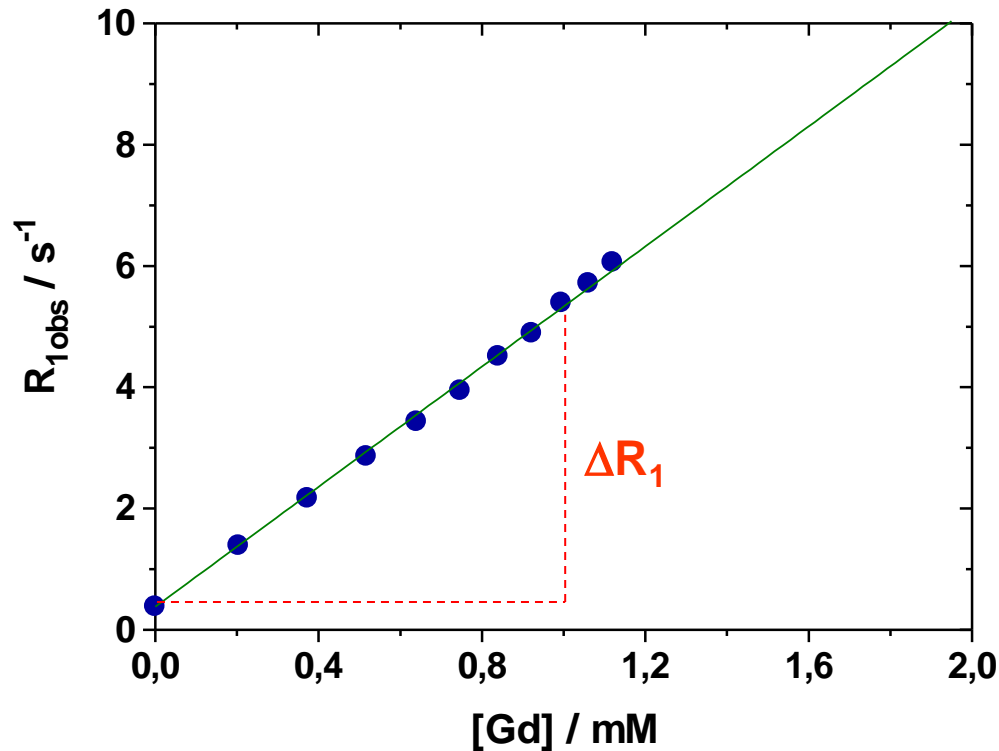
relaxivity – r_1 [mM⁻¹ s⁻¹]



$f(B, T, q, r, \tau_R, \tau_M, T_{e1,2})$

Relaxivity: definitions

$$R_i = [CA]r_{ip} + R_{iw} \quad i = 1,2$$



20 MHz; 298 K

$$r_{1p} = 4.95$$

$$R_{1w} = 0.38$$

Commercial Agents

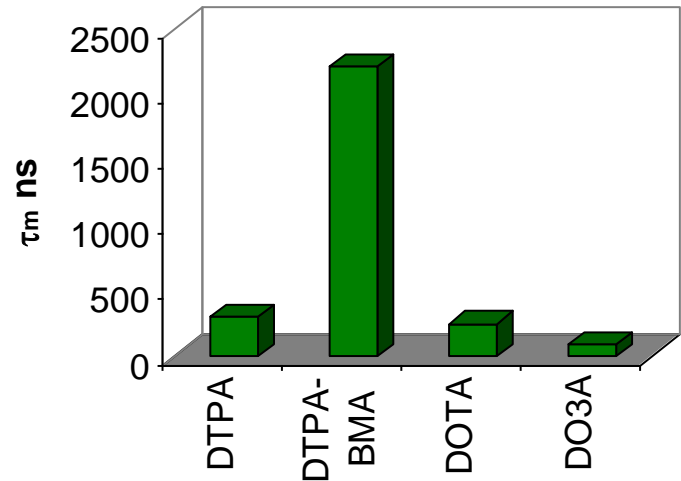
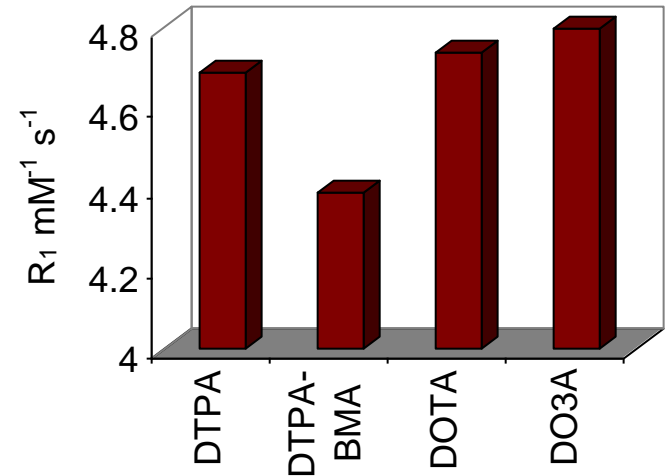
Low relaxivity:

polyaminocarboxylates with $r_1 = 4 - 5 \text{ mM}^{-1}\text{s}^{-1}$:

- only one inner sphere coordinated water molecule: $q = 1$
- slow water exchange rate : $\tau_M = 150$ to 2500 ns
- fast tumbling molecule : $\tau_R \approx \text{ps}$

Disadvantages:

- injection of gram quantities of Gd complex
- limited to imaging blood streams / liver & kidneys
- do not allow site specific targeting

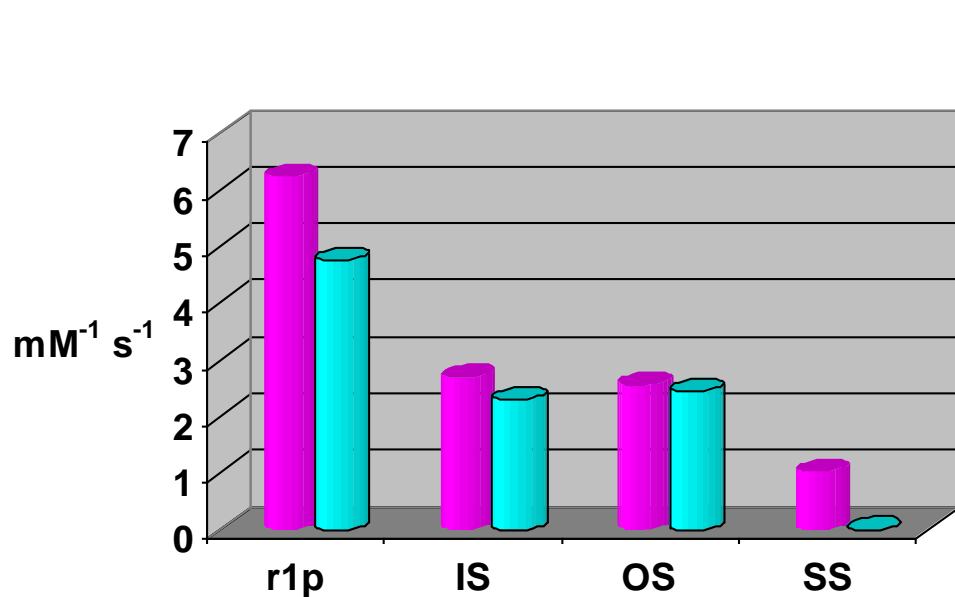


Caravan, P et al. *Chem. Rev.* **1999**, 99, 2293-2352.

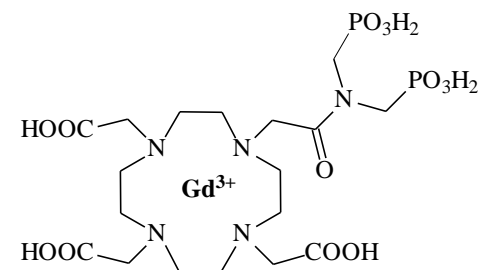
contributions to relaxivity

$$r_i = r_{ip}^{IS} + r_{ip}^{OS} + r_{ip}^{SS} \quad (i = 1, 2)$$

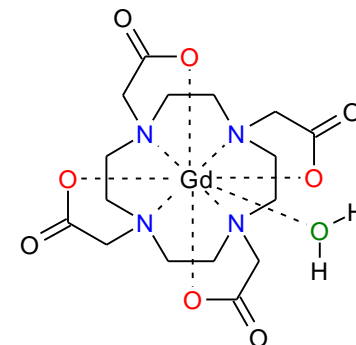
- need to specify magnetic field strength and temperature (... and pH)



20 MHz – 298 K



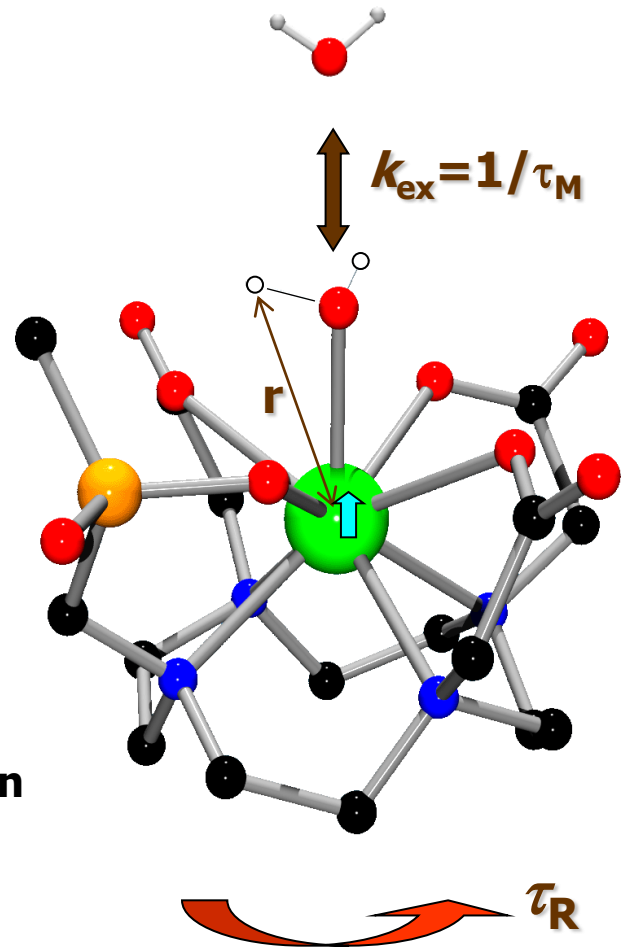
● DO3A-NP2
● DOTA



Inner-sphere contribution to the relaxivity

$$r_{1p}^{IS} = \frac{[CA]q}{55.6} \frac{1}{T_{1M} + \tau_M}$$

- q : n° of metal-bound water molecules
- τ_M : their mean residence lifetime
- T_{1M} : the longitudinal nuclear magnetic relaxation time of the bound water protons



Solomon-Bloembergen-Morgan (SBM) equations

$$\frac{1}{T_{1M}} = \frac{2}{15} \frac{\gamma_H^2 g^2 S(S+1) \beta^2}{r_H^6} \left[\frac{3\tau_{C1}}{1 + \omega_H^2 \tau_{C1}^2} + \frac{7\tau_{C2}}{1 + \omega_S^2 \tau_{C2}^2} \right]$$

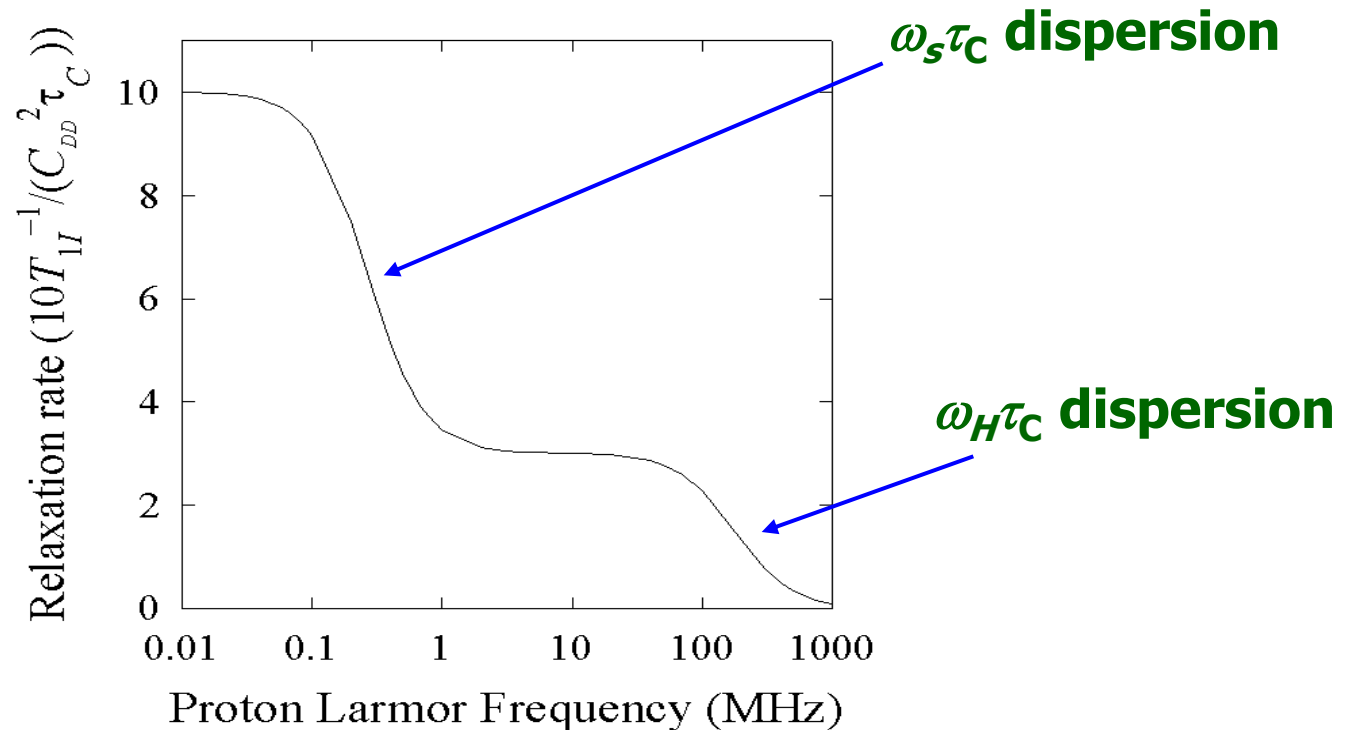
$$\frac{1}{\tau_{Ci}} = \frac{1}{\tau_R} + \frac{1}{\tau_M} + \frac{1}{T_{ie}}$$

$$\left(\frac{1}{T_{1e}} \right)^{\text{ZFS}} = \frac{1}{25} \Delta^2 \tau_v \{4S(S+1) - 3\} \left(\frac{1}{1 + \omega_S^2 \tau_v^2} + \frac{4}{1 + 4\omega_S^2 \tau_v^2} \right)$$

$$\left(\frac{1}{T_{2e}} \right)^{\text{ZFS}} = \Delta^2 \tau_v \left[\frac{5.26}{1 + 0.372\omega_S^2 \tau_v^2} + \frac{7.18}{1 + 1.24\omega_S^2 \tau_v^2} \right]$$

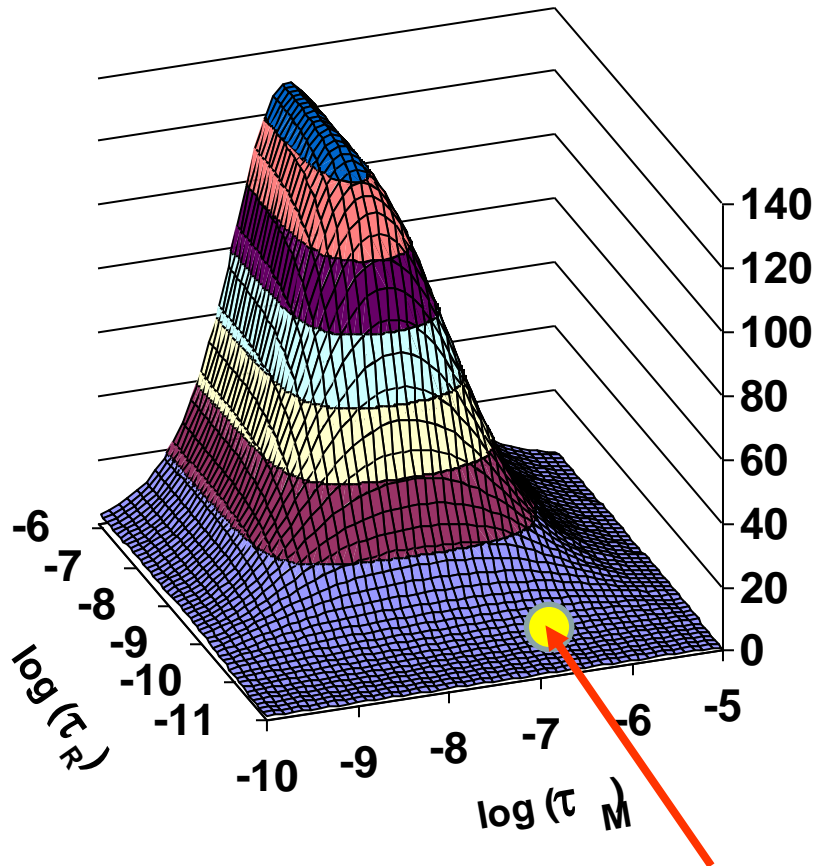
Solomon equation

$$\frac{1}{T_{1M}} = \frac{2}{15} \frac{\gamma_H^2 g^2 S(S+1) \beta^2}{r_H^6} \left[\frac{3\tau_{C1}}{1 + \omega_H^2 \tau_{C1}^2} + \frac{7\tau_{C2}}{1 + \omega_S^2 \tau_{C2}^2} \right]$$

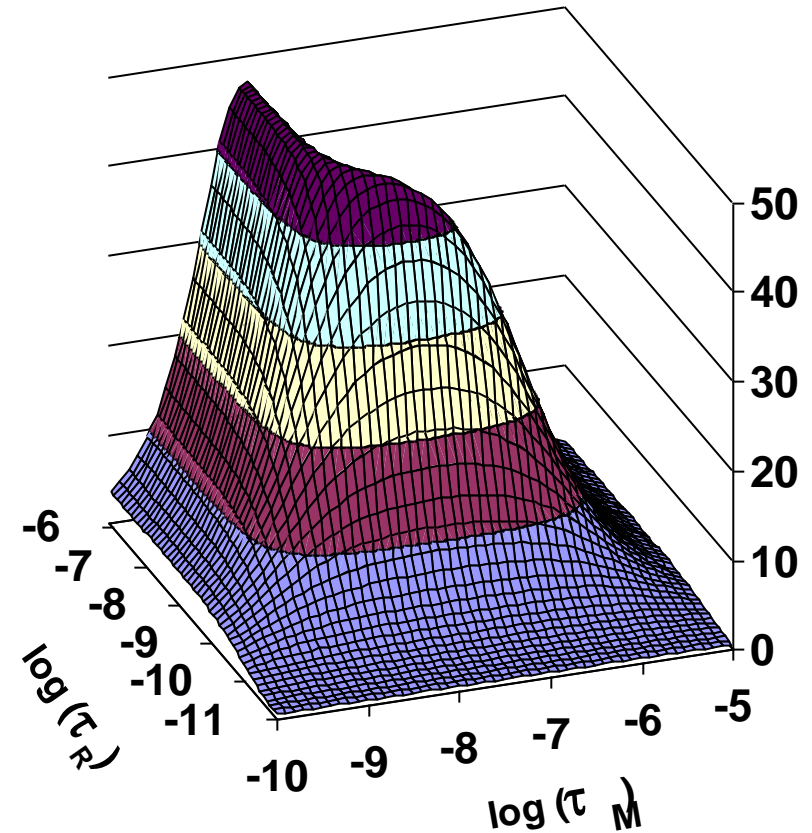


Dependence of r_{1p} on molecular parameters and B_0

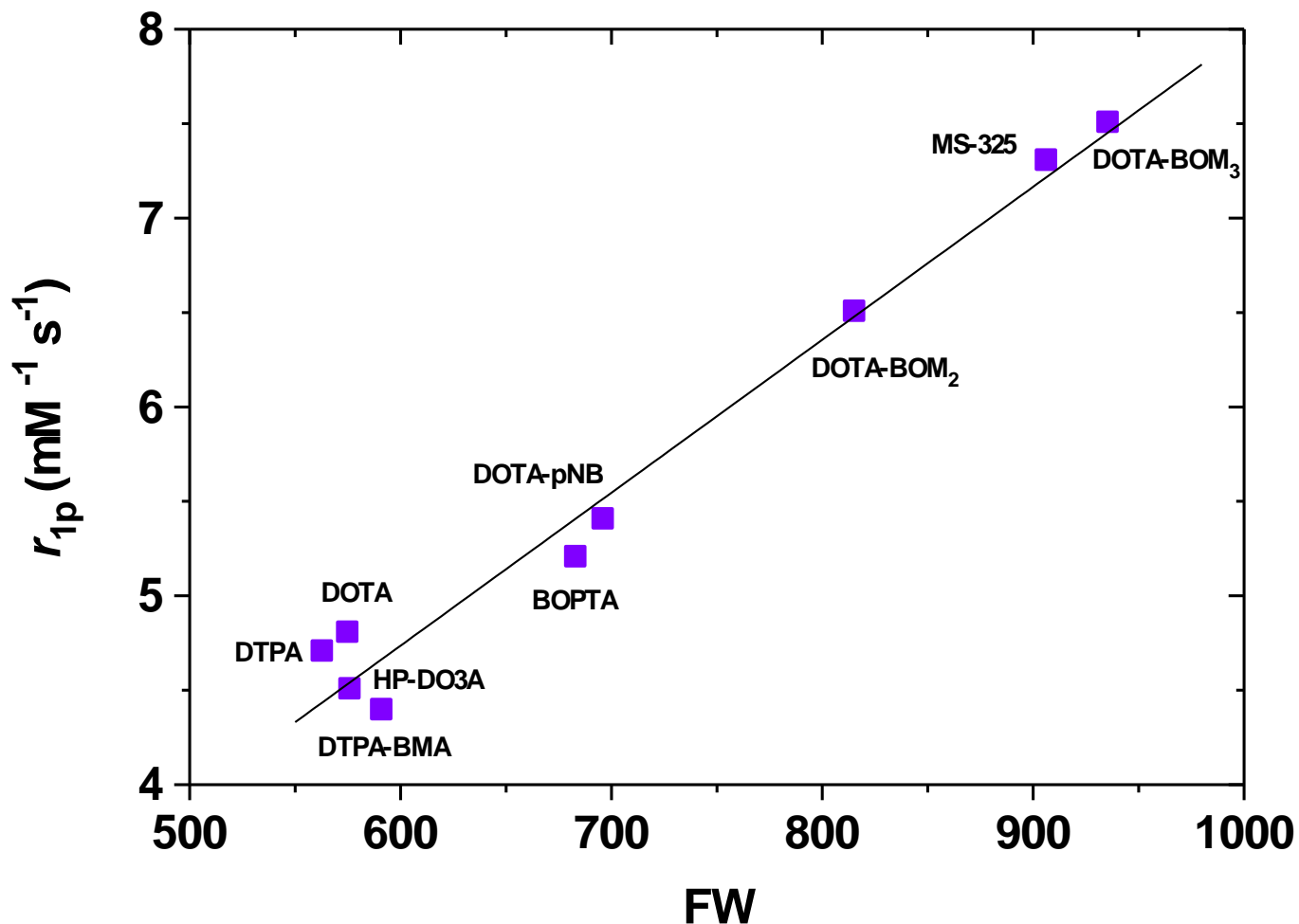
0.5 T / $q=1$ / $r = 3.0 \text{ \AA}$



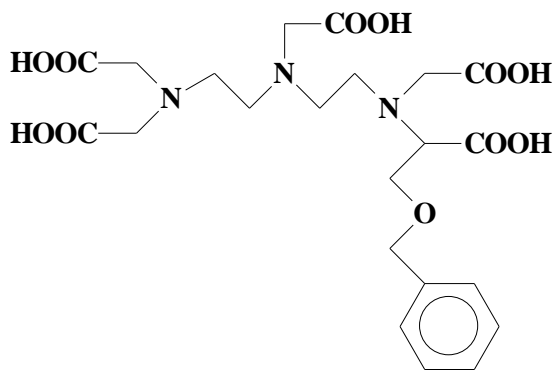
1.5 T / $q=1$ / $r = 3.0 \text{ \AA}$



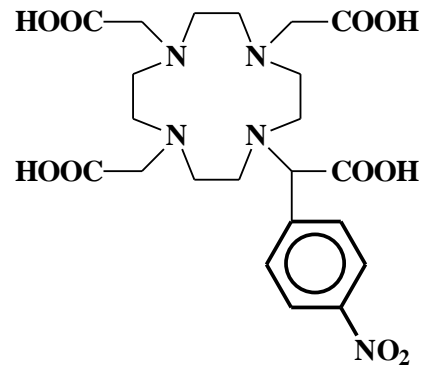
Rotational dynamics



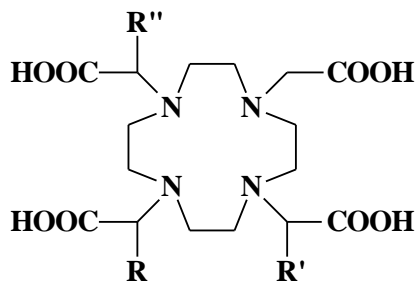
Plot of the relaxivity (20 MHz and 25°C) for monoaquo polyaminocarboxylate Gd(III) complexes versus molecular weight. The correlation coefficient R is 0.988



BOPTA

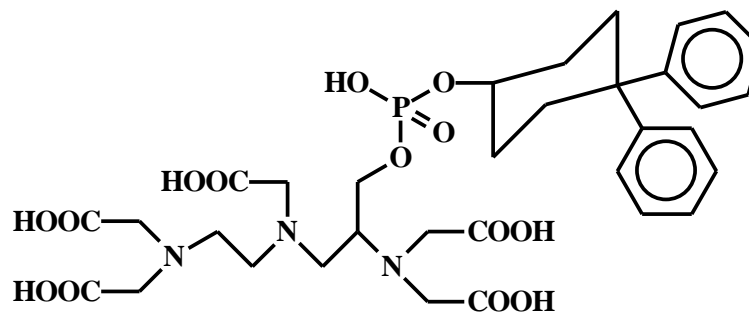


DOTA-pNB



DOTA-(BOM)₂
 (R = H, R', R'' = -CH₂OCH₂C₆H₅)

DOTA-(BOM)₃
 (R, R', R'' = -CH₂OCH₂C₆H₅)

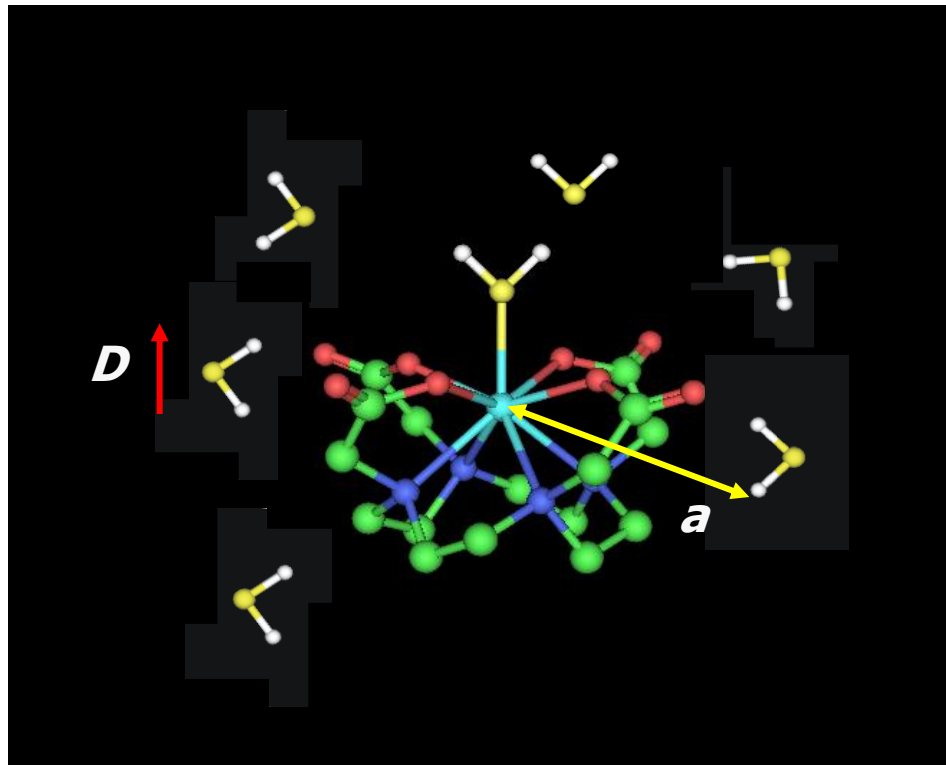


MS-325

Outer-sphere contribution to the relaxivity

→ **translational diffusional model of Freed**

modulation of the dipolar interaction of the paramagnetic centre with the water molecules diffusing next to the surface of the complex

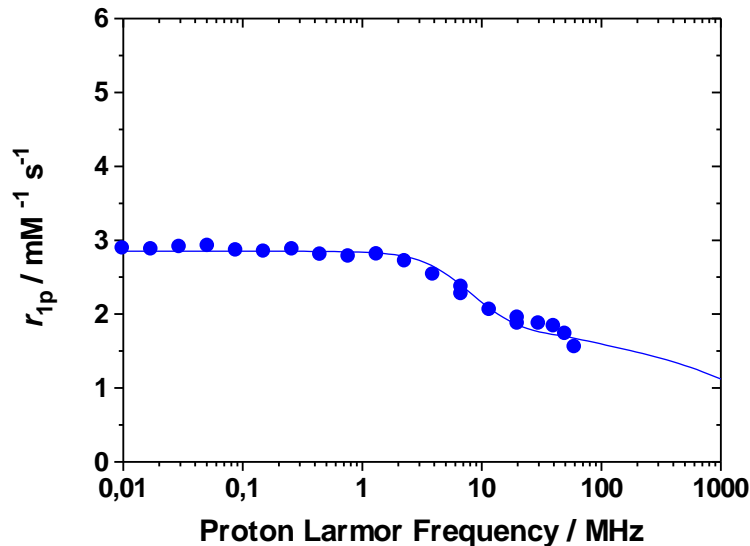


Outer-sphere contribution to the relaxivity

$$r_{1p}^{OS} = C^{OS} \left(\frac{1}{aD} \right) [7J(\omega_S) + 3J(\omega_H)]$$

$$J(\omega) = \text{Re} \left[\frac{1 + 1/4 \left(i\omega\tau_d + \frac{\tau_d}{T_{je}} \right)^{1/2}}{1 + \left(i\omega\tau_d + \frac{\tau_d}{T_{je}} \right)^{1/2} + 4/9 \left(i\omega\tau_d + \frac{\tau_d}{T_{je}} \right) + 1/9 \left(i\omega\tau_d + \frac{\tau_d}{T_{je}} \right)^{3/2}} \right]$$

with $j = 1, 2$; $\tau_d = a^2/D$



$$q = 0$$

$$a = 4.2 \text{ \AA}$$

$$D = 2.24 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$$

298 K

Outer-sphere contribution to the relaxivity

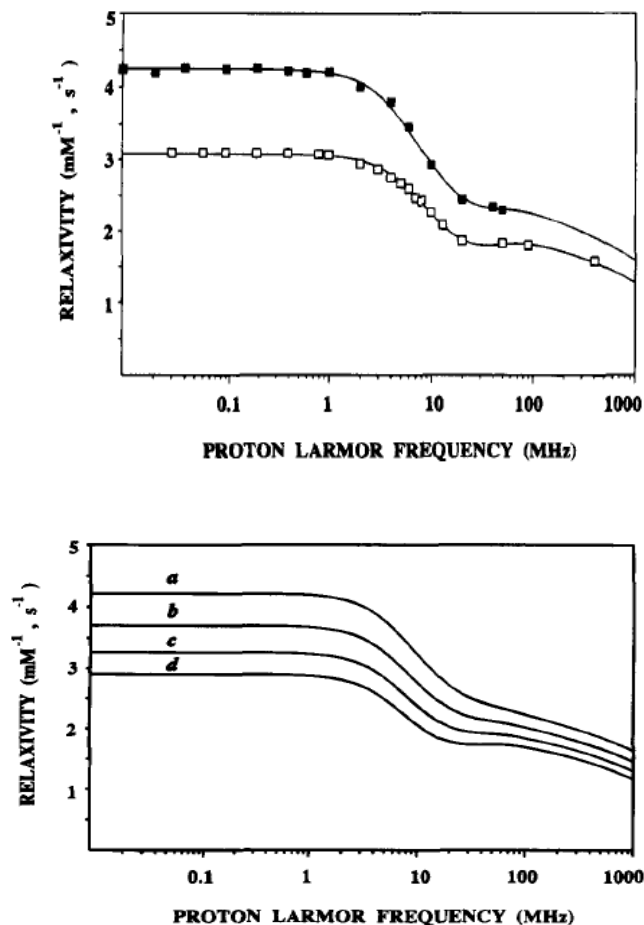


Figure 4. Simulated outer sphere NMRD profiles for values of the distance a of 3.6 (a), 3.9 (b), 4.2 (c), and 4.5 Å (d). Values typical of Gd(III) complexes were utilized for the other parameters: $\tau_{\text{SO}} = 80$ ps; $\tau_{\text{v}} = 15$ ps; $D = 2.0 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$.

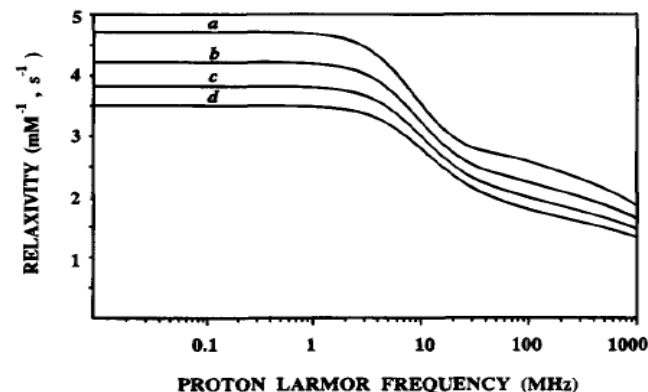
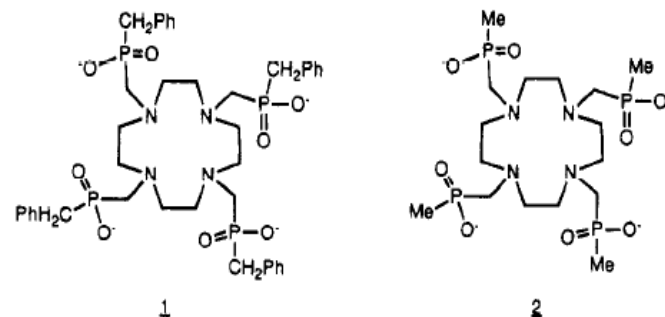


Figure 5. Simulated outer sphere NMRD profiles for values of the diffusion coefficient D of 1.7×10^{-5} (a), 2.0×10^{-5} (b), 2.3×10^{-5} (c), and $2.6 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$ (d). Values typical of Gd(III) complexes were utilized for the other parameters: $\tau_{\text{SO}} = 80$ ps; $\tau_{\text{v}} = 15$ ps; $a = 3.6$ Å.

Separation of the *inner* and *outer sphere* contributions



measurement of the T-dependency of r_{1p} at a fixed frequency

- the *outer sphere* relaxivity increases by lowering T because of the increase of D and of $T_{1,2e}$
- the T-dependence of r_{1p}^{IS} depends on the relative values of T_{1M} and τ_M :

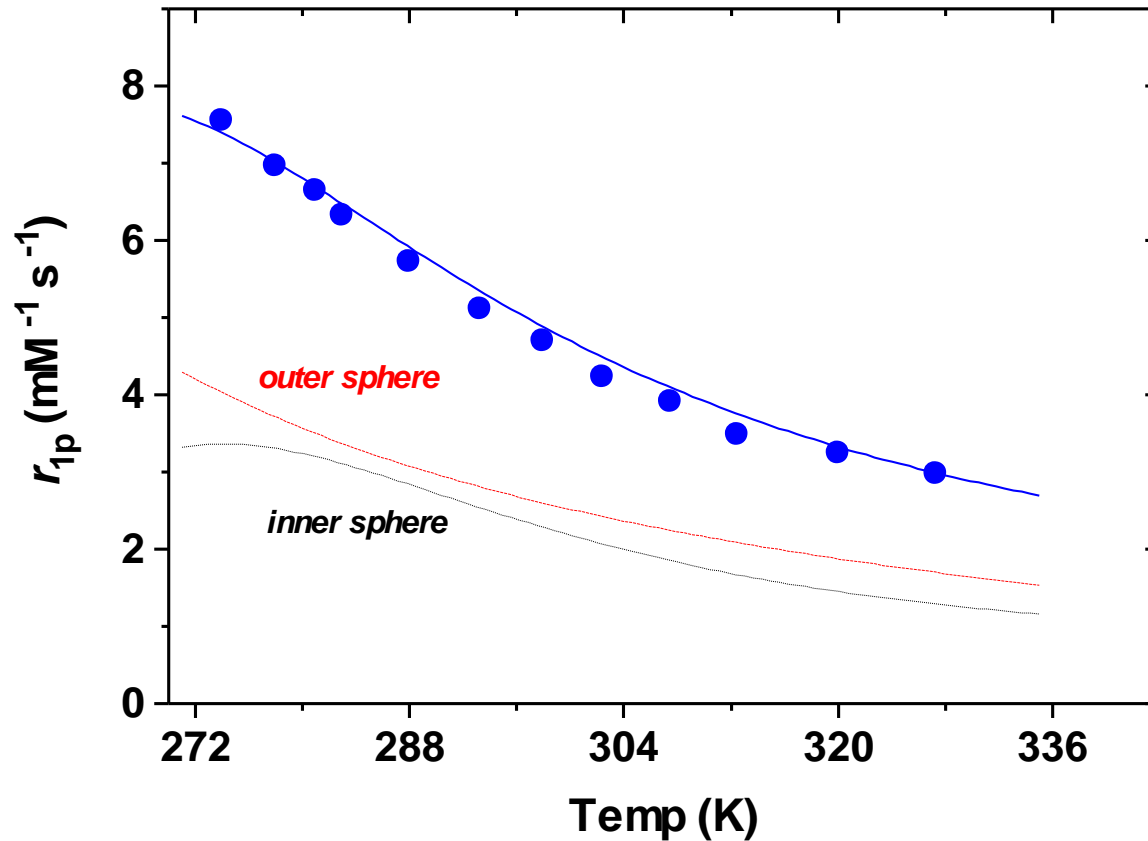
$$r_{1p}^{IS} = \frac{[CA]q}{55.6} \frac{1}{T_{1M} + \tau_M}$$



- i. $\tau_M < T_{1M}$ (fast exchange regime)**
- ii. $\tau_M \geq T_{1M}$ (slow/intermediate exchange regime)**

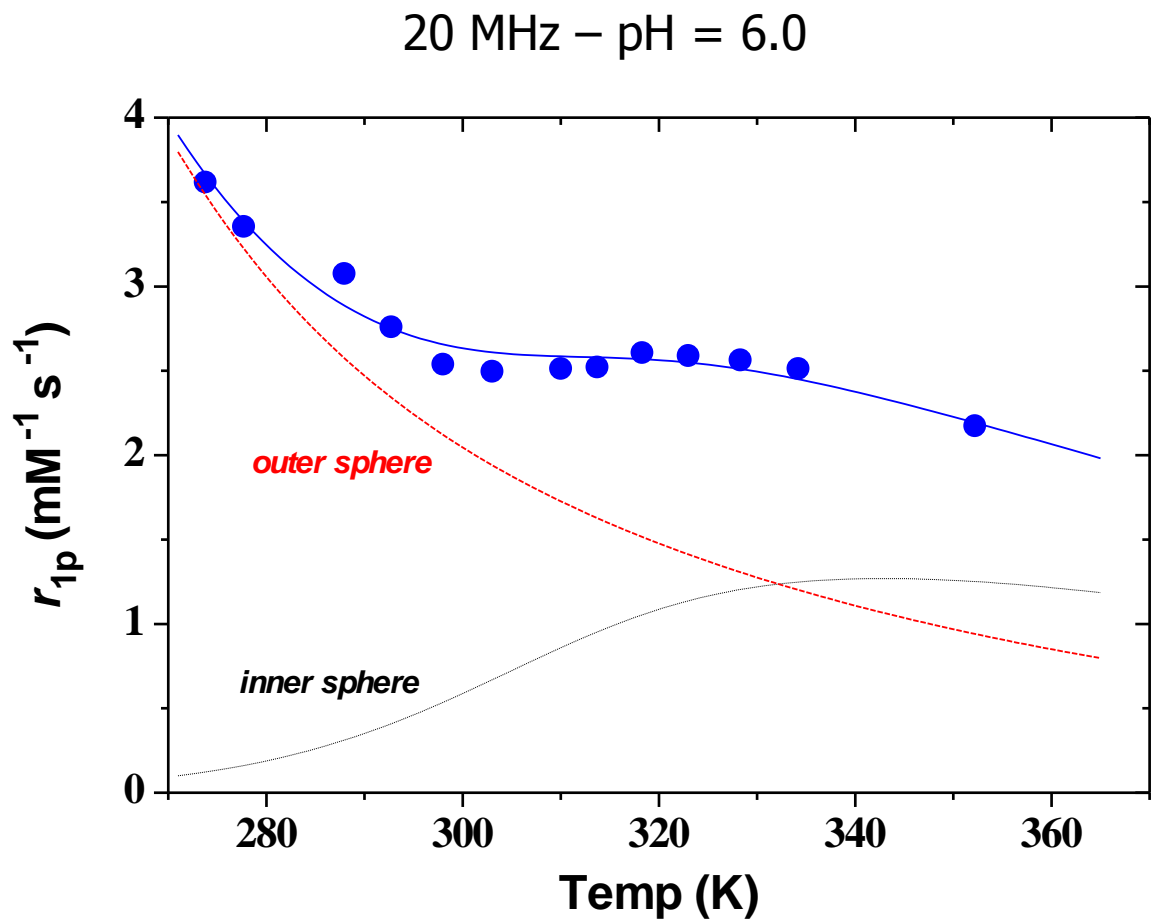
$\tau_M < T_{1M}$: fast exchange regime

20 MHz – pH = 7.1



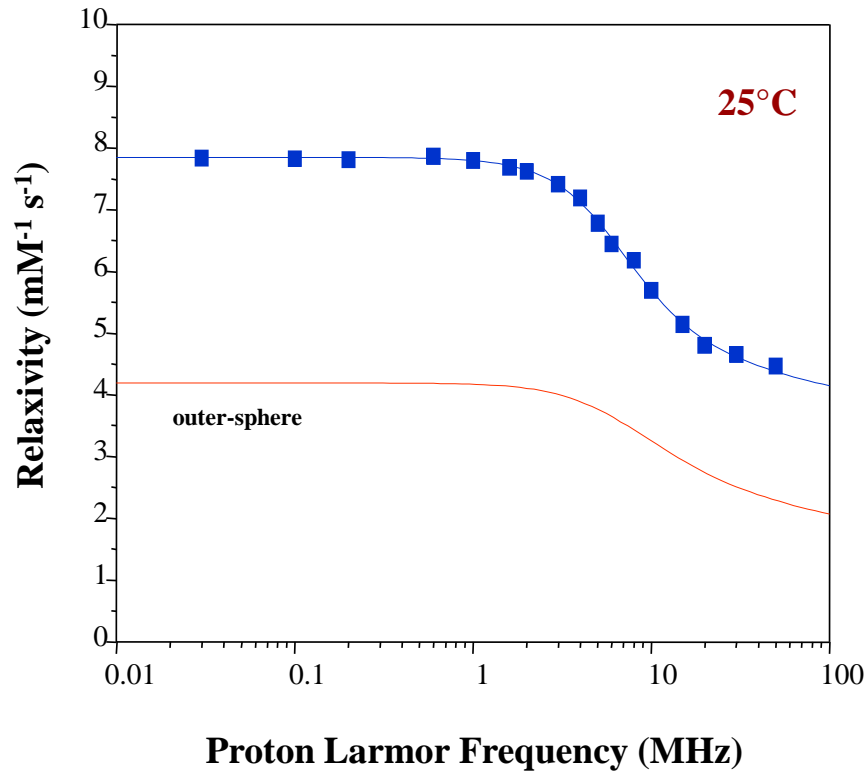
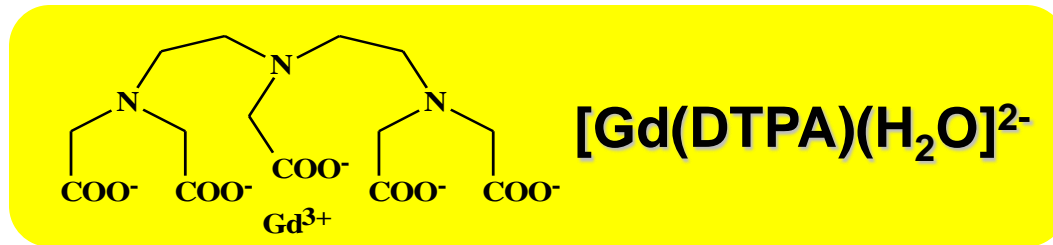
Na[Gd(DOTA)]

$\tau_M \geq T_{1M}$: slow/intermediate exchange regime



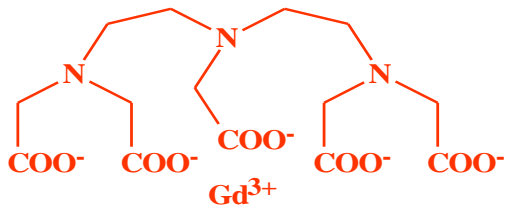
[Gd(DOTAM)]tfl3

NMRD profiles

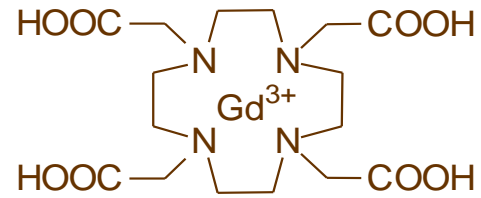


$$\begin{aligned}q &= 1 \\r &= 3 \text{ \AA} \\ \tau_R &= 70 \text{ ps} \\ \tau_M &= 300 \text{ ns} \\ 0.47 T_{1E} &= 1 \text{ ns}\end{aligned}$$

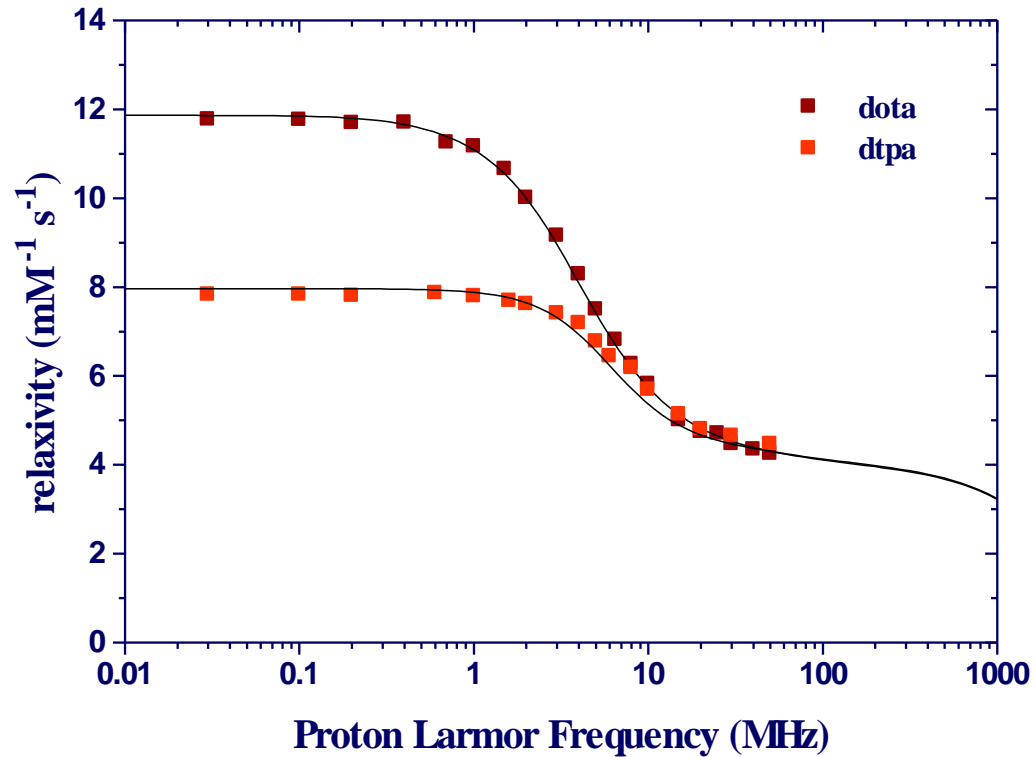
NMRD profiles



dtpa



dota



NMRD parameters

hydration number q

reorientational correlation time τ_R

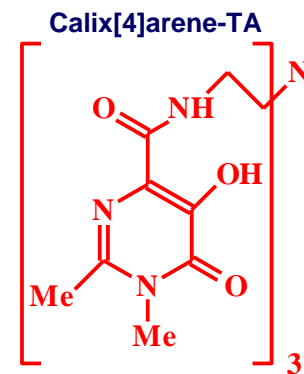
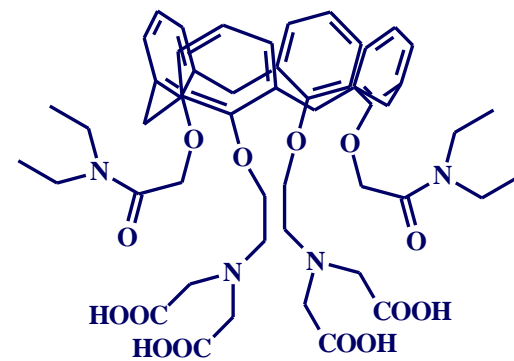
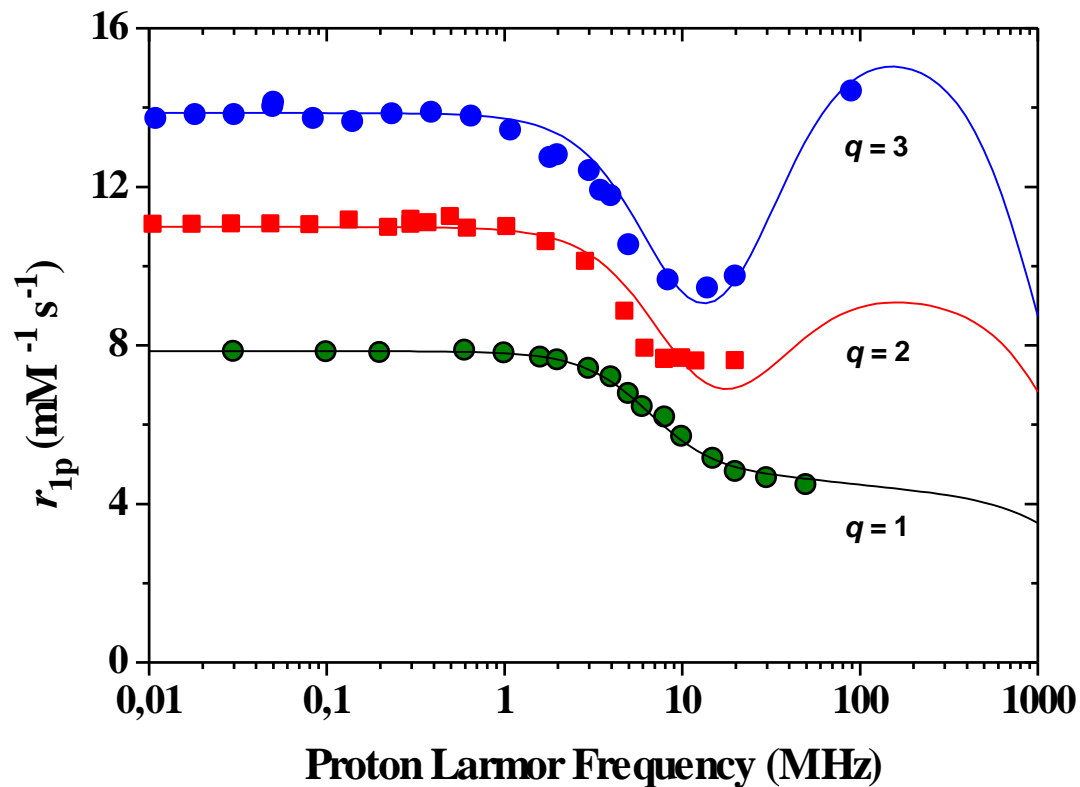
exchange lifetime τ_M

Gd-H_w distance r

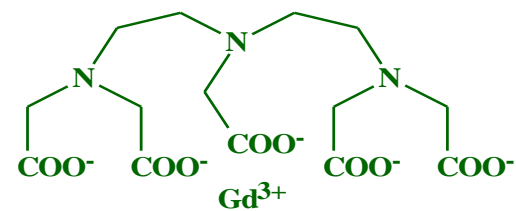
electronic relaxation times (Δ^2 ; τ_V)

a and D (outer sphere)

the effect of increasing q ...



TrenHOPY



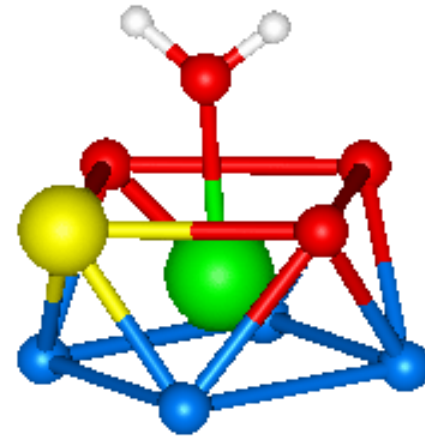
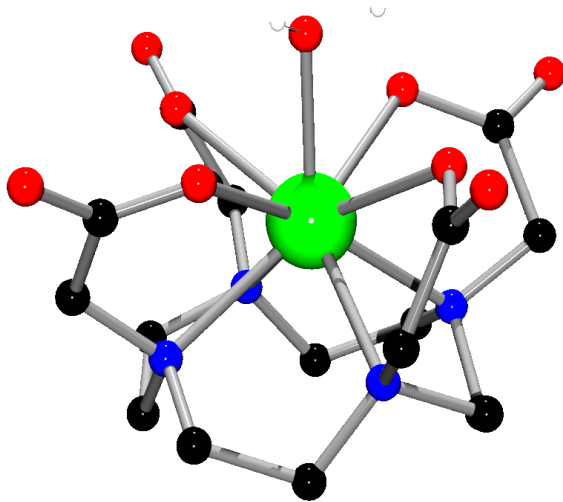
DTPA

¹H NMRD profiles (25°C) of three Gd(III) complexes differing in the number q of *inner sphere* water molecules:

GdDTPA (open circles), GdHOPY (squares), GdCalix[4]arene (filled circles)

the Gd-H_w distance r

Estimation from X-ray data? Gd-O_w ≠ Gd-H_w
Gd-H_w depends from tilt angle of the water molecule



tilt angle: variation with nature of donor groups?

pulsed ENDOR spectra: 3.1 ± 0.1 Å (Caravan et al.)

Rotational dynamics

the reorientational correlation time τ_R

A) Debye-Stokes equation

$$\tau_R = 4\pi\eta r_{\text{eff}}^3 / 3k_B T$$

B) ^{17}O NMR longitudinal relaxation time of the Gd(III)-bound water molecules

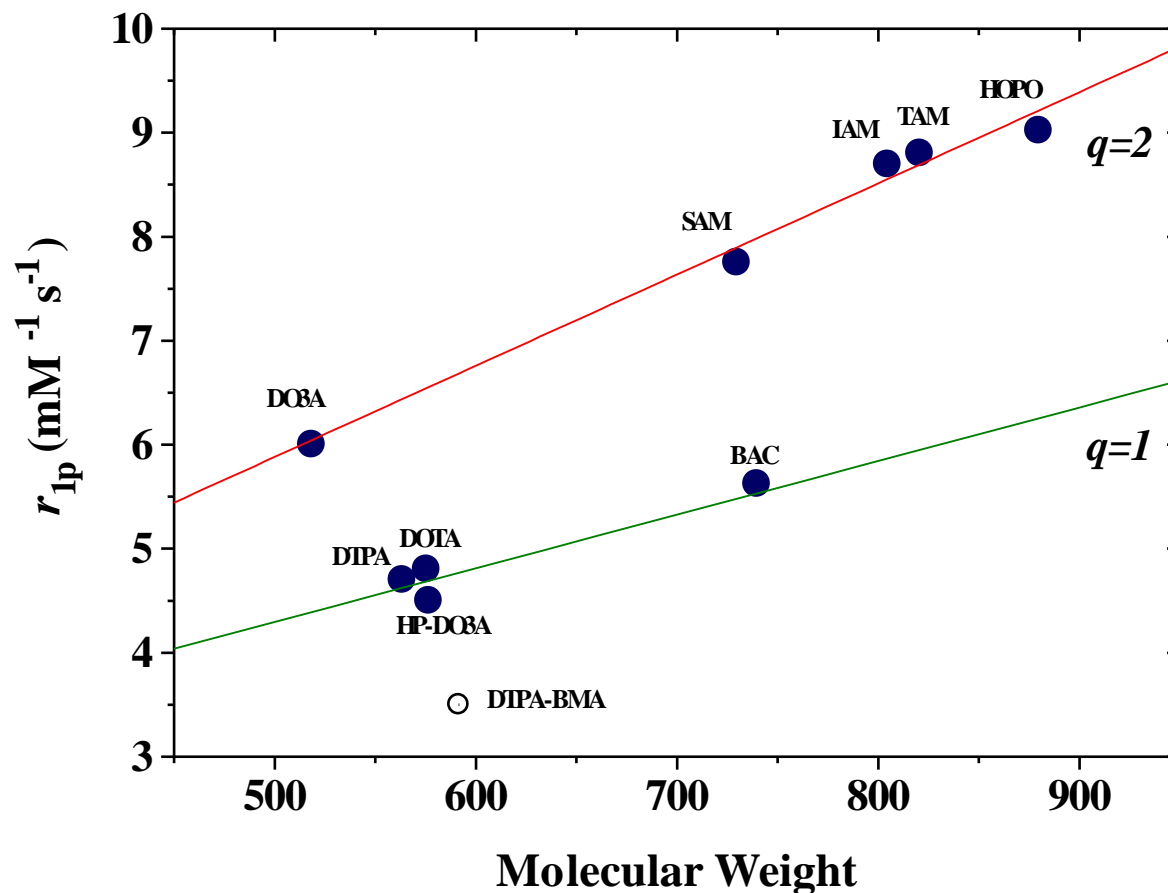
$$\frac{1}{T_{1q}} = \frac{3\pi^2}{10} \left(\frac{2I + 3}{I^2(2I - 1)} \right) \chi^2 \left(1 + \eta^2 / 3 \right) \left[0.2 \left(\frac{\tau_R}{1 + \omega_I^2 \tau_R^2} \right) + 0.8 \left(\frac{\tau_R}{1 + 4\omega_I^2 \tau_R^2} \right) \right]$$

C) ^{13}C NMR longitudinal relaxation time and NOE on a diamagnetic (Y, La, Lu) analog

$$R_1^{\text{dd}} = n \frac{4\gamma_C^2 \gamma_H^2 \hbar^2 S(S + 1)}{3r_{\text{C-H}}^6} \tau_R$$

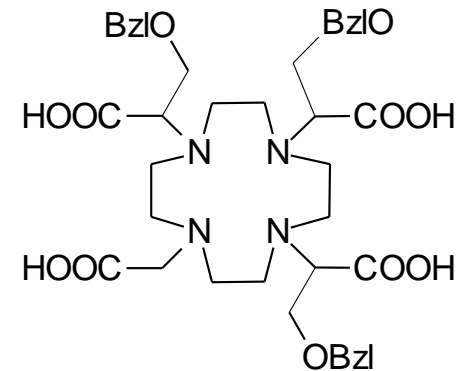
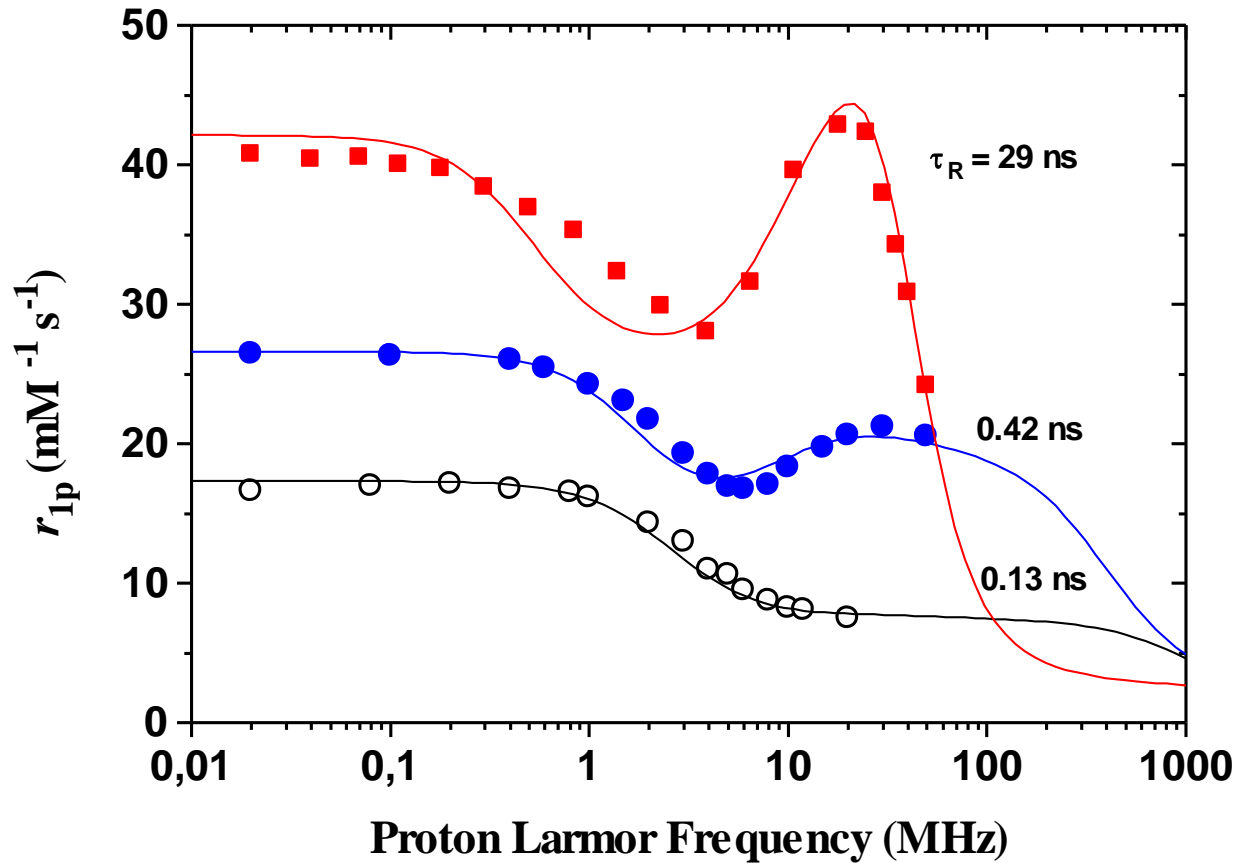
$$R_1^{\text{obs}} / R_1^{\text{dd}} = \eta_{\text{max}} / \eta$$

the reorientational correlation time τ_R



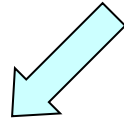
The linear relationship confirms the dependence of r_{1p} on the tumbling rate of the complexes, the close similarity of the outer sphere contributions, and excludes significant variations in the distance values $r_{\text{Gd-H}}$

the effect of slowing down rotation...

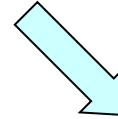


^1H NMRD profiles (25°C) of GdDOTA-BOM₃ (open circles), its inclusion complex with β -cyclodextrin (filled circles) and of GdDOTA-BOM₃-HSA adduct (squares). The different shapes and amplitudes of the profiles are primarily due to the different rotational correlation times of the paramagnetic complexes.

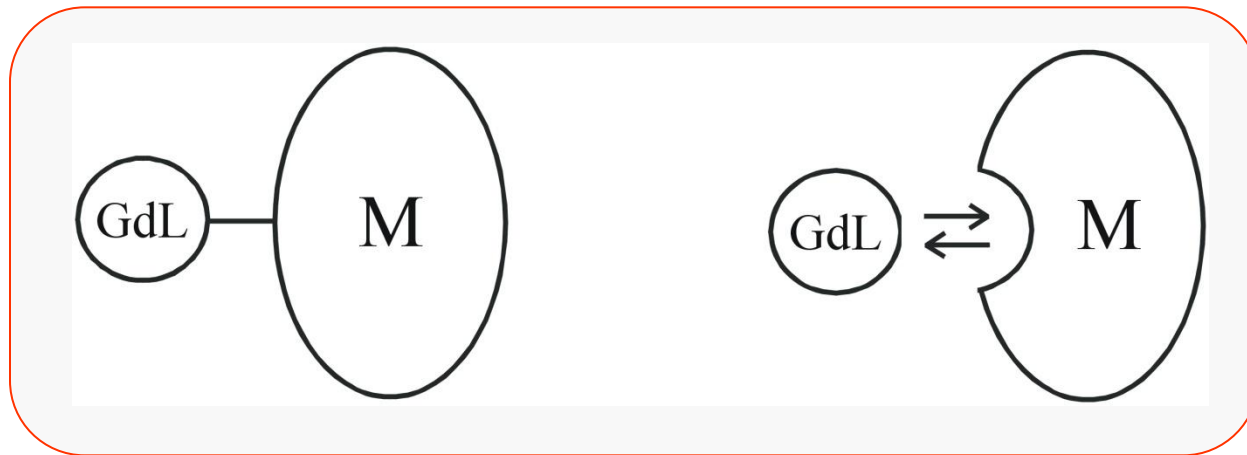
How to increase τ_R ?



Covalent binding



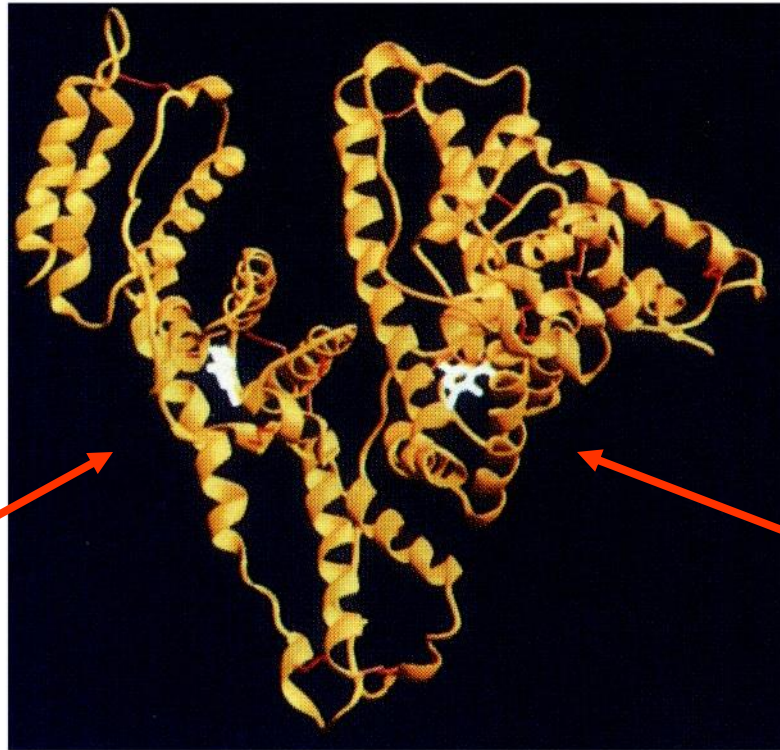
Non Covalent binding



M = macromolecule

(proteins, synthetic or natural polymers, dendrimers,...)

HSA as target molecule for Gd^{III}-based CAs



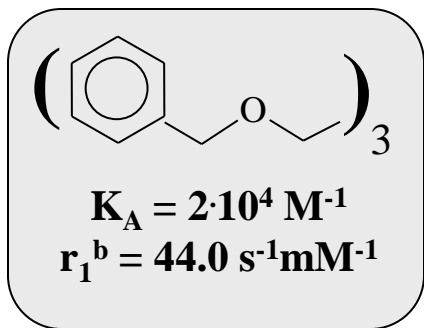
IIIA
binding site

IIA
binding site

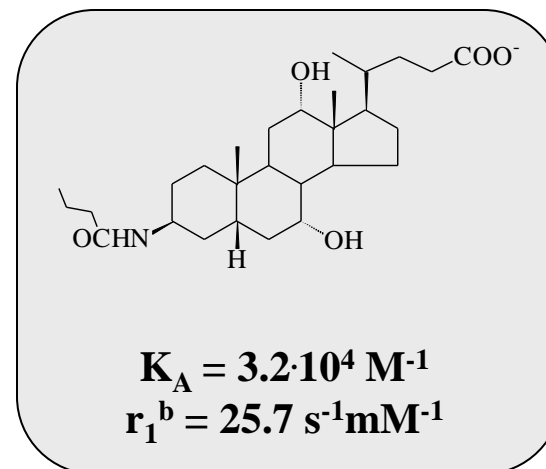
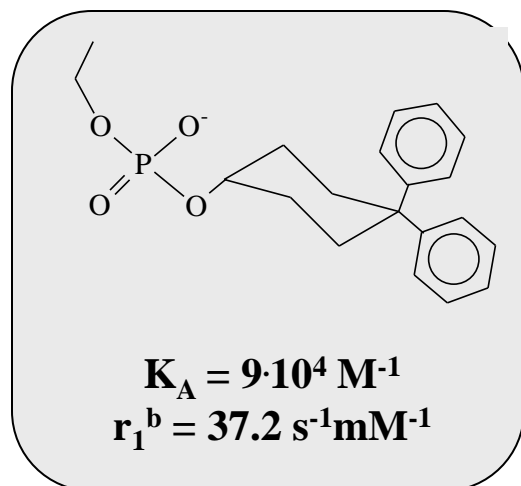
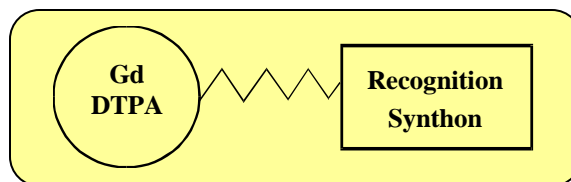
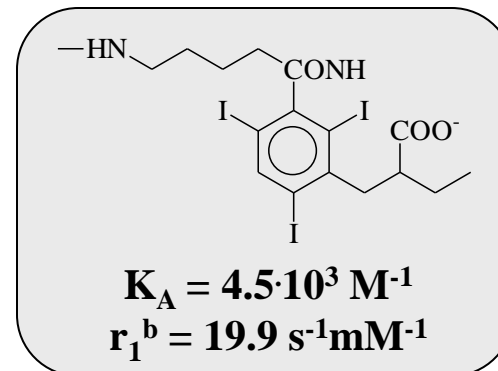


Development of CAs for MR Angiography (MRA)

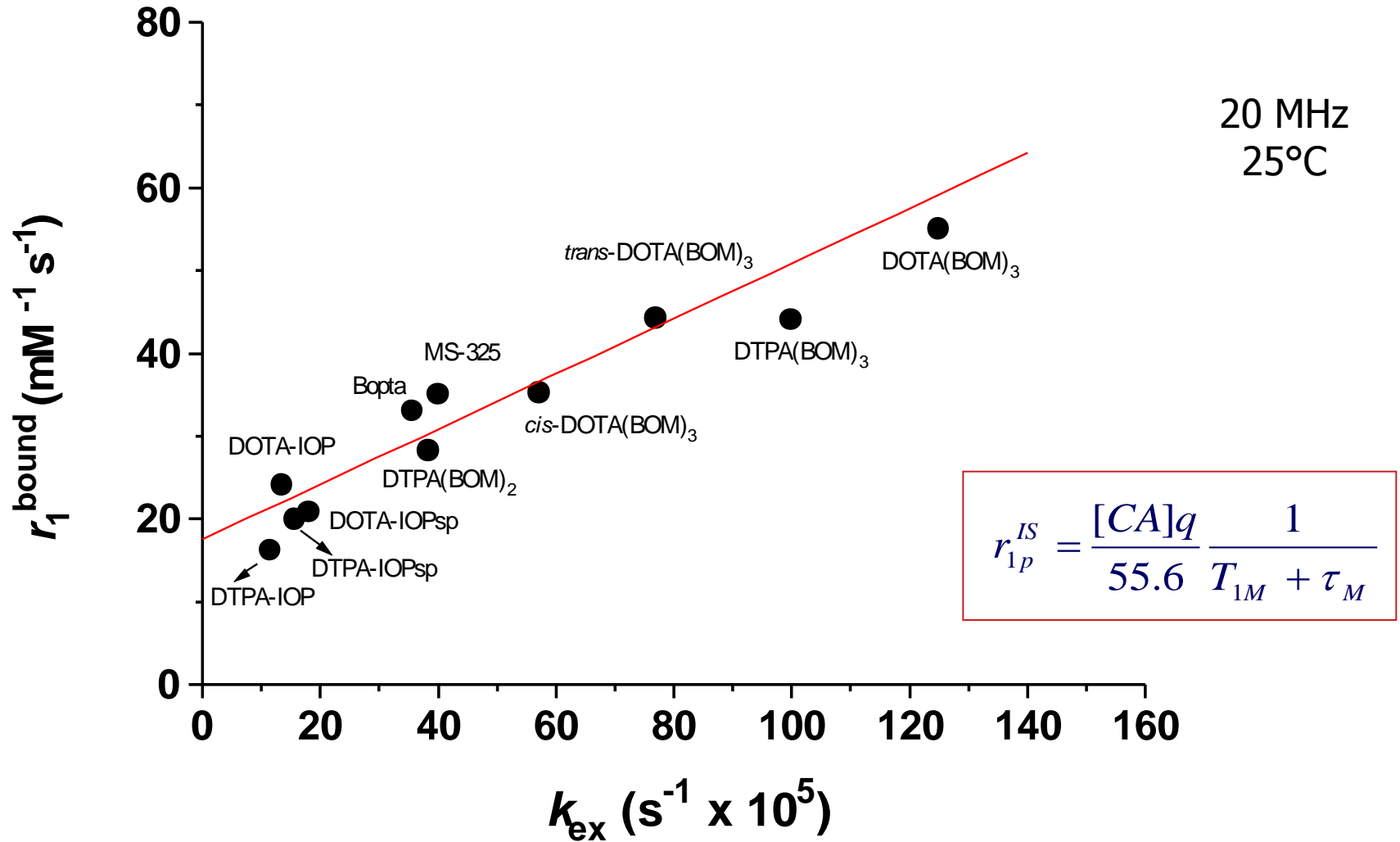
Non covalent binding to HSA



25°C - 0.47 T

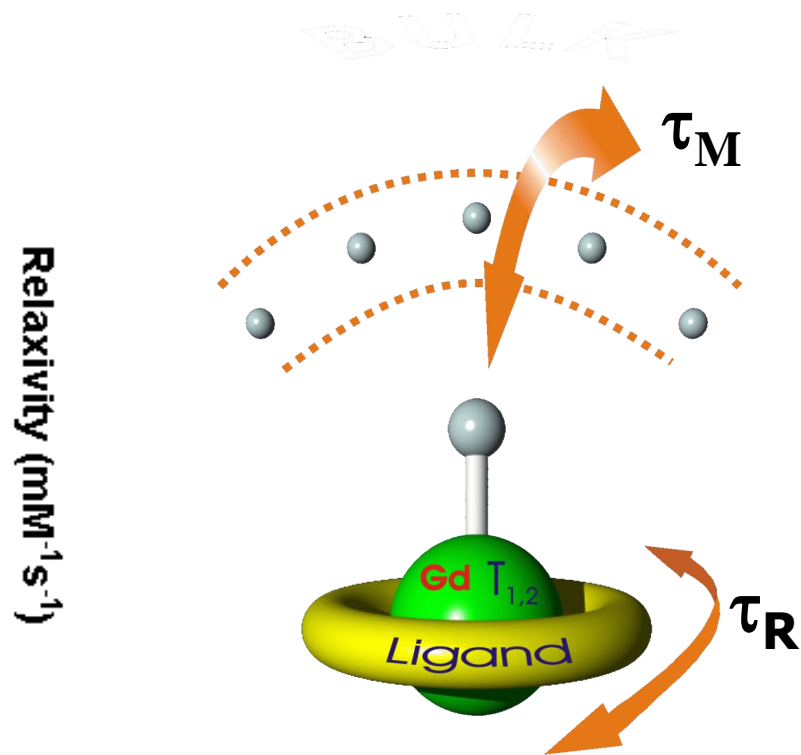
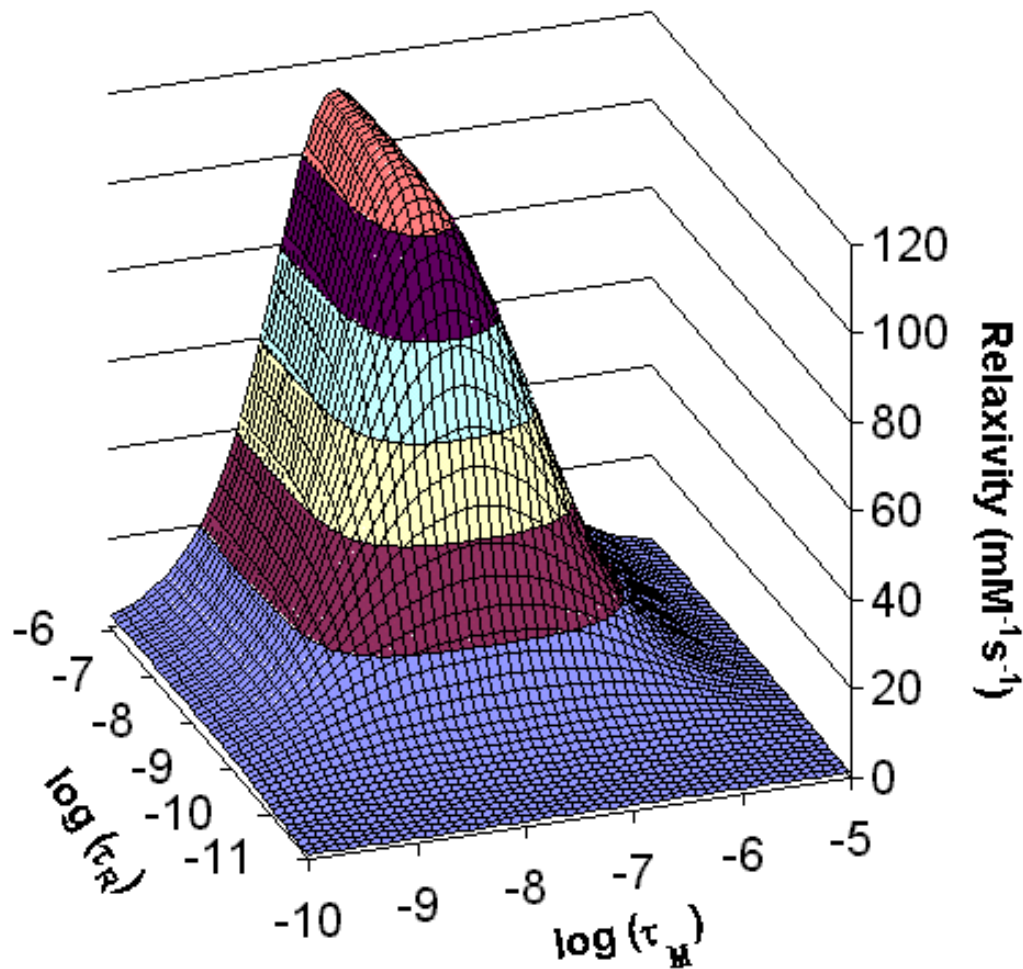


Non-covalent HSA conjugates



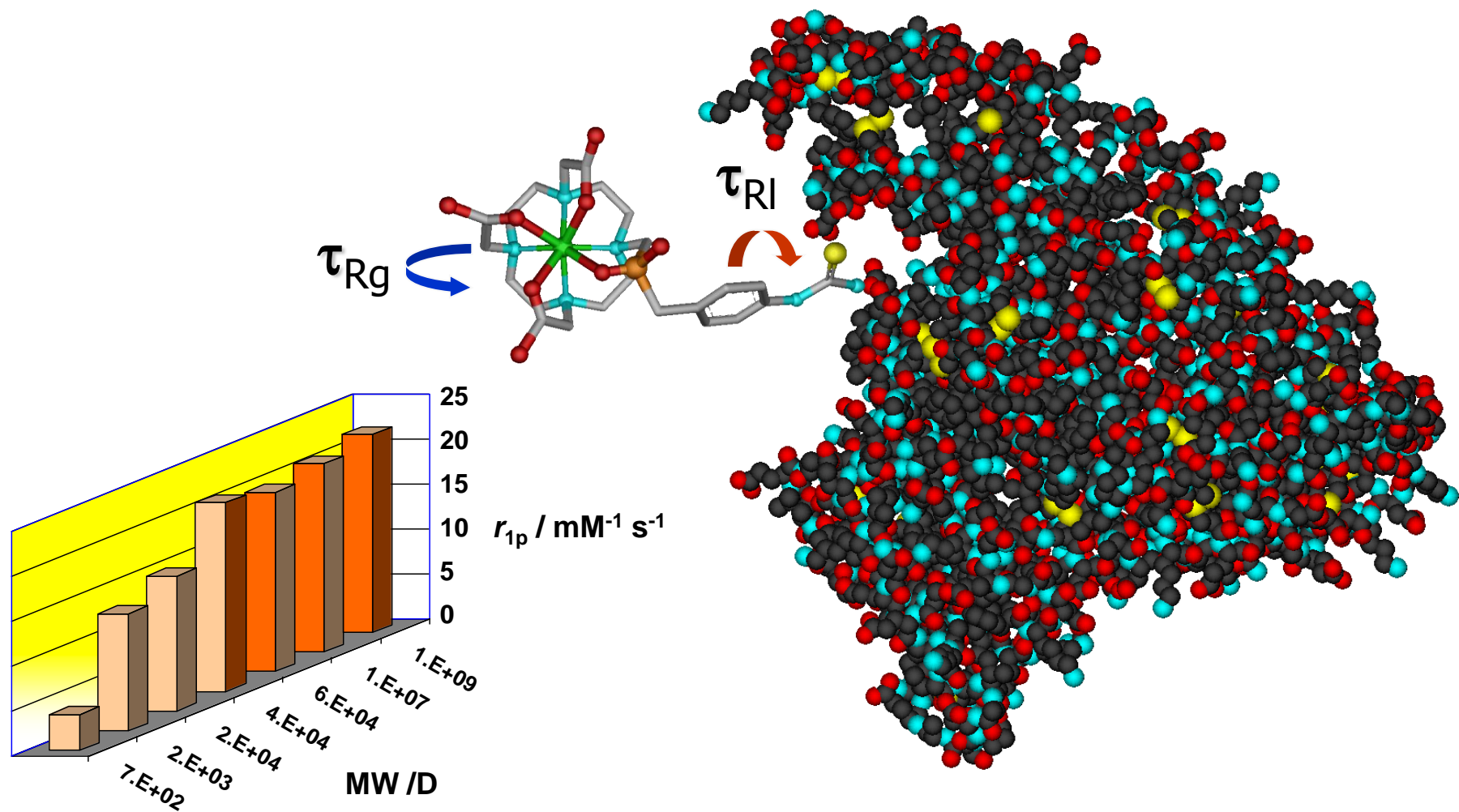
Aime, S.; Botta, M.; Fasano, M.; Terreno, E., *Protein-Bound Metal Chelates*, in *The Chemistry of Contrast Agents in Medical Magnetic Resonance Imaging*, (A. E. Merbach, É Tóth, Eds.), Wiley: New York, 2001

the limiting factors: rotation and exchange...



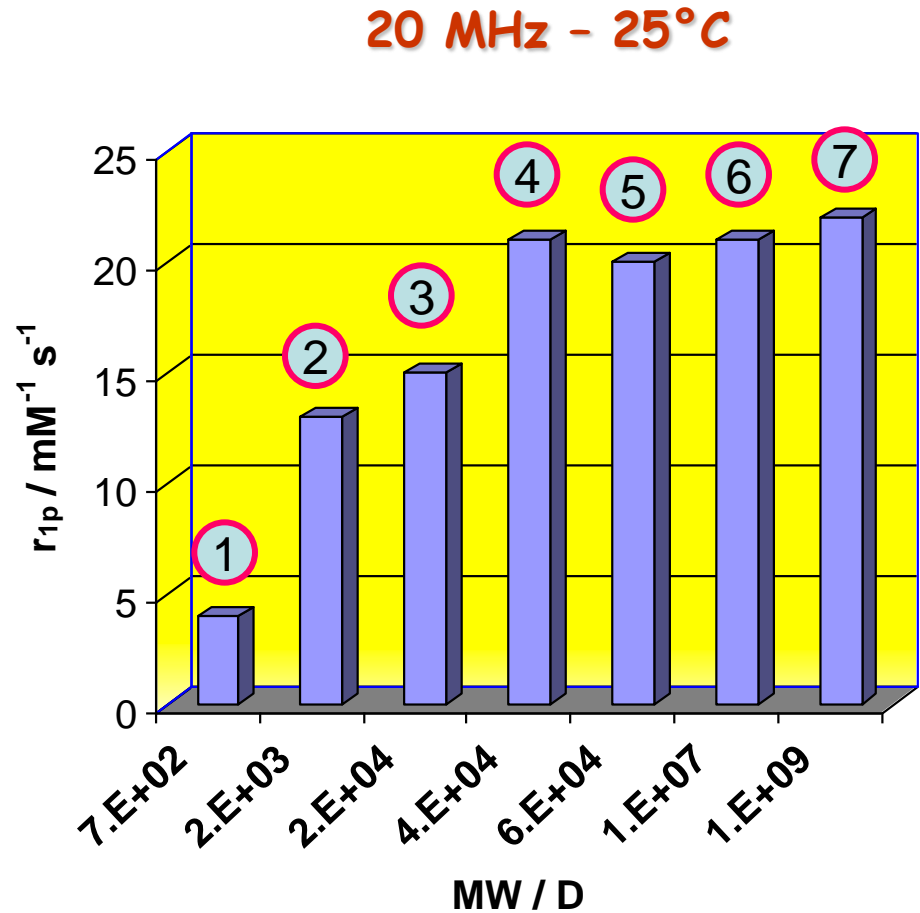
The problem of internal rotation

another limiting factor



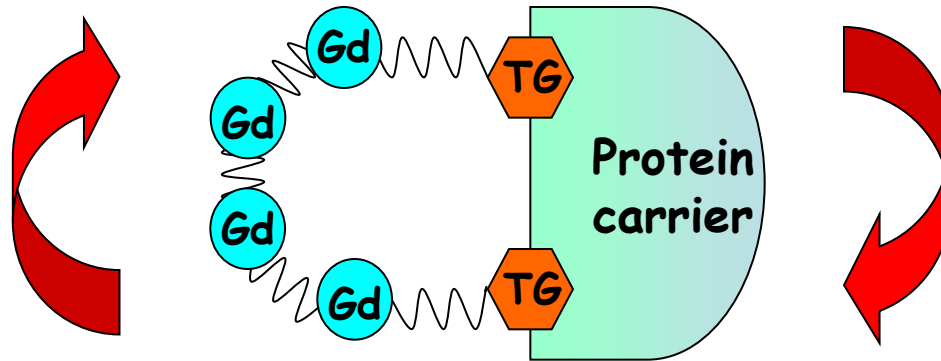
Relaxivity of macromolecular contrast agents

- ① Small sized compounds
- ② Fullerenes
- ③ Small dendrimers (ex. Gadomer-17)
- ④ Protein carriers
- ⑤ Dendrimers (ex. PAMAM-G5)
- ⑥ Micelles
- ⑦ Liposomes



The problem of internal rotation

Approaches to solve the problem (1)



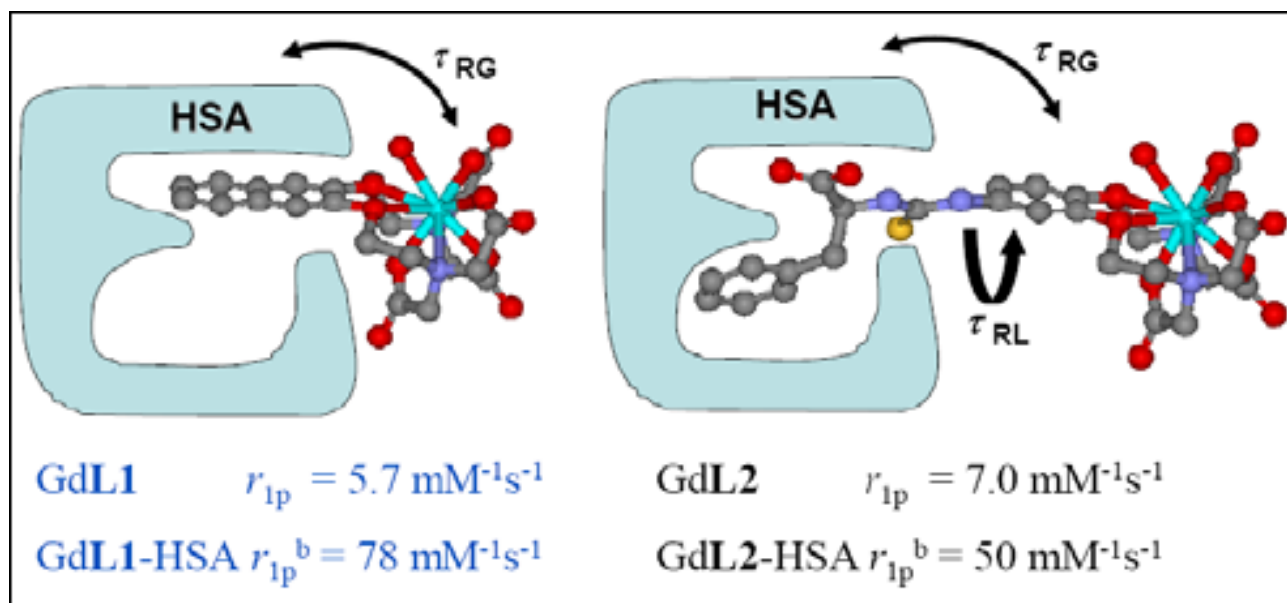
**GdDTPA tetramer
non targeted
 r_{1p} per Gd = 24**



**GdDTPA tetramer
twice targeted to HSA
 r_{1p}^b per Gd = 41**

HSA Conjugates

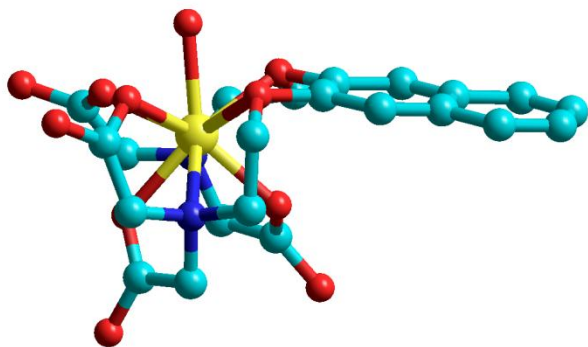
Approaches to solve the problem (2)



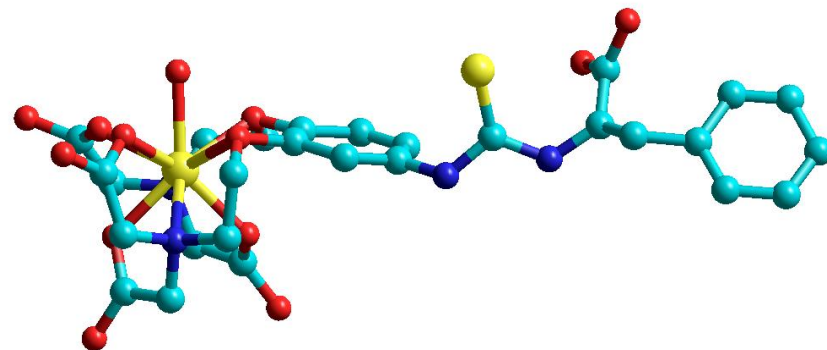
Maximizing the relaxivity of HSA-bound gadolinium complexes
by simultaneous optimization of rotation and water exchange

Chem. Commun., **2007**, 4044-4046

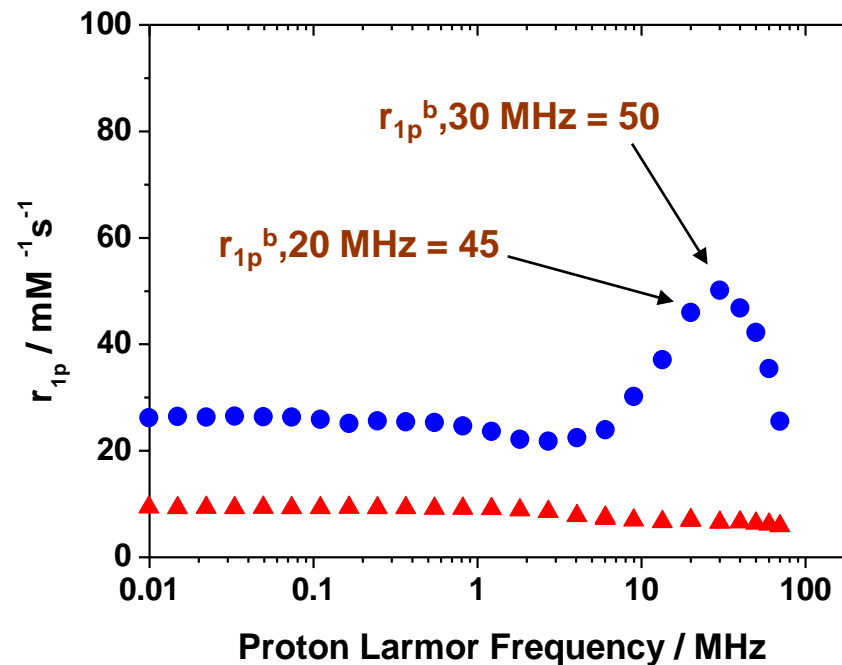
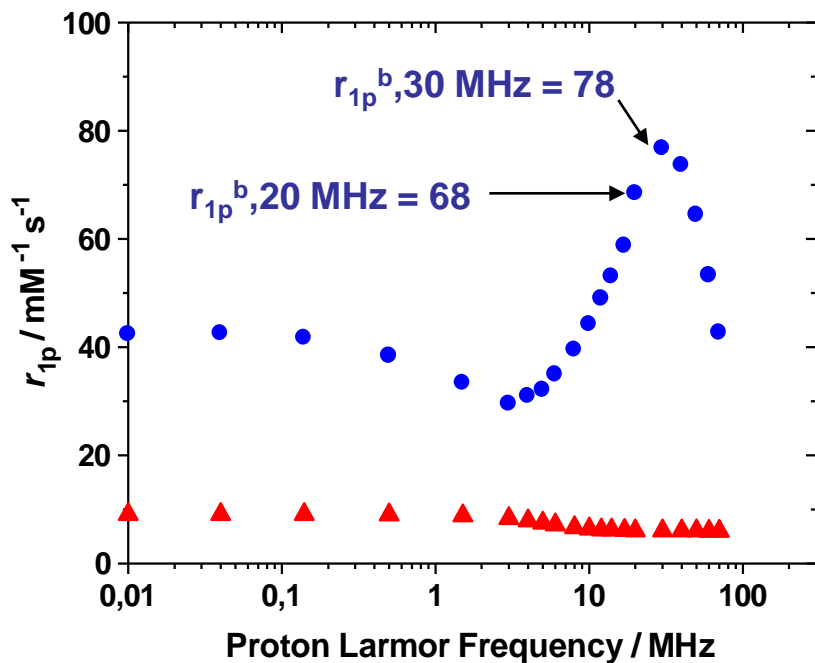
Binding to HSA: ^1H NMRD



GdL1



GdL2

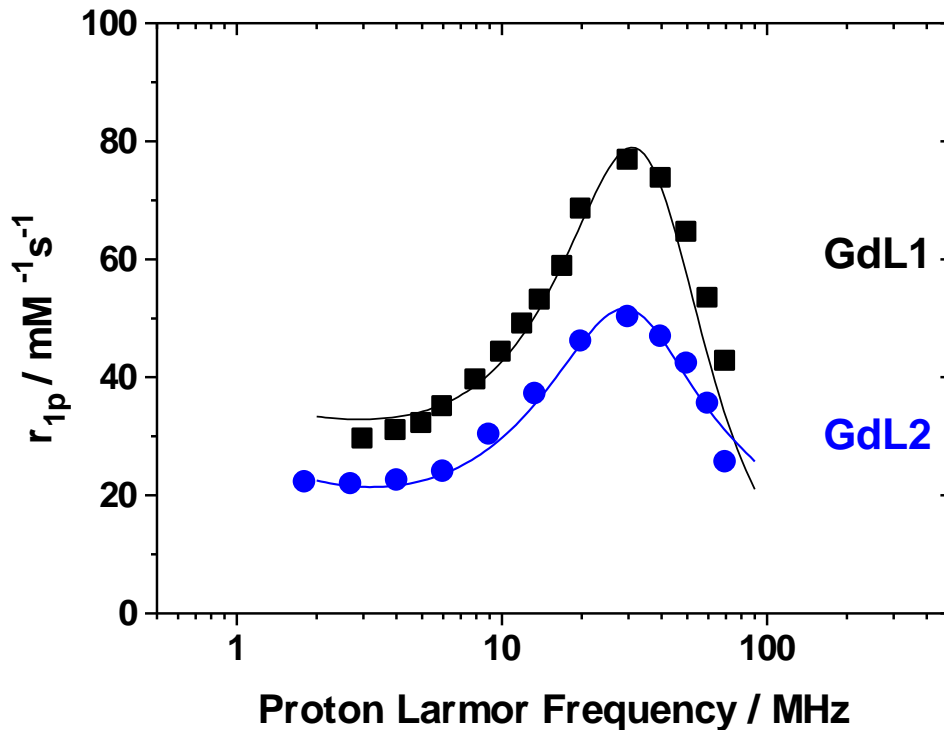


$$r_{lp}^{IS} = \frac{q}{55.6} \times \frac{1}{T_{IM} + \tau_M}$$

Lipari-Szabo model free approach:

$$\frac{1}{T_{IM}} = \frac{2}{15} \frac{\gamma_{HG}^2 S(S+1) \beta^2}{r_{Gd-H}^6} \left[\frac{3S^2 \tau_{Clg}}{1 + \omega_H^2 \tau_{Clg}^2} + \frac{3(1-S^2) \tau_{Cl}}{1 + \omega_S^2 \tau_{Cl}^2} \right]$$

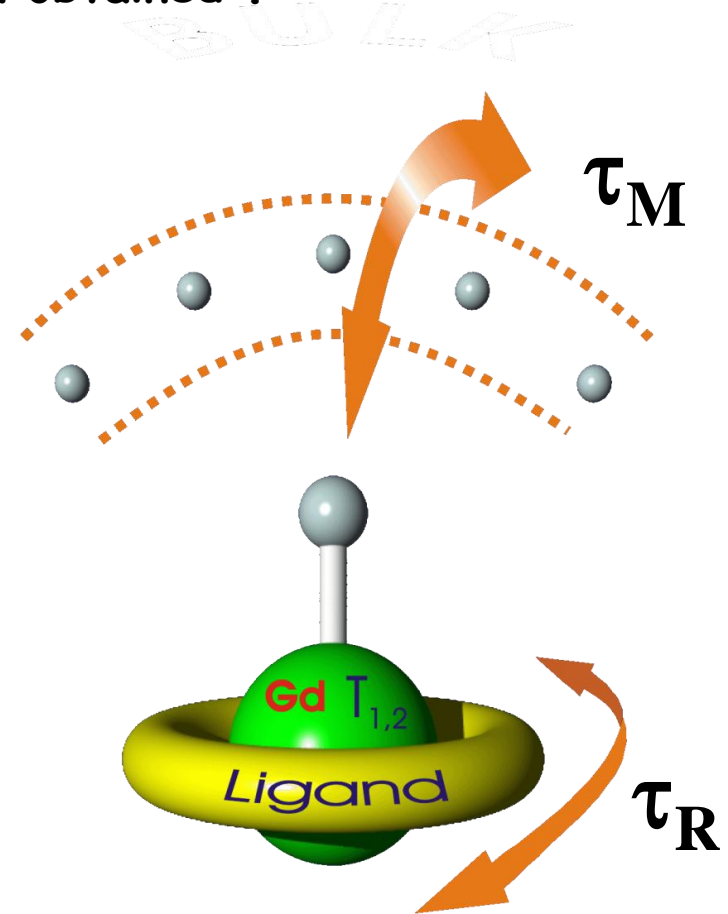
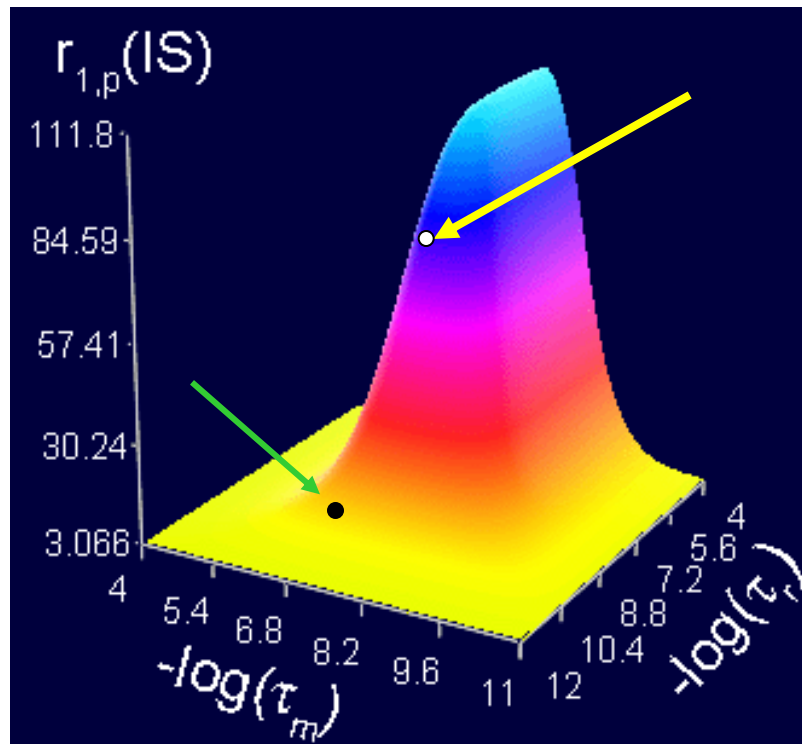
$$\frac{1}{\tau_{Cl}} = \frac{1}{\tau_M} + \frac{1}{\tau} + \frac{1}{T_{le}} \quad \Rightarrow \quad \frac{1}{\tau} = \frac{1}{\tau_{Rg}} + \frac{1}{\tau_{Rl}}$$



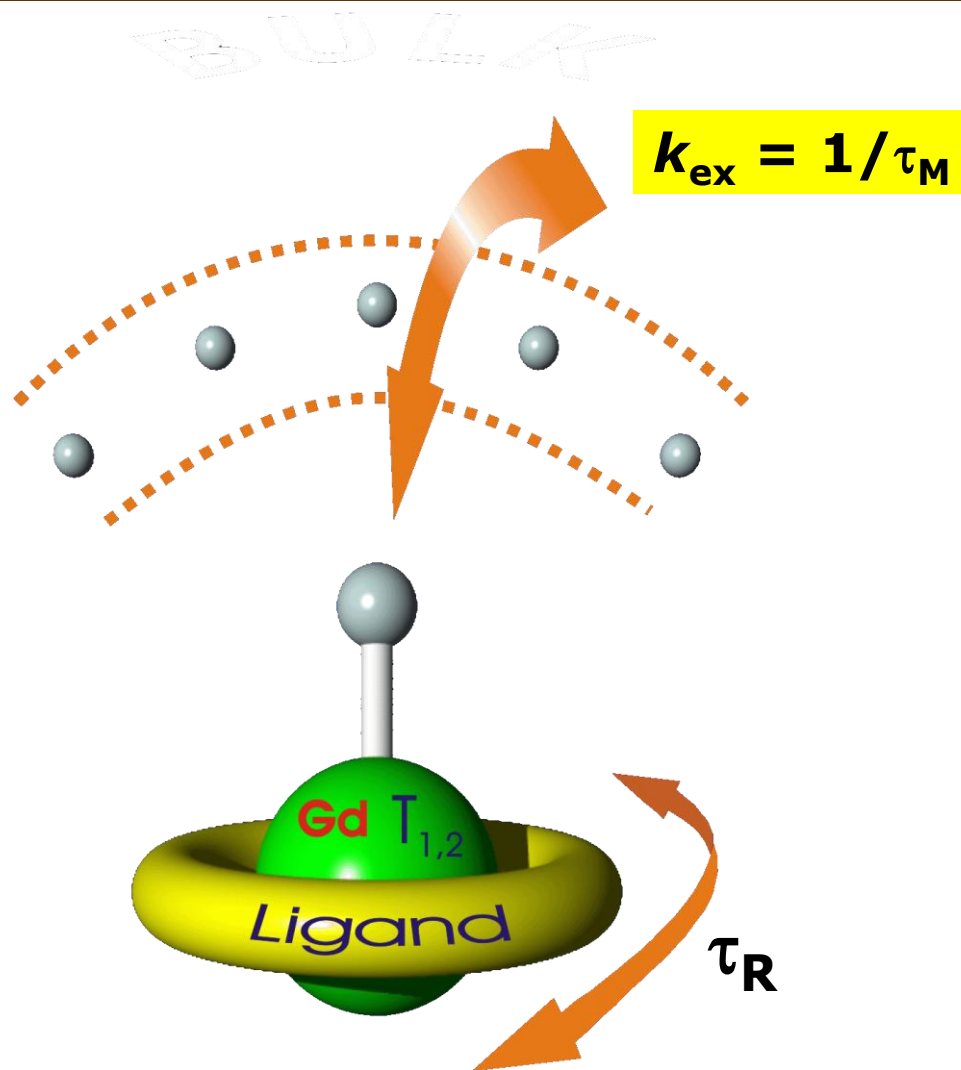
	GdL1	GdL2
$\tau_{Rg}(ns)$	20	20
$\tau_{Rl}(ns)$	9.0	1.1
S^2	0.64	0.31

Conclusions

By optimizing both the rotational dynamics and the rate of water exchange for the first time the high relaxivity value predicted by the theory has been obtained .

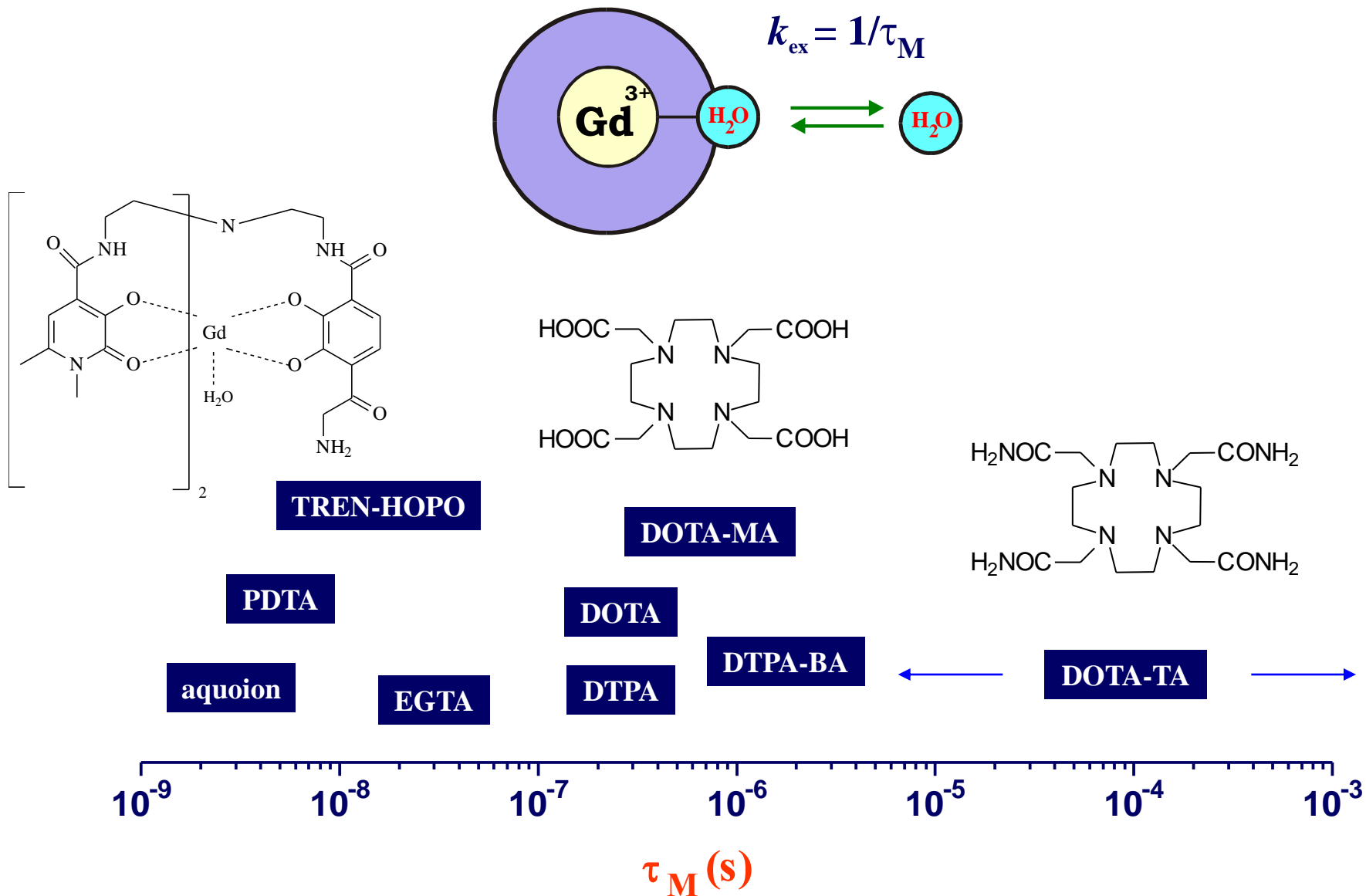


the limiting factors: rotation and exchange...

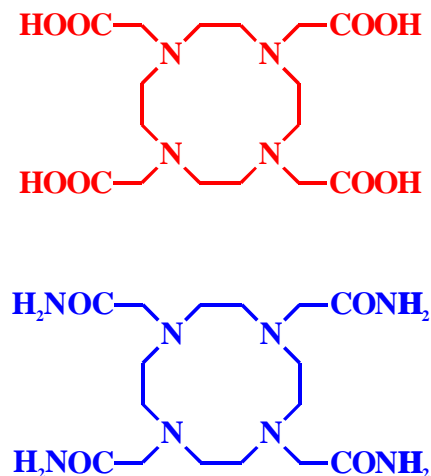
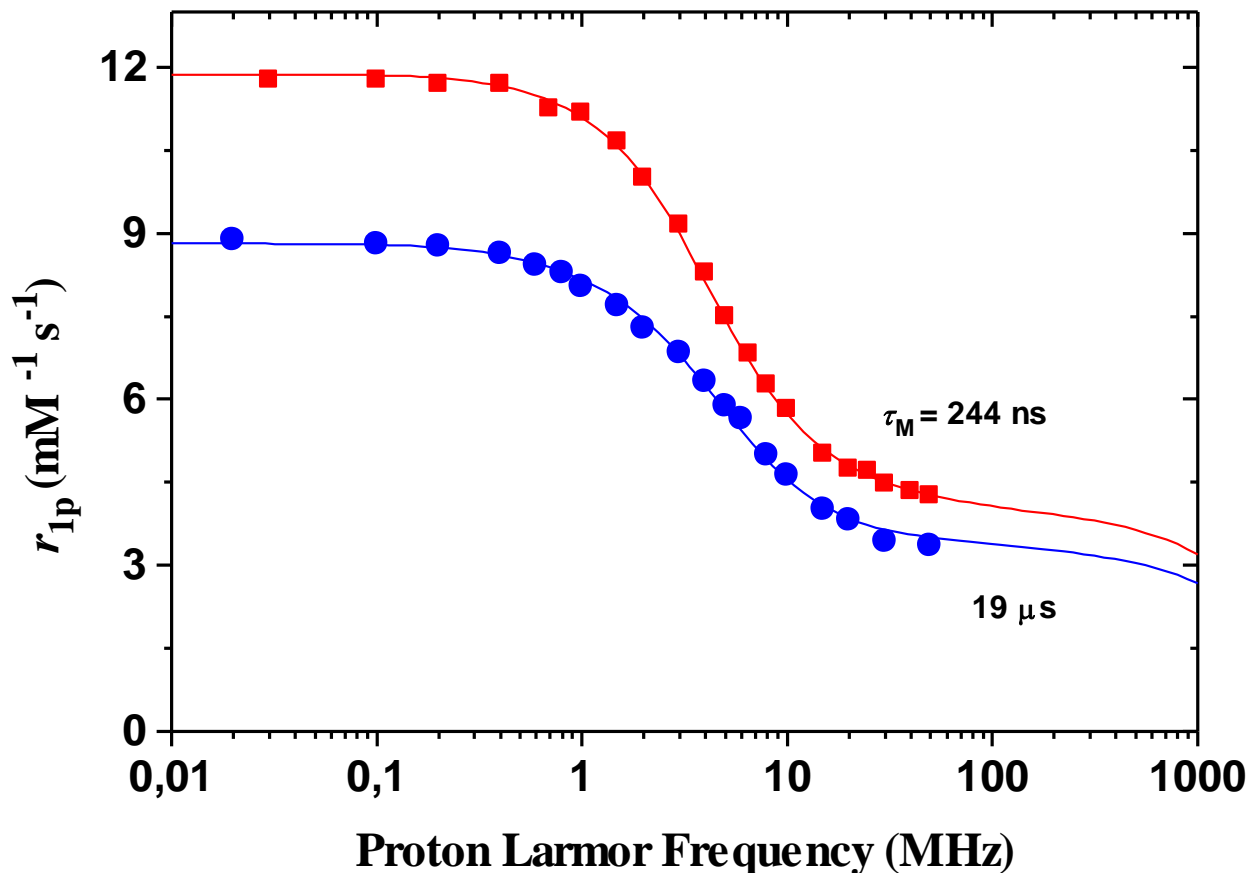


water exchange rate

the reorientational exchange lifetime τ_M



the effect of slowing down water exchange...

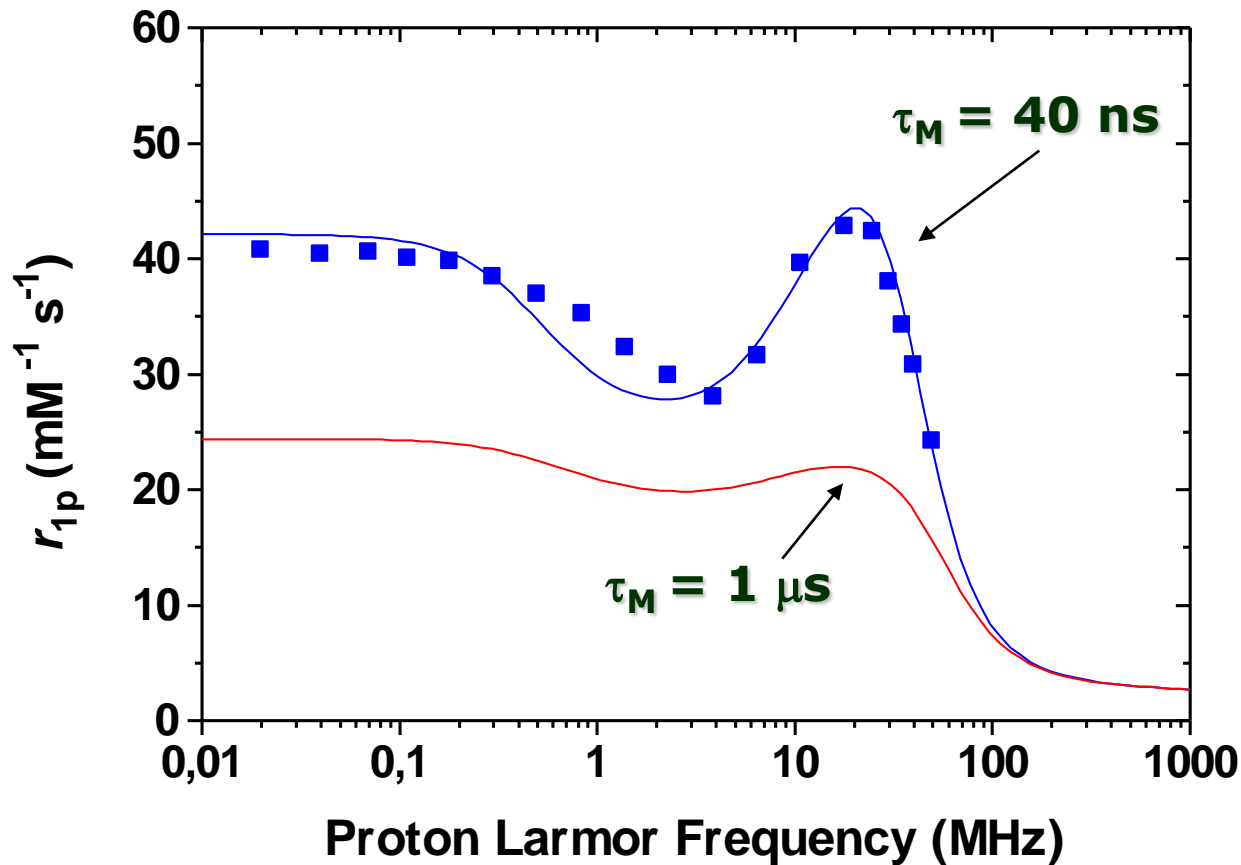


$$r_{1p}^{IS} = \frac{[\text{CA}]q}{55.6} \frac{1}{T_{1M} + \tau_M}$$

¹H NMRD profile (25°C) of $[\text{Gd}(\text{DOTA})]^-$ (squares) and $[\text{Gd}(\text{DOTAM})]^{3+}$ (circles) showing the effect on relaxivity due to a slow rate of water exchange

the effect of slowing down water exchange...

...on a macromolecular system: HSA-bound Gd complex



$$r_{1p}^{IS} = \frac{[CA]q}{55.6} \frac{1}{T_{1M} + \tau_M}$$

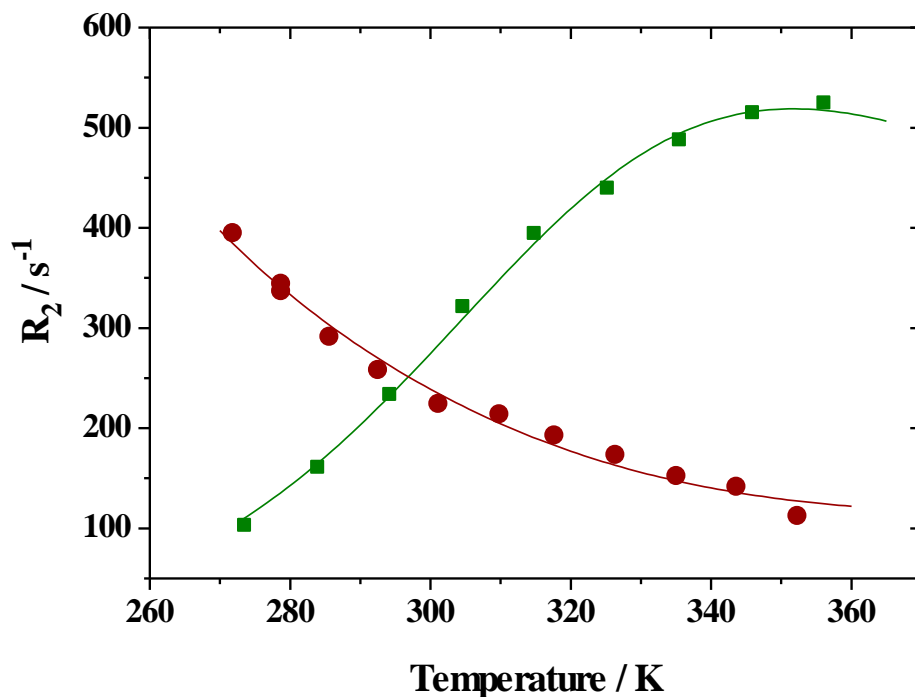
Techniques: ^{17}O NMR

$$R_{2p}^{\text{O}} = \frac{[C]q}{55.6} (\tau_{\text{M}}^{\text{O}})^{-1} \frac{R_{2M}^{\text{O}^2} + (\tau_{\text{M}}^{\text{O}})^{-1} R_{2M}^{\text{O}} + \Delta\omega_{\text{M}}^{\text{O}^2}}{\left(R_{2M}^{\text{O}} + (\tau_{\text{M}}^{\text{O}})^{-1}\right)^2 + \Delta\omega_{\text{M}}^{\text{O}^2}}$$

Swift-Connick Equations

$$R_{2M}^{\text{O}} = \frac{1}{3} \left(\frac{A}{\hbar}\right)^2 S(S+1) \left(\tau_{\text{E1}} + \frac{\tau_{\text{E2}}}{1 + \omega_s^2 \tau_{\text{E2}}^2} \right)$$

$$\tau_{\text{Ei}}^{-1} = \tau_{\text{M}}^{\text{O}^{-1}} + T_{\text{iE}}^{-1}$$

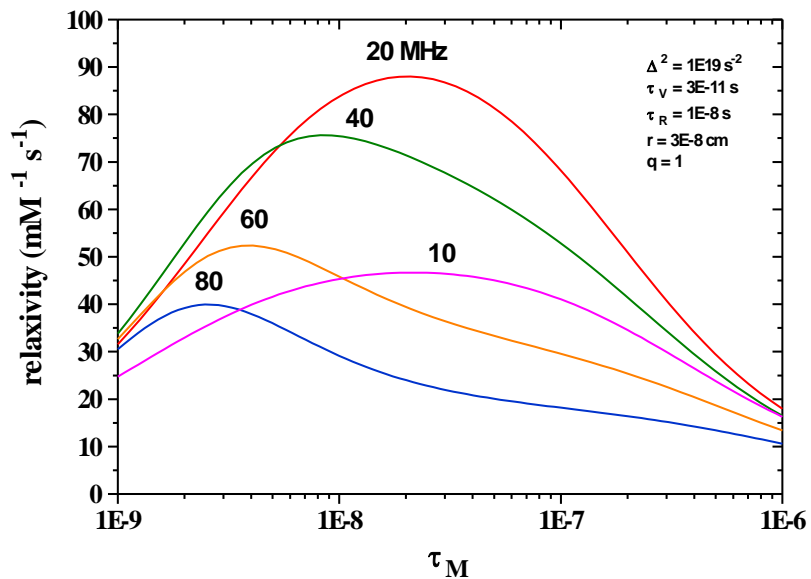


■ slow exchange

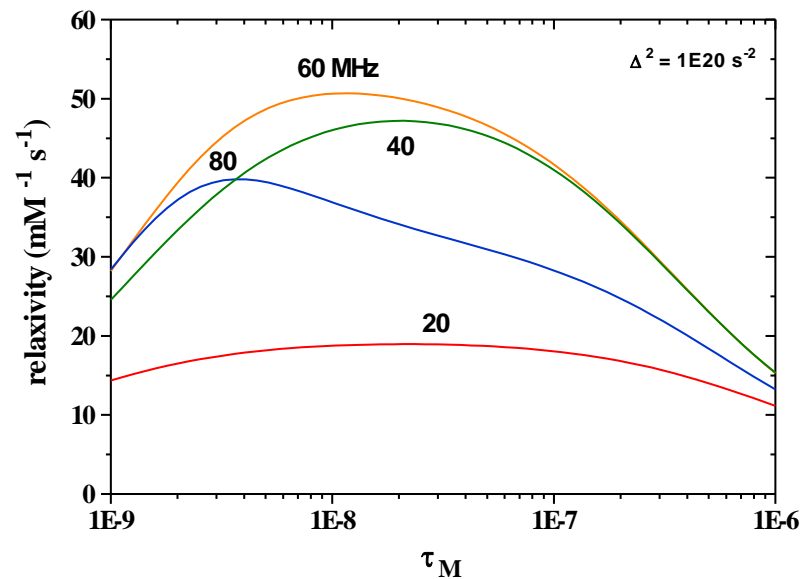
● fast exchange

the reorientational exchange lifetime τ_M

⇒ the optimal τ_M value depends on B_0




DOTA-like complexes



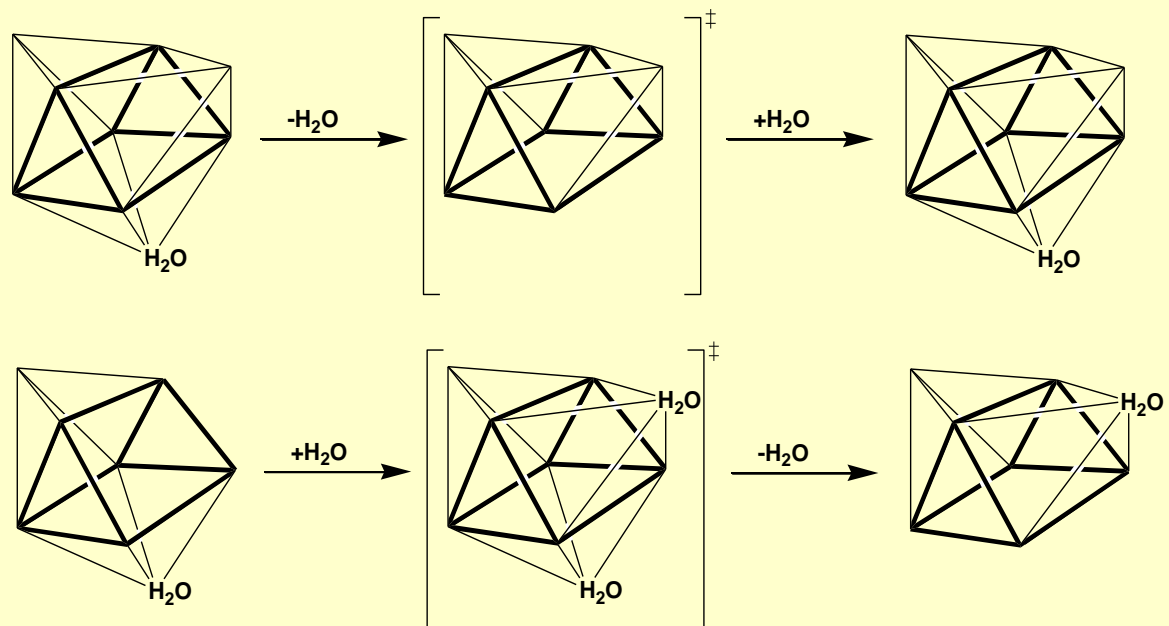
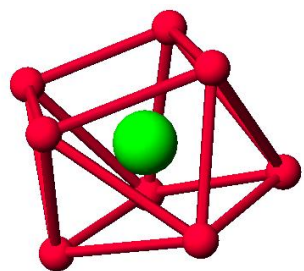
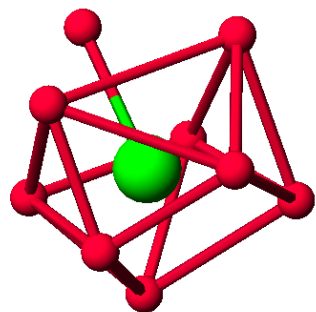
DTPA-like complexes

The Factors That Control Water Exchange

- **8- or 9-coordinate ground state (mechanism of the exchange process)**
- **charge of the complex**
- **molecular geometry** 
- **steric interactions at the H₂O binding site**
- **electronic effects**
- **(assistance of 2nd sphere H₂O molecules and nature of the counterion)**

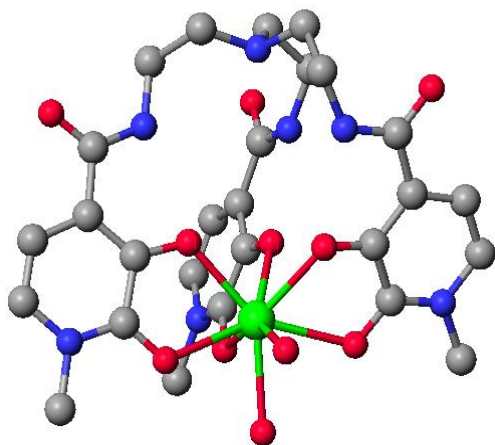
k_{ex} and mechanism of the exchange process

water exchange mechanisms

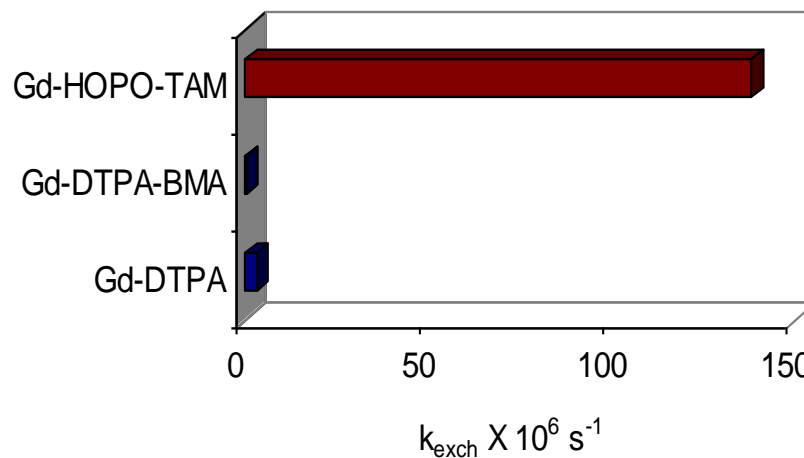
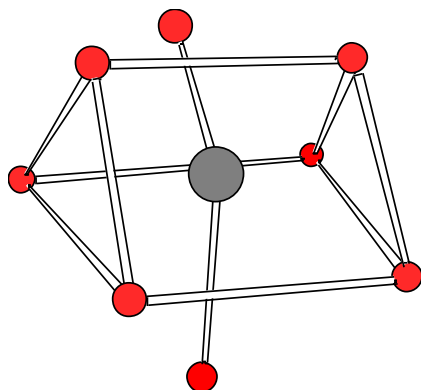
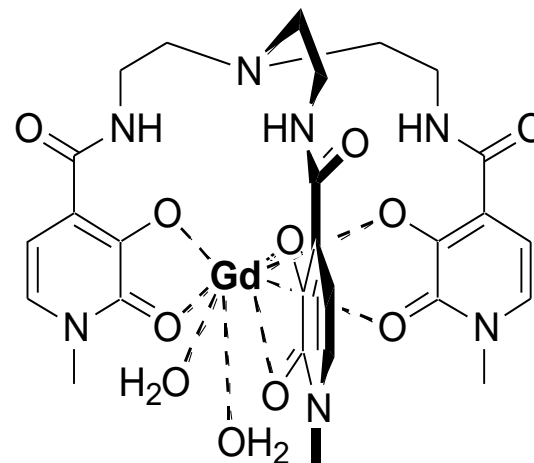


Dissociative (top) and associative (bottom) mechanisms of water exchange at an idealized tricapped trigonal prismatic Gd^{3+} metal center. The dissociative mechanism operates through a coordinatively unsaturated 8-coordinate intermediate, while in the associative mechanism a 9-coordinate intermediate is formed.

k_{ex} and mechanism of the exchange process



Gd(TREN-trisHOPO)(H₂O)₂
X-Ray structure

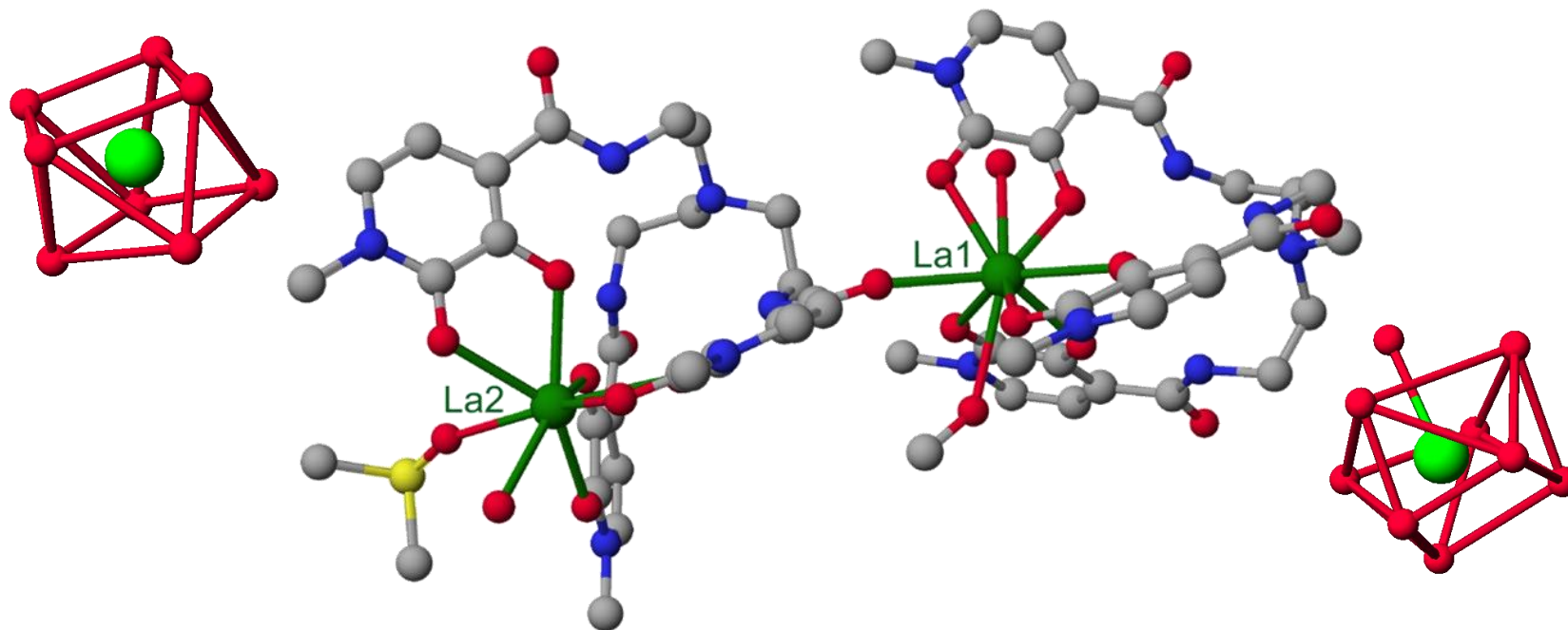


Xu, J.; Franklin, S. J.; Whisenhunt, D. W.; Raymond, K. N. *J. Am. Chem. Soc.* **1995**, *117*, 7245.

Cohen, S. M.; Xu, J.; Radkov, E.; Raymond, K. N.; Botta, M.; Barge, A.; Aime, S. *Inorg. Chem.* **2000**, *39*, 5747.

k_{ex} and mechanism of the exchange process

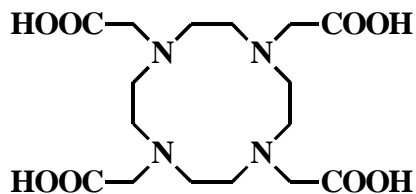
Crystal structure of La-TREN-1-Me-3,2-HOPO
dimer of an 8 and 9 coordinate complex



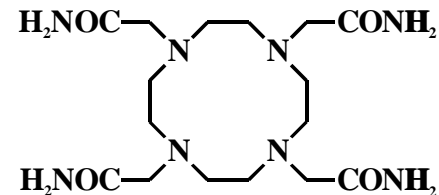
Molecule assembles as a dimer, **La1: 9-coordinate; La2: 8-coordinate**
La1: monocapped square antiprismatic geometry; La2: square antiprismatic geometry
Water molecules replaced by DMSO and methanol in the solid state

Inorg. Chem. **2000**, 39, 5747.
J. Am. Chem. Soc. **2003**, 125, 14274.

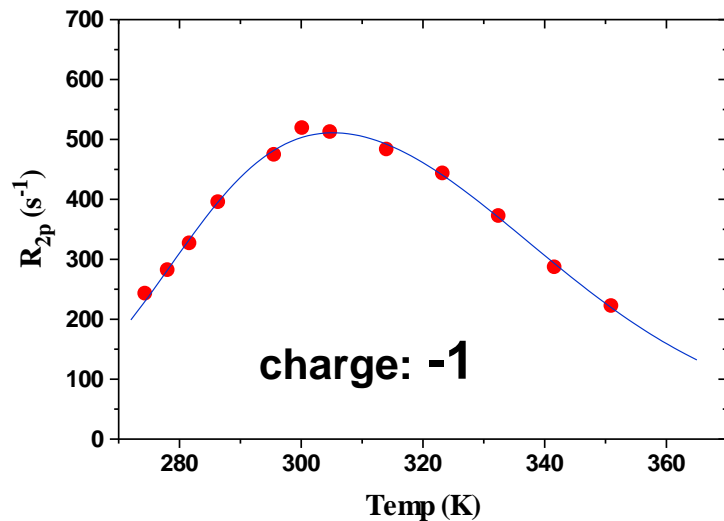
k_{ex} and charge of the complex



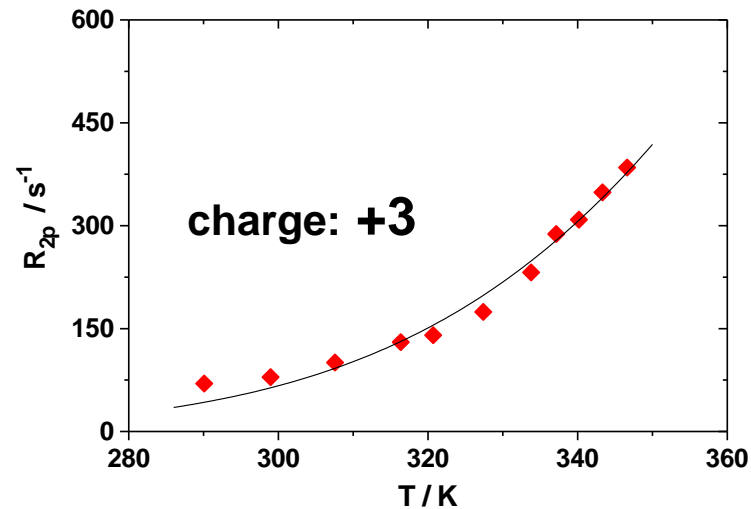
DOTA



DOTAM



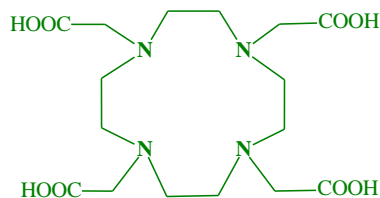
$$\tau_M = 300 \text{ ns}$$



$$\tau_M = 14 \mu s$$

k_{ex} and molecular geometry

DOTA



...and derivatives

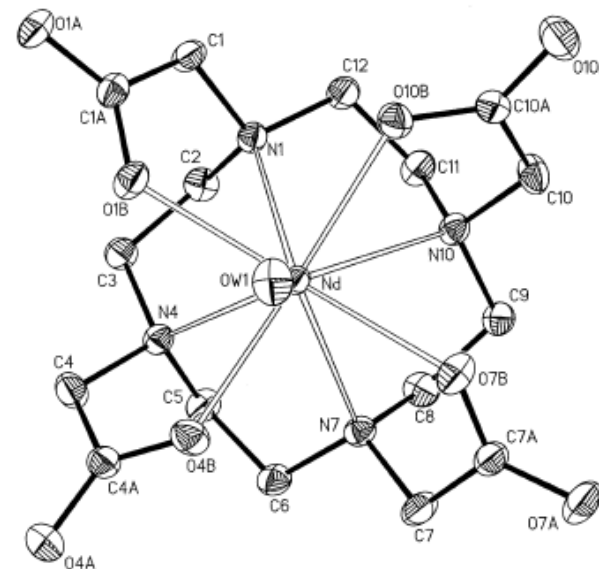
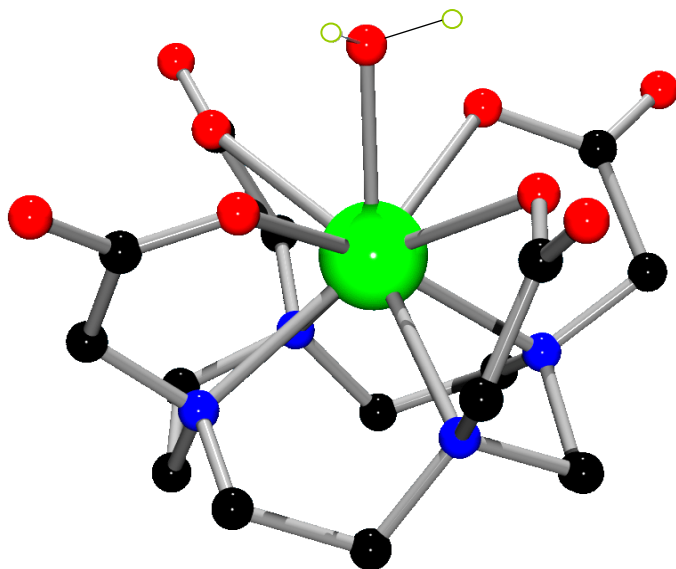
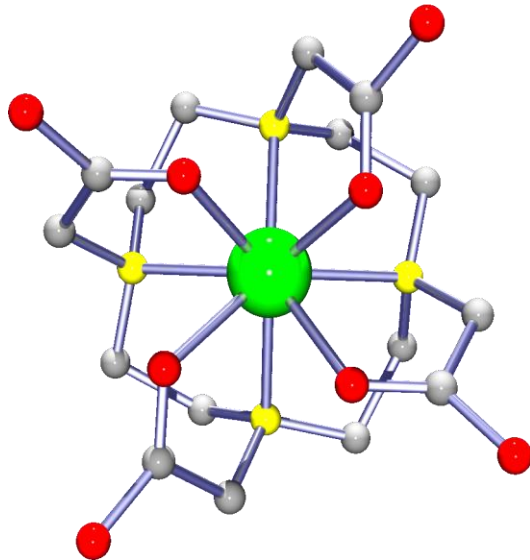
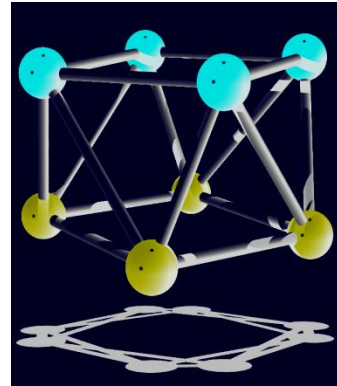
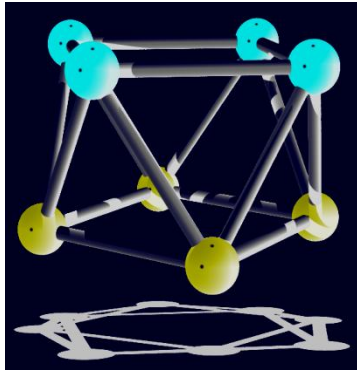


Figure 4. SA geometry in $[\text{Nd}(\text{DOTA})(\text{H}_2\text{O})]^-$, ORTEP view.

SAP

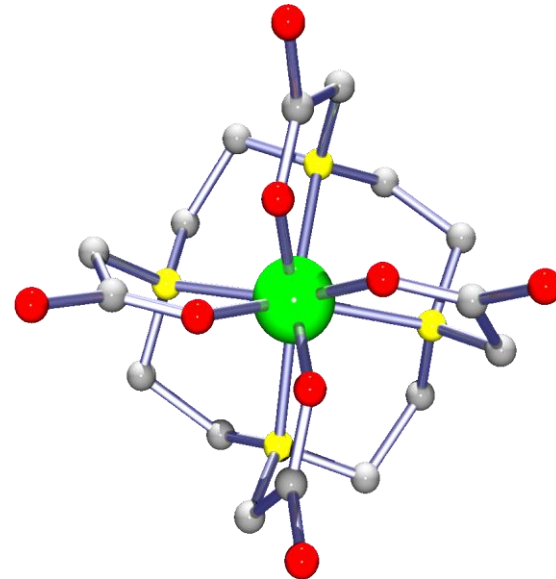
square antiprismatic geometry

k_{ex} and molecular geometry



SAP
Pr-Ho

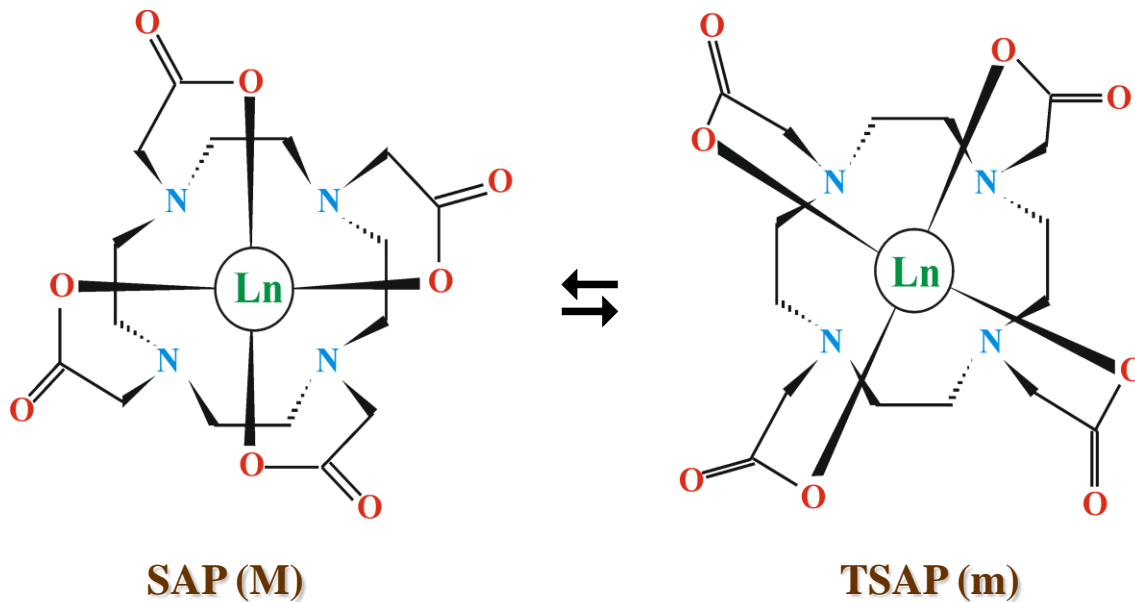
in the solid state



TSAP
La-Ce

k_{ex} and molecular geometry

LnDOTA complexes are present in solution as a mixture of the 2 isomers



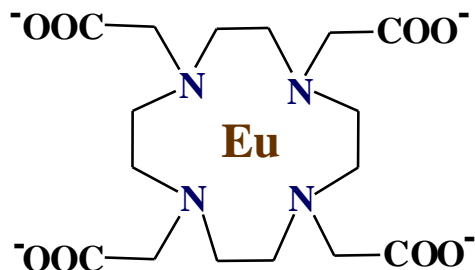
$$k_{ex} (m) \gg k_{ex} (M)$$

for Eu complexes of tetraamide DOTA derivatives

Angew. Chem. Int. Ed., **1998**, 37, 2673

J. Am. Chem. Soc., **1999**, 121, 5762

k_{ex} and molecular geometry



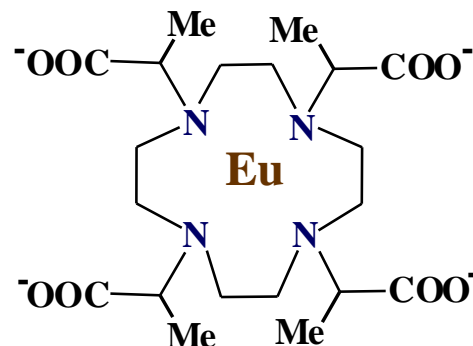
DOTA

SAP >> TSAP

$$K^{298} = [m]/[M] = 0.21$$

$$\Delta H^\circ = 3.4 \text{ (kJ/mol)}$$

$$\Delta S^\circ = -1 \text{ (J/mol}\cdot\text{K)}$$



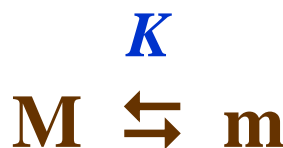
DOTMA

SAP << TSAP

$$K^{298} = [m]/[M] = 4.5$$

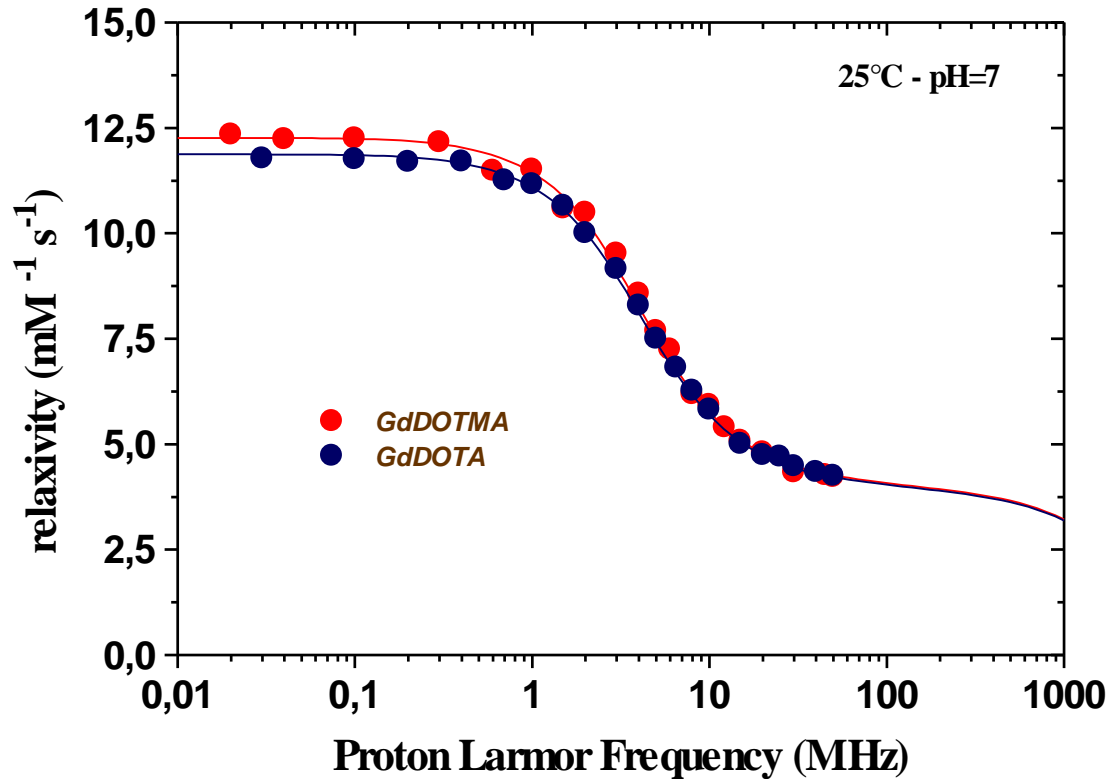
$$\Delta H^\circ = 6.0 \text{ (kJ/mol)}$$

$$\Delta S^\circ = 33 \text{ (J/mol}\cdot\text{K)}$$



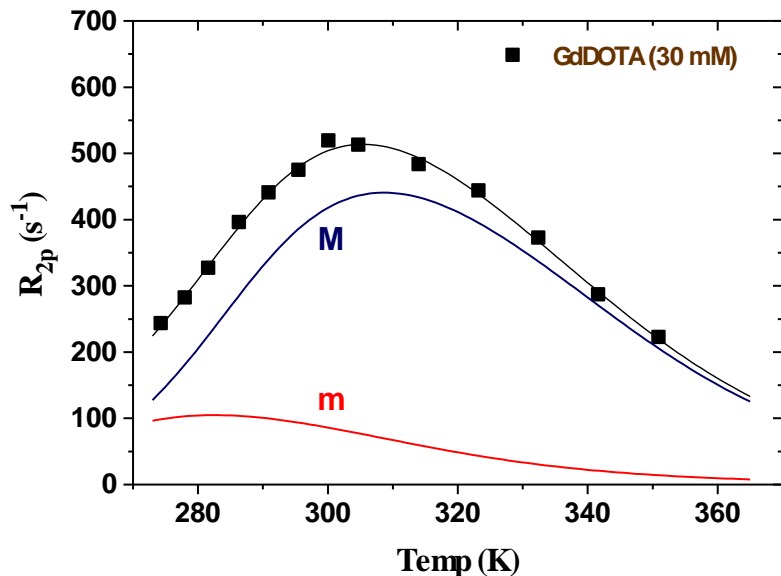
(*Inorg.Chem.*, 1997, 36, 2059)

k_{ex} and molecular geometry



- **The electronic relaxation times of the two isomers are very similar**

k_{ex} and molecular geometry



$$\tau_M (M) = 376 \text{ ns}$$

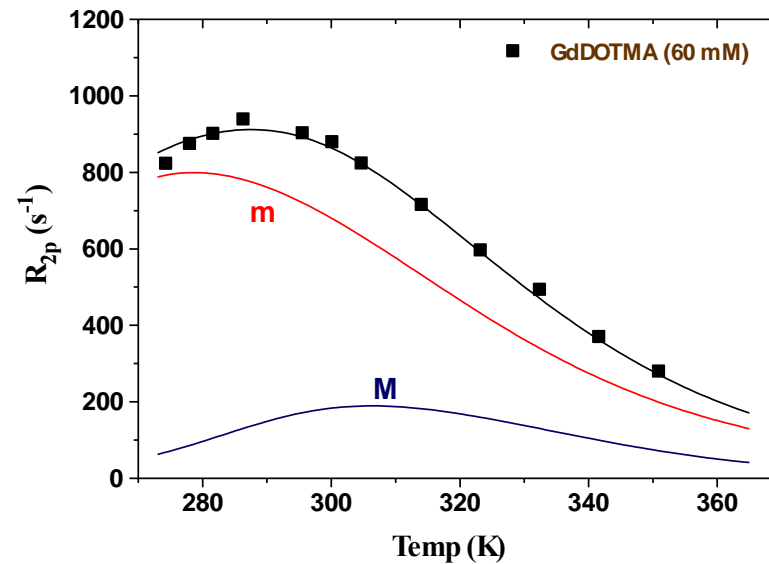
$$\tau_M (m) = 41 \text{ ns}$$

$$\Delta H_M (M) = 55 \text{ kJ/mol}$$

$$\Delta H_M (m) = 48 \text{ kJ/mol}$$

$$\Delta^2 = 1.2 \times 10^{19} \text{ s}^{-2}$$

$$\tau_V = 6.4 \text{ ps}$$



$$\tau_M (M) = 380 \text{ ns}$$

$$\tau_M (m) = 42 \text{ ns}$$

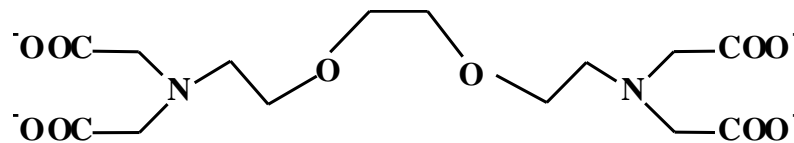
$$\Delta H_M (M) = 54 \text{ kJ/mol}$$

$$\Delta H_M (m) = 40 \text{ kJ/mol}$$

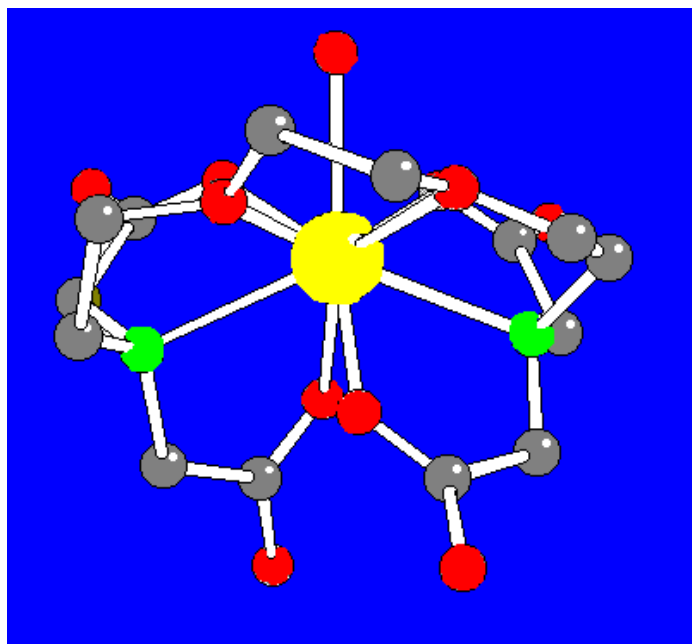
$$\Delta^2 = 1.1 \times 10^{19} \text{ s}^{-2}$$

$$\tau_V = 6.0 \text{ ps}$$

k_{ex} and steric interactions



EGTA



The high water exchange rate of $[\text{Gd}(\text{EGTA})(\text{H}_2\text{O})]^-$ is explained in terms of a limiting dissociative exchange mechanism favoured by the **steric constraints** on the water binding site

$$k_{\text{ex}} = 3.1 \times 10^7 \text{ s}^{-1}$$

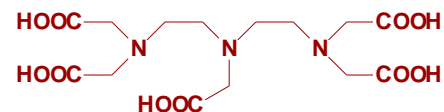
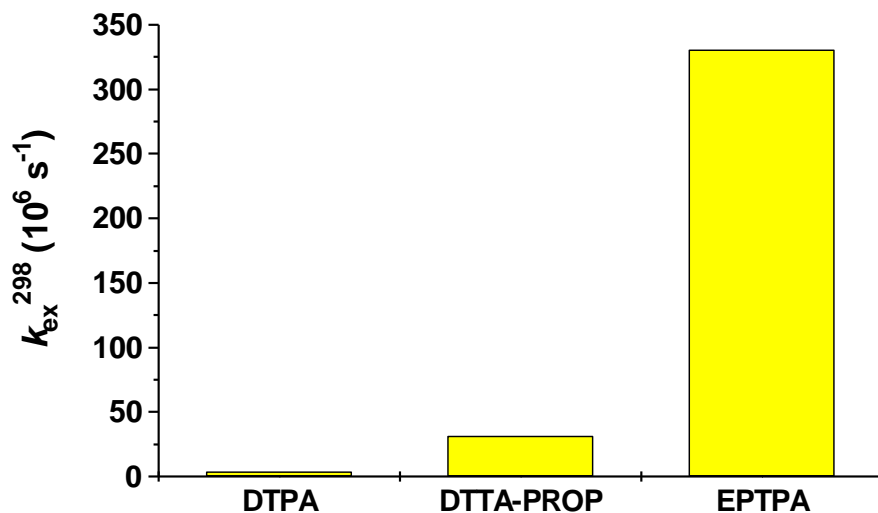


Inorg. Chem., **1997**, 36, 5104

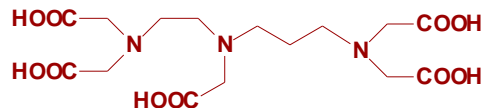
acceleration of water exchange by inducing steric compression at the water binding site

E. Ruloff, É. Tóth, R. Scopelliti, R. Tripier, H. Handel, A. E. Merbach, *Chem. Commun.*, **2002**, 2630
S. Laus, R. Ruloff, É. Tóth, A. E. Merbach, *Chem. Eur. J.*, **2003**, 9, 3555

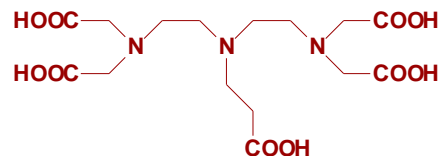
k_{ex} and steric effects



DTPA

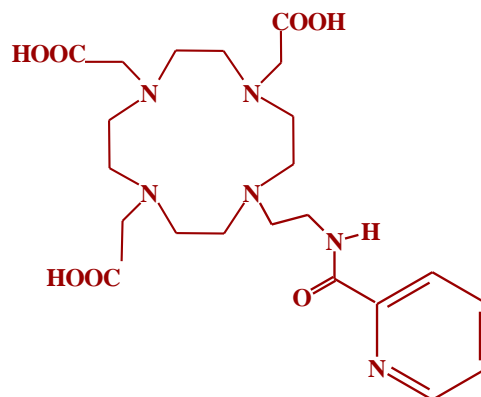
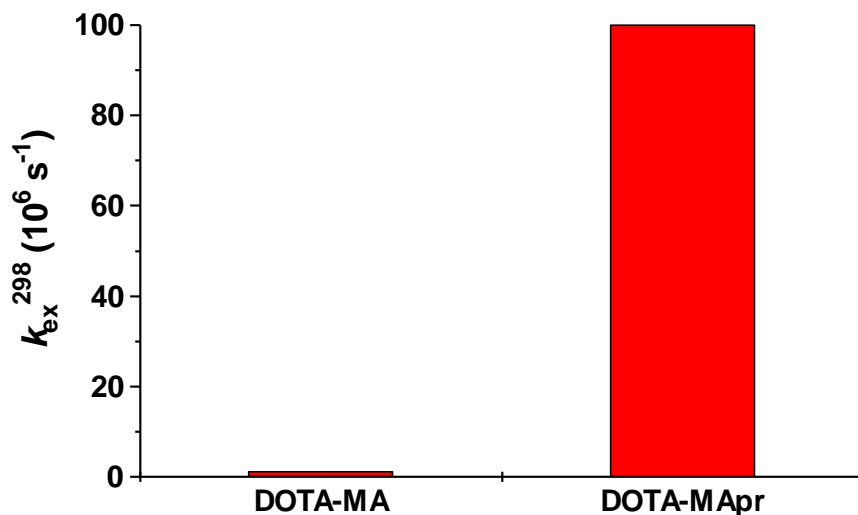


EPTPA



DTTA-PROP

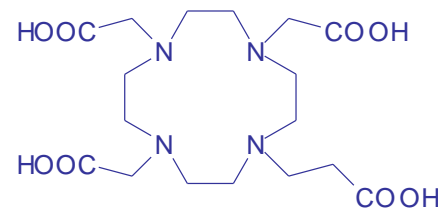
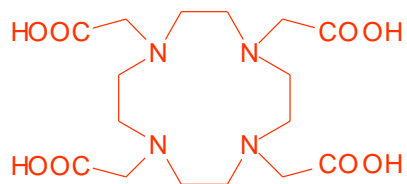
Merbach, Toth et al. *Chem. Eur. J.*, **2003**, 3555



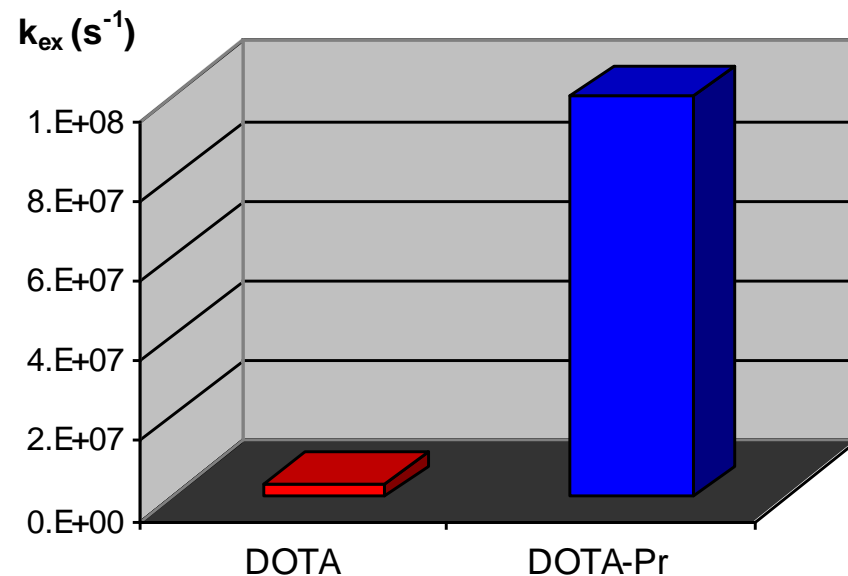
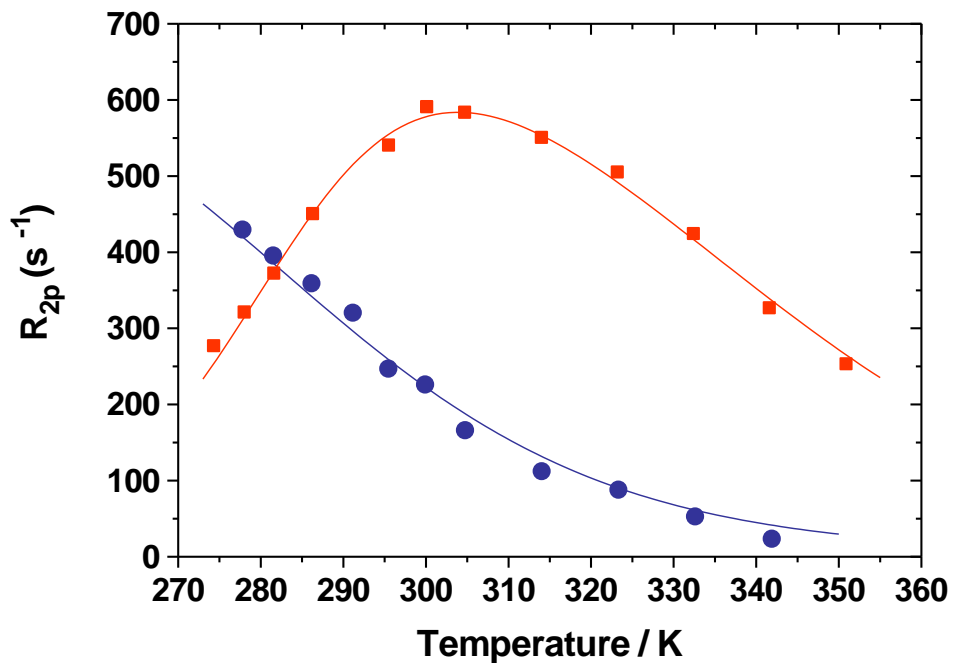
DOTA-MApr

Parker, Botta et al. *Dalton Trans.*, **2004**, 1441

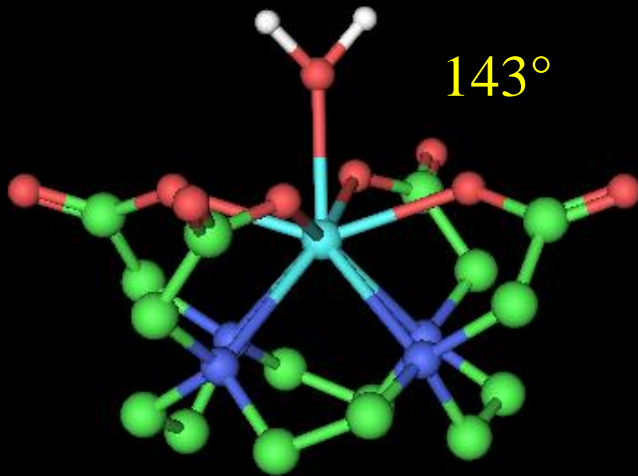
k_{ex} and steric effects



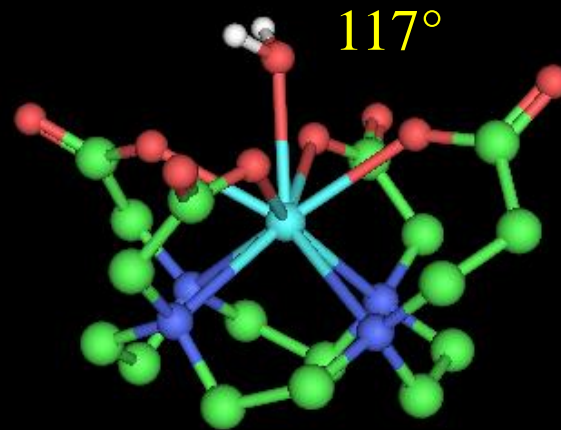
^{17}O NMR - 2.1 T



k_{ex} and steric effects

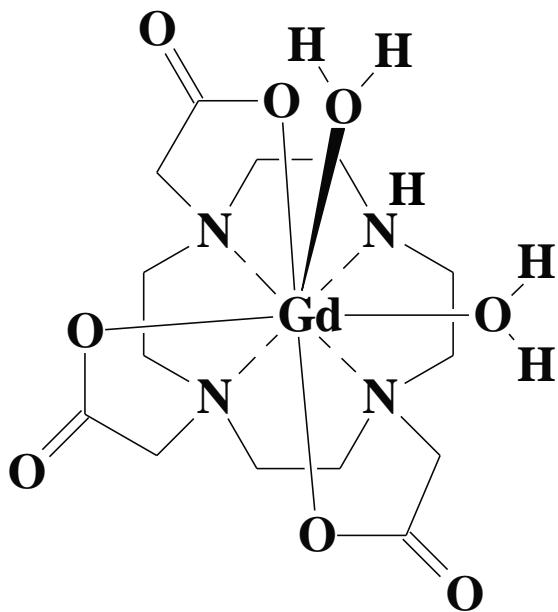


GdDOTA



GdDOTA-Pr

k_{ex} and electronic effects



GdDO3A

$$q = 1.88$$

$$\tau_M = \underline{91 \text{ ns}} (25^\circ\text{C})$$

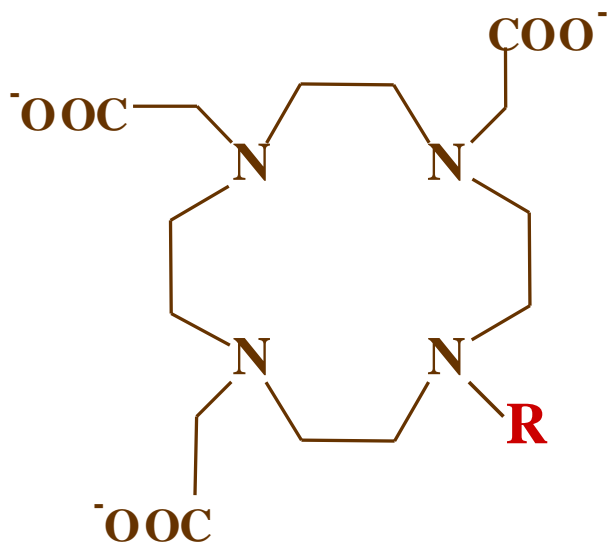
$$\Delta^2 = 3 \times 10^{19} \text{ s}^{-2}$$

$$\tau_V = 16 \text{ ps}$$

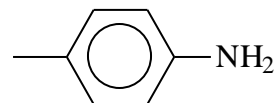
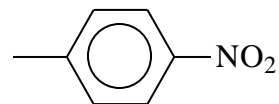
*Merbach, Toth, Helm,... *Magn. Reson. Chem.*, **1999**, 37, 701

k_{ex} and electronic effects

Can τ_M be modulated by introducing a substituent at the secondary nitrogen atom ?

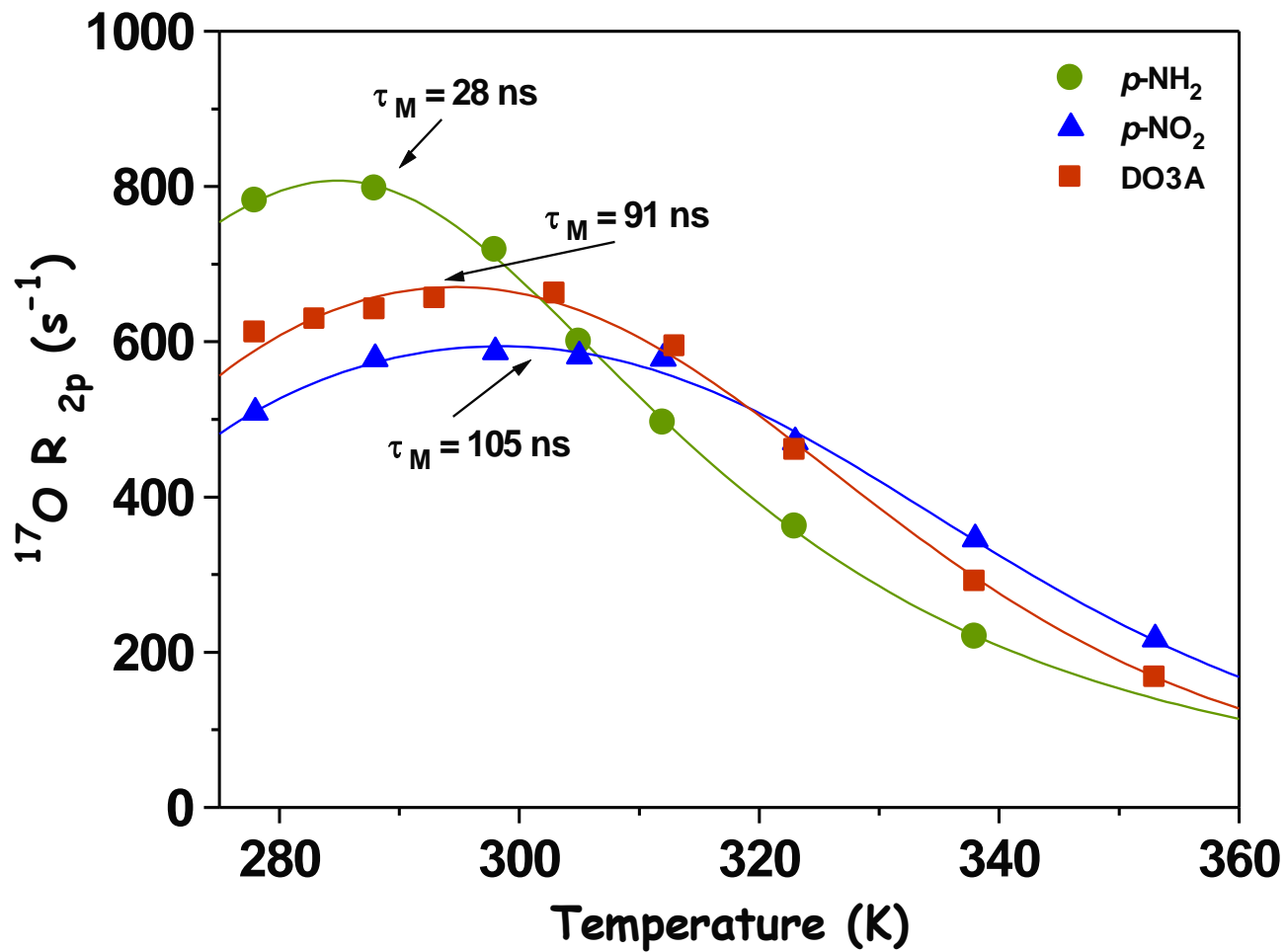


$R =$

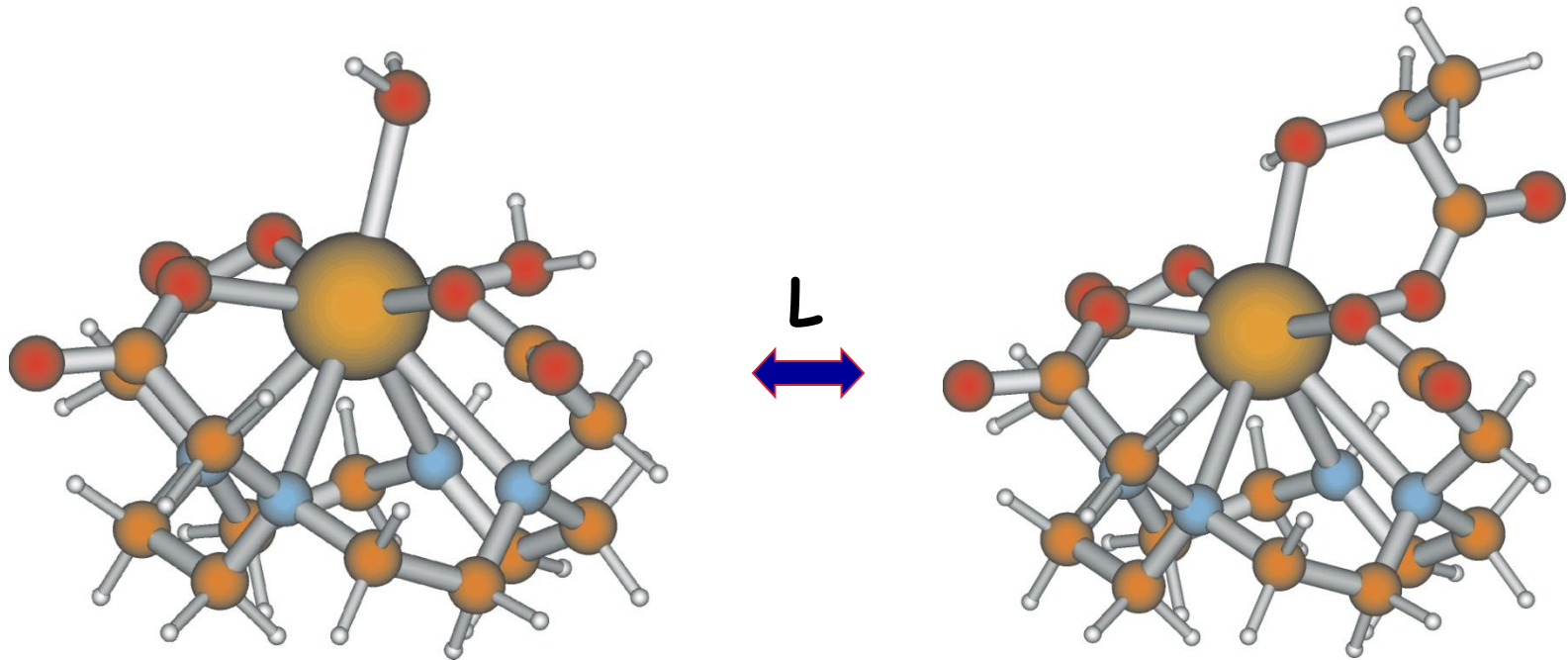


k_{ex} and electronic effects

^{17}O NMR - 7 T

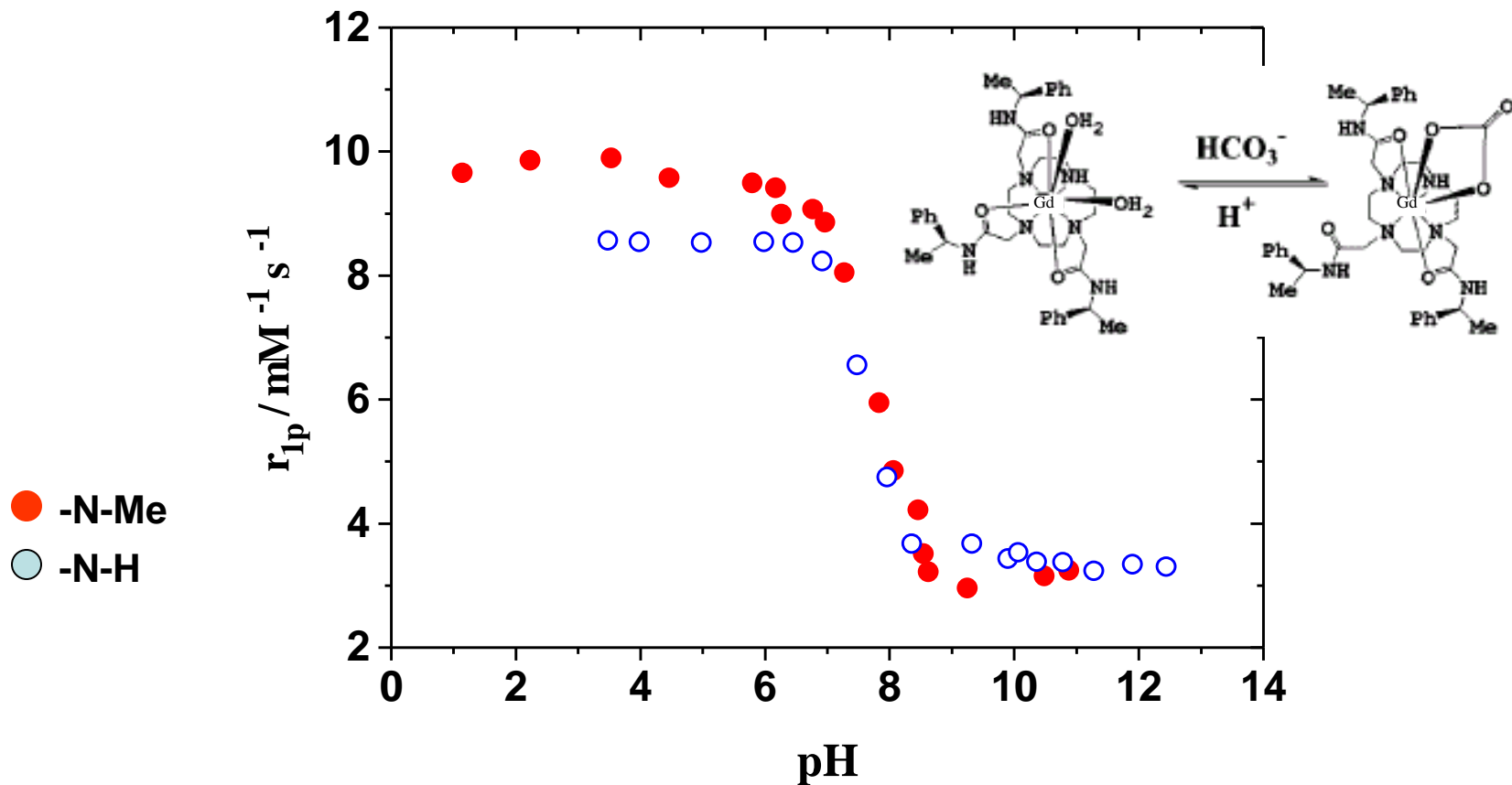


$q=2$ complexes: possible formation of ternary systems!

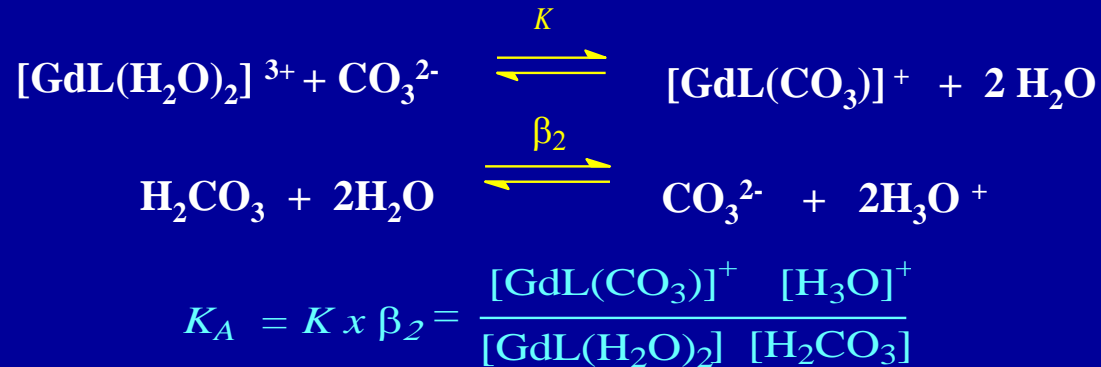


Ternary complexes with carbonate

20 MHz - 25°C



Ternary complexes with carbonate

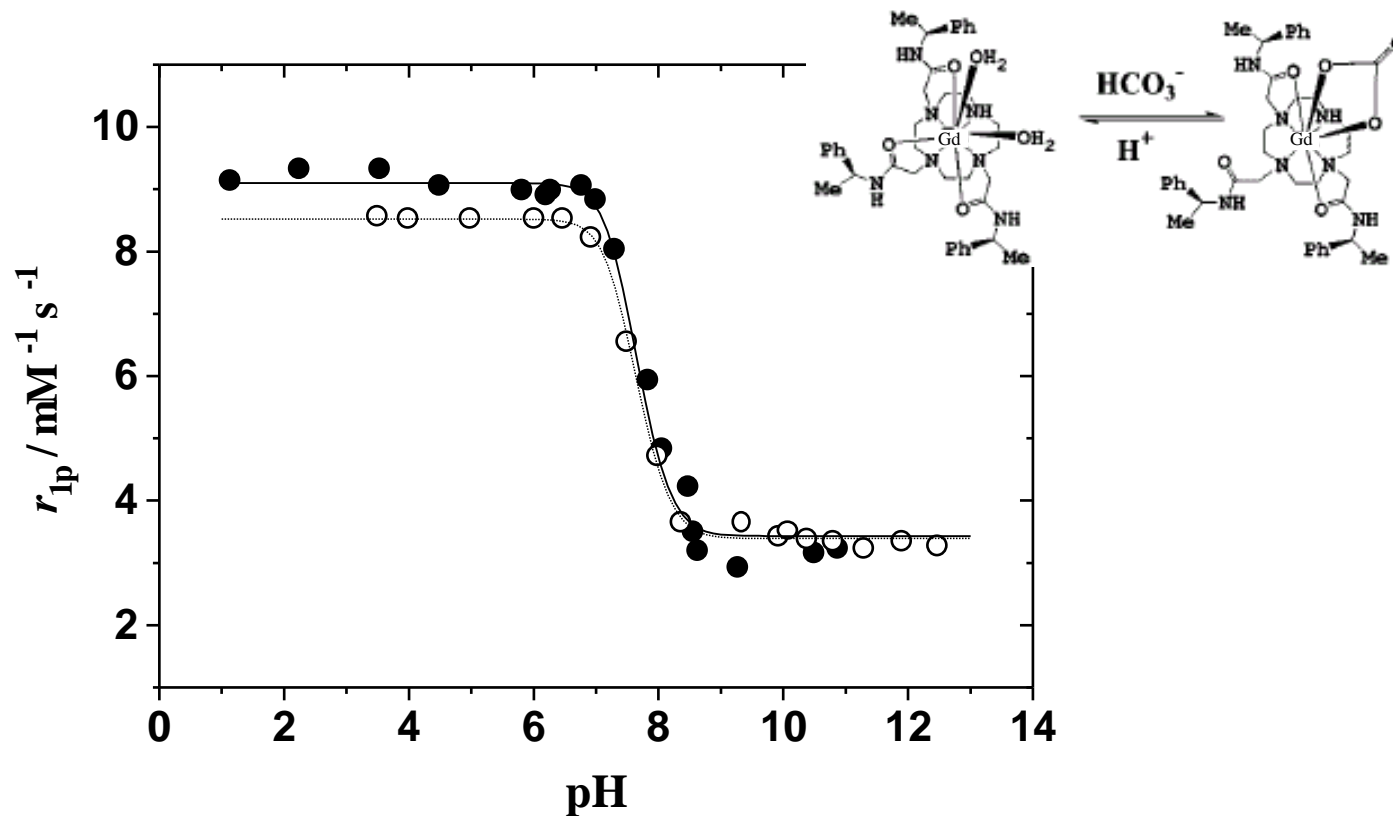


$$R_{1p} = ([C_F]r_{1p}^F + [C_B]r_{1p}^B) \cdot 1000$$

$$[C_B] = \frac{\left(K\beta_2 L_T + K\beta_2 C_T + [H^+]^2 + \beta_2 \right) - \sqrt{\left(K\beta_2 L_T + K\beta_2 C_T + [H^+]^2 + \beta_2 \right)^2 - 4K\beta_2^2 L_T C_T}}{2K\beta_2}$$

Ternary complexes with carbonate

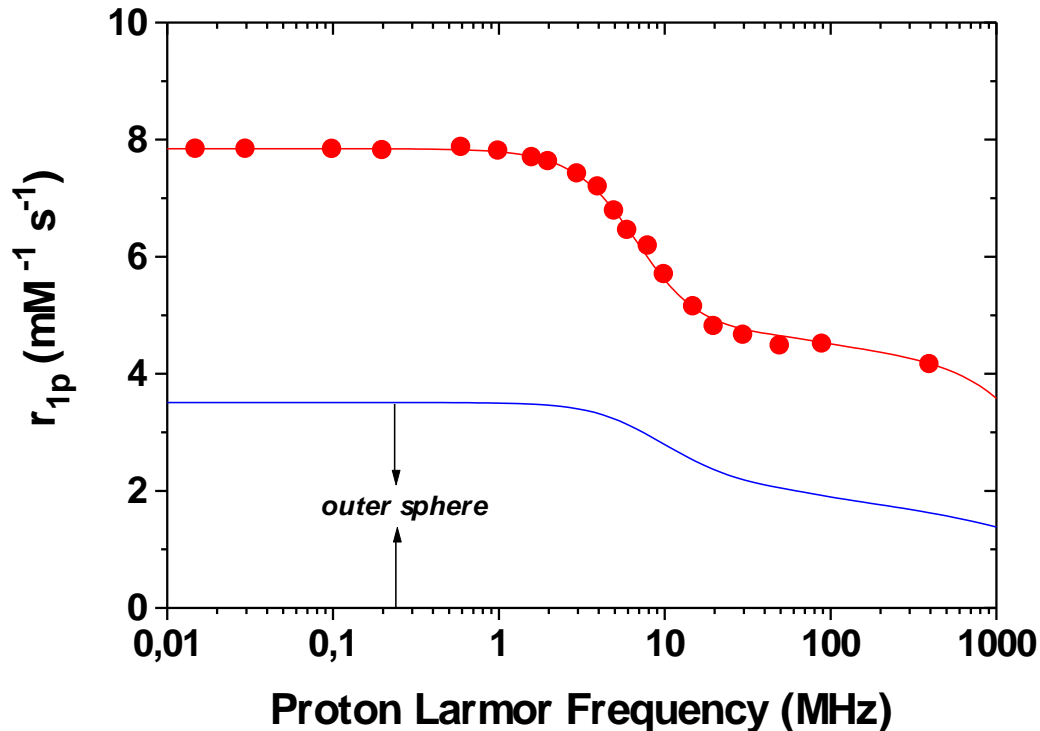
$$K = \begin{cases} 1.9 \times 10^6 \text{ M} & \bullet \text{ -N-Me} \\ 1.1 \times 10^6 \text{ M} & \circ \text{ -N-H} \end{cases}$$



“second sphere” relaxivity

second sphere contribution to relaxivity

$[\text{Gd}(\text{DTPA})(\text{H}_2\text{O})]^{2-}$: nature of the hydration spheres

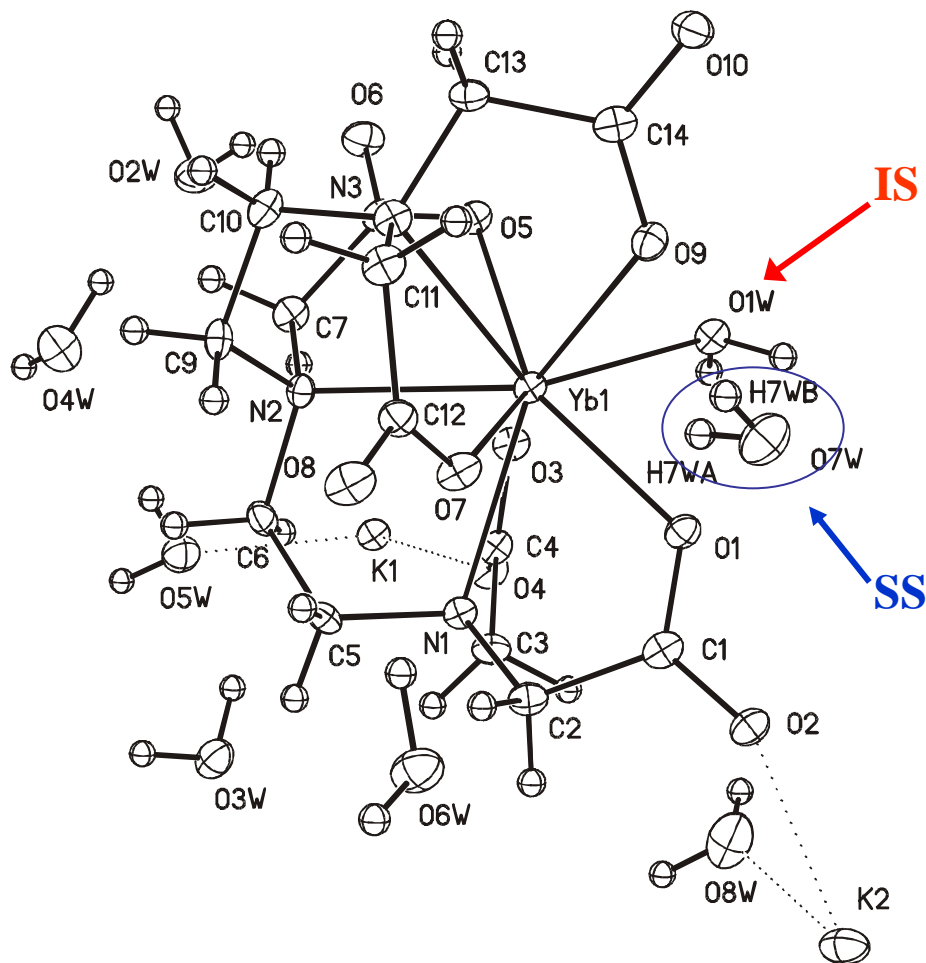
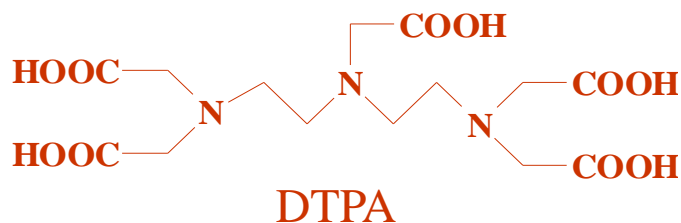


$q = 1$
 $r = 3 \text{ \AA}$
 $\tau_R = 74 \text{ ps}$
 $\tau_M = 330 \text{ ns}$
 $a = 3.5 \text{ \AA}$
 $D = 2.24 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$

second sphere contribution to relaxivity

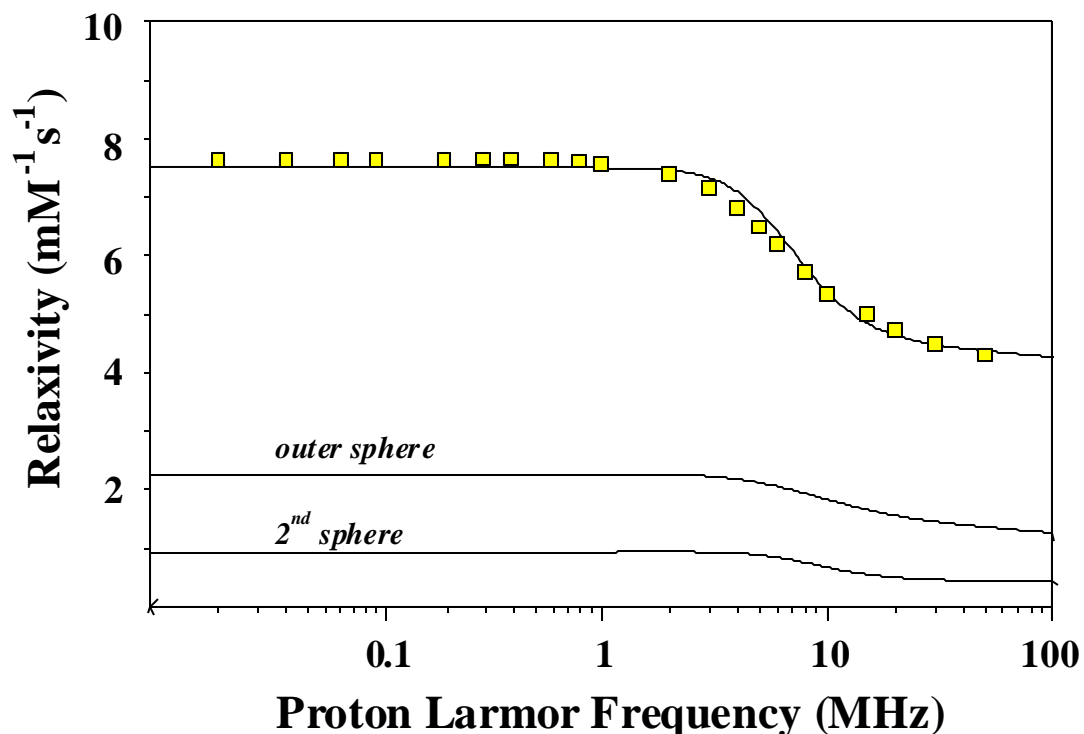
three well-defined hydration shells can be observed consisting of:

- i) **one coordinated water molecule;**
- ii) **several water molecules in the outer coordination sphere of the Yb^{III} ion and**
- iii) **one water molecule surprisingly close to the Yb and H-bonded to proximate -COOH groups**



Eur. J. Inorg. Chem., **2000**, 971-977

second sphere contribution to relaxivity



$$q = 1$$

$$r = 2.95 \text{ \AA}$$

$$\tau_R = 65 \text{ ps}$$

$$\tau_M = 330 \text{ ns}$$

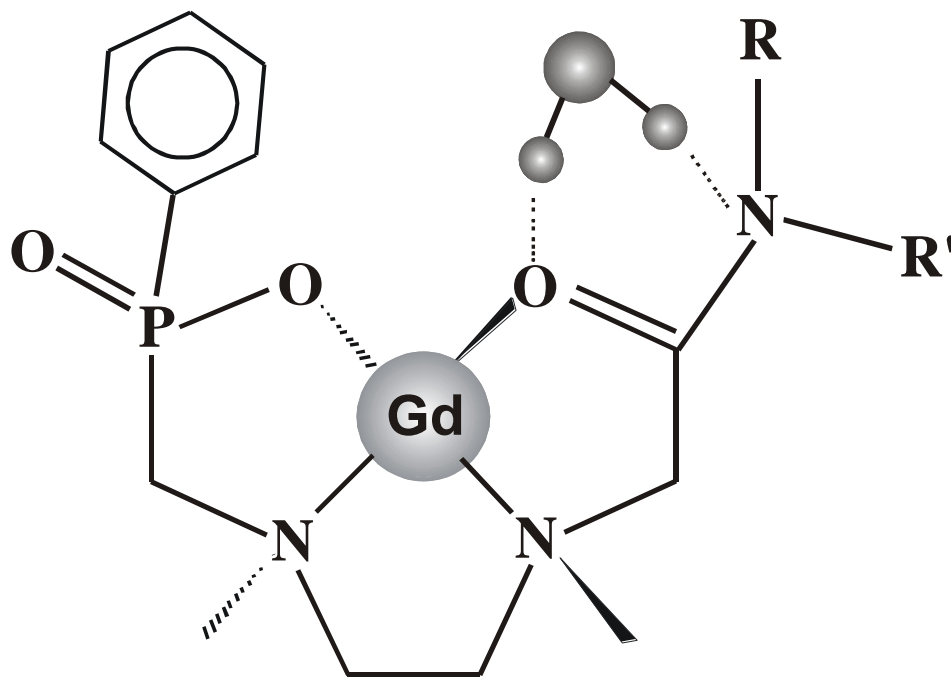
$$a = 4.5 \text{ \AA}$$

$$q' = 1$$

$$r' = 3.28 \text{ \AA}$$

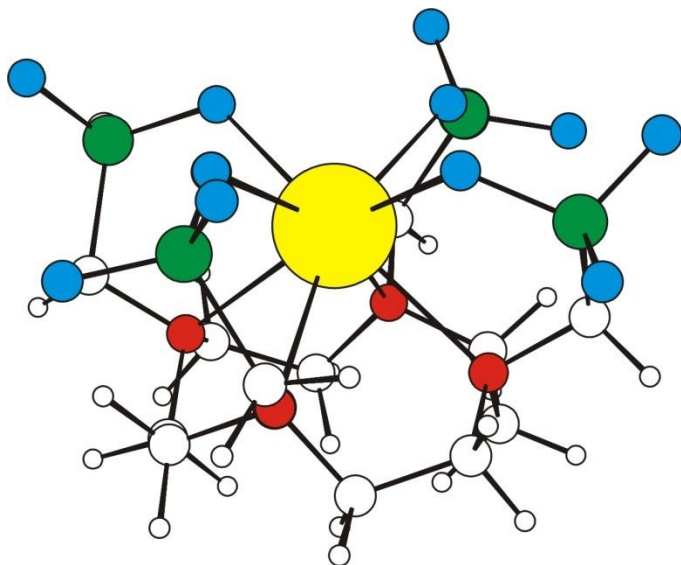
“The contribution of the SS term is about 10% of the total relaxivity but is very important for a correct and more realistic interpretation of the experimental data. Furthermore, its contribution can be optimized and the relaxivity enhanced through a suitable ligand design.”

second sphere contribution to relaxivity



Schematic representation of the *second sphere* water molecule affecting the relaxivity of the complex GdDOTMP-MBu₂

second sphere contribution to relaxivity



GdDOTP

$$q = 0$$

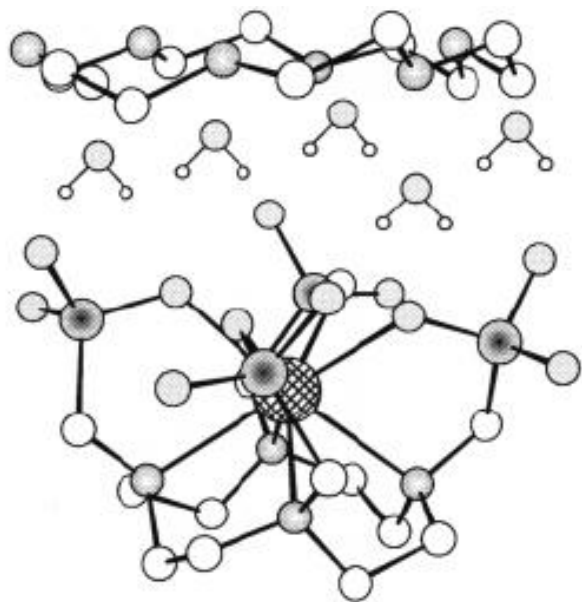
$$r_{1p} = 4.58$$



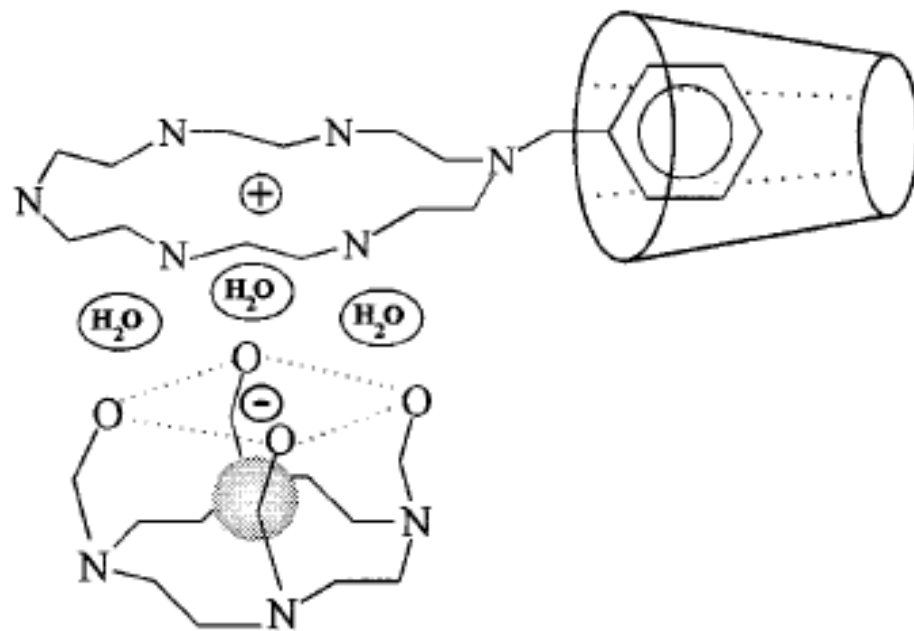
**strong contribution from
water molecules H-bonded
to phosphonate groups**

second sphere contribution to relaxivity

enhancement of r_{1p} (ss)



$$r_{1p} = 7.7$$



$$r_{1p} = 18.1$$

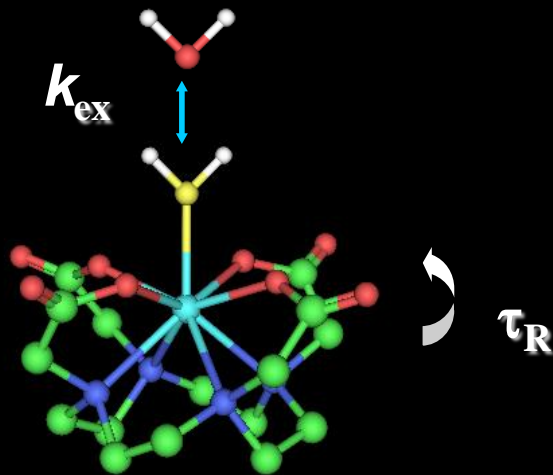
$$r_{1p} (\text{os}) = 2.5$$

Relaxivity enhancement
at
high fields (> 1 T)

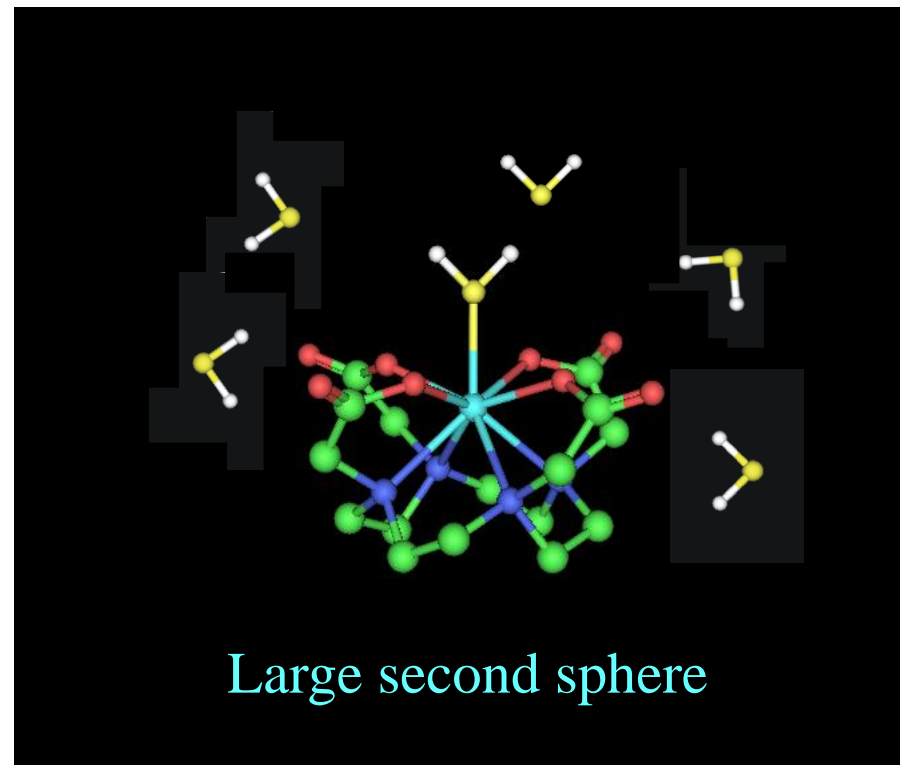
Relaxivity enhancement at high fields

medium molecular weight conjugates

- efficient coupling of the local motion of the Gd-OH₂ vector with the rotational motion of the whole complex
- fast rate of water exchange
- large number second sphere water molecules



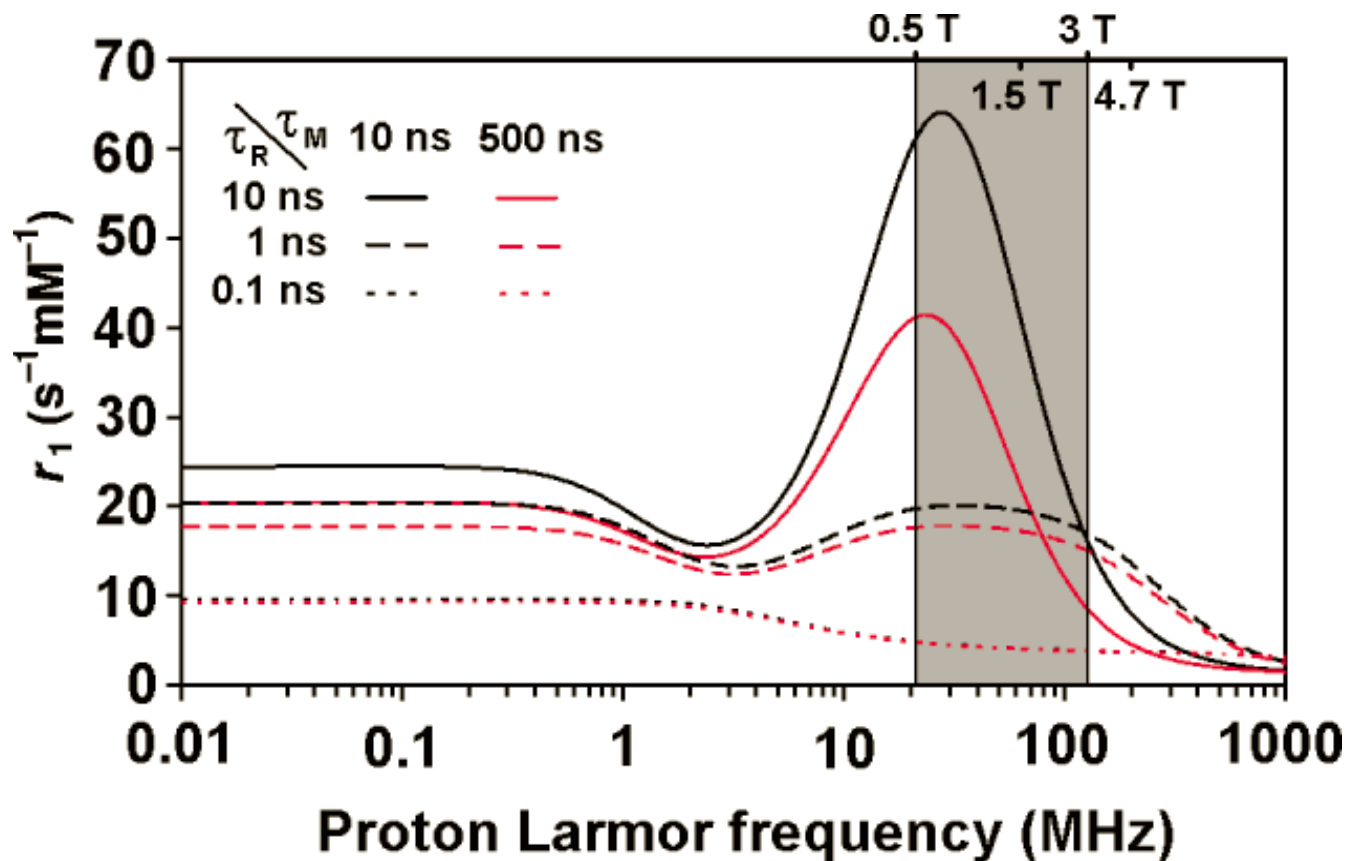
Slow rotation and fast exchange



Large second sphere

Relaxivity enhancement at high fields

medium molecular weight conjugates

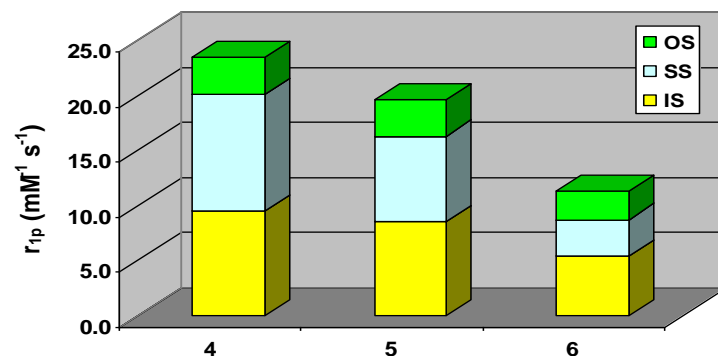
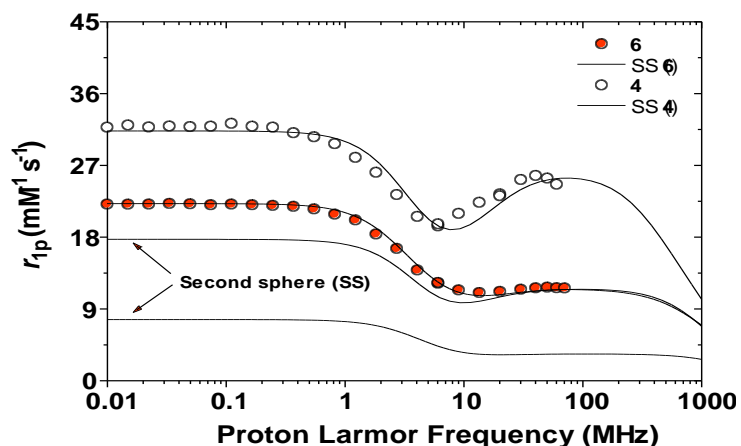
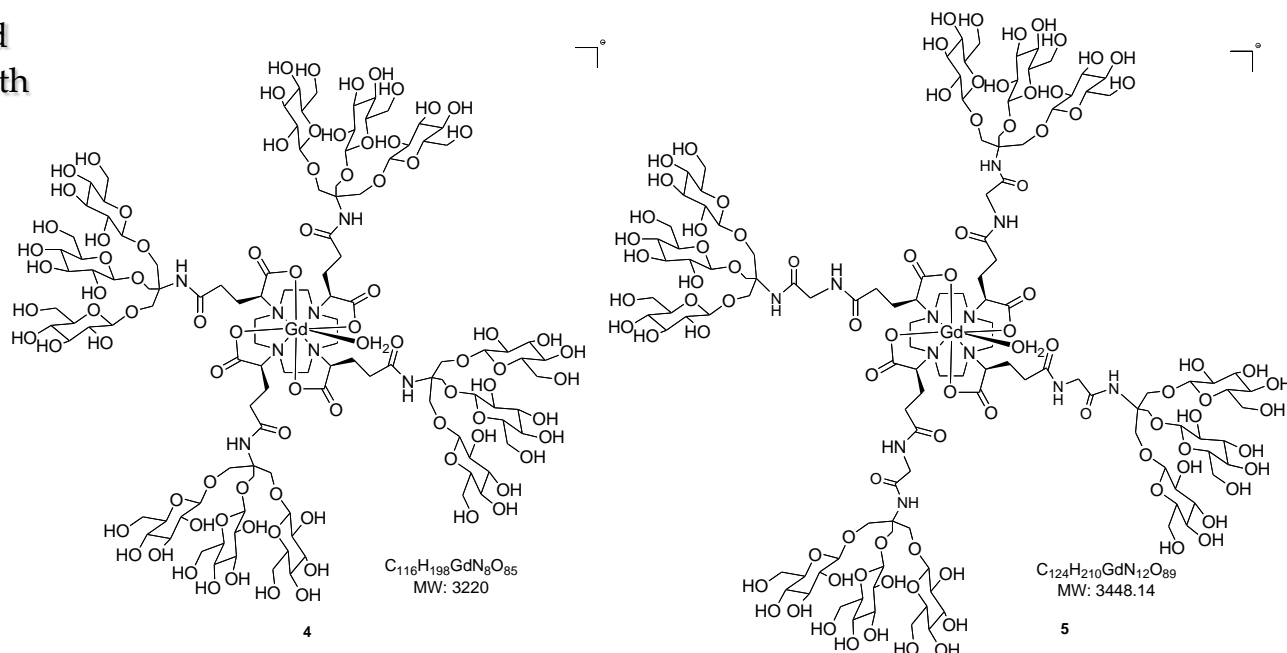


Relaxivity enhancement at high fields

hydrophilic dendritic structure, packed with second sphere H₂O molecules, with the Gd ion at the focal point

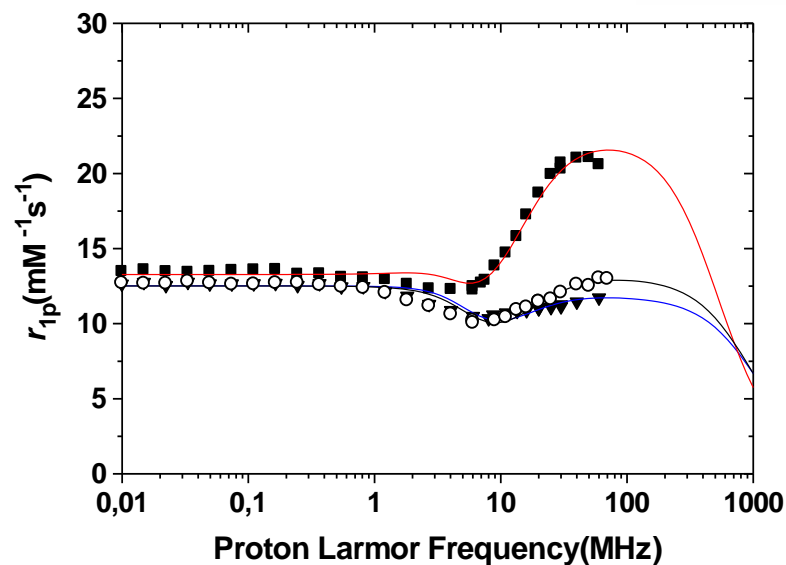
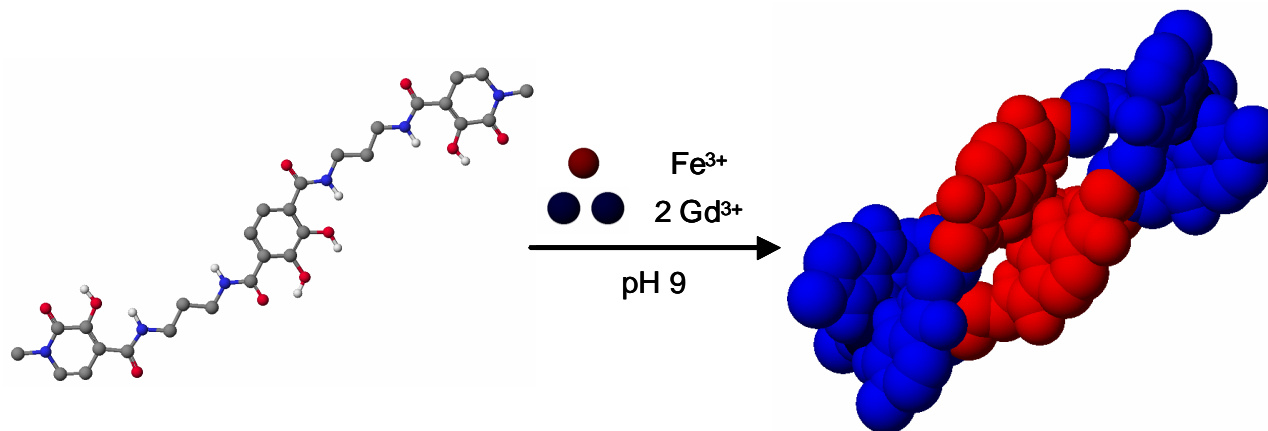
q=1

efficient coupling of the local motion of the Gd-OH₂ vector with the rotational motion of the whole complex fast rate of water exchange large number second sphere water molecules



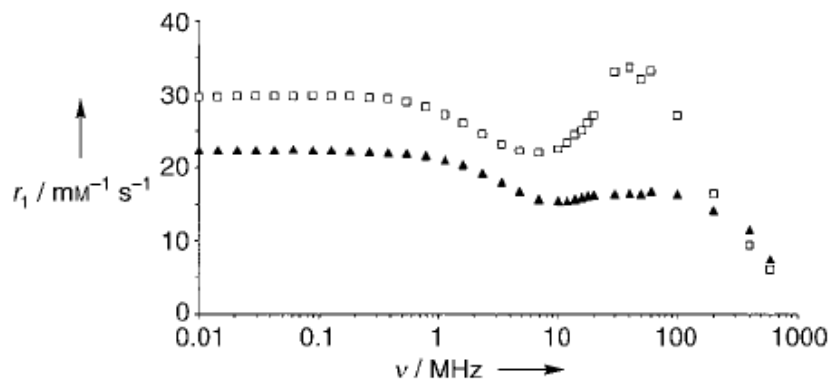
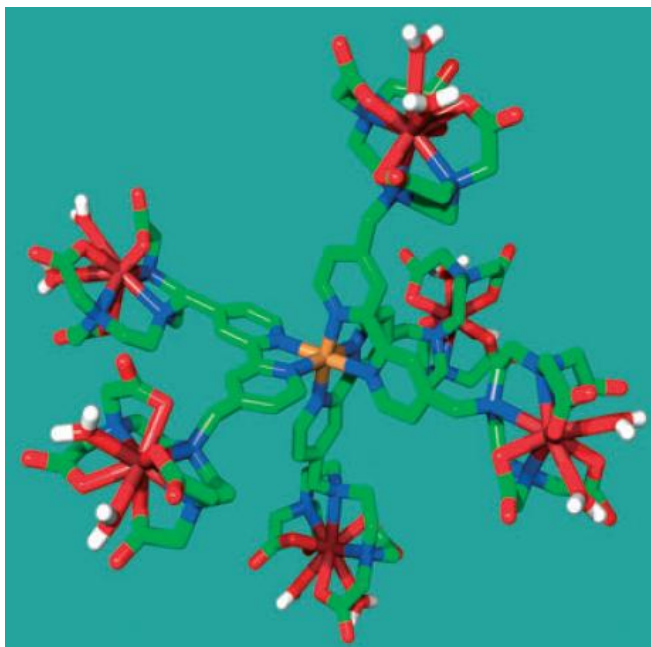
Relaxivity enhancement at high fields

Fe(III) Templated Gd(III) Self-Assemblies



Relaxivity enhancement at high fields

Metallostar → MW = 3744 g mol⁻¹

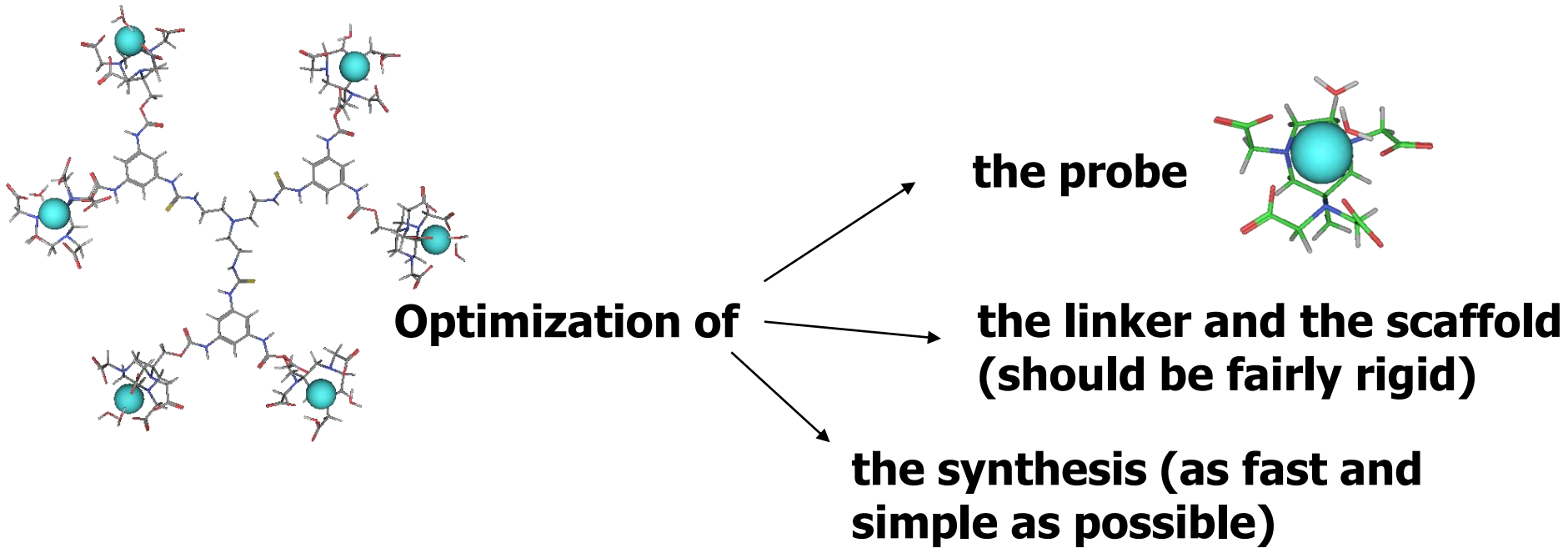


J. B. Livramento, E. Toth, A. Sour, A. Borel, A. E. Merbach, and R. Ruloff, *Angew. Chem. Int. Ed.* **2005**, *44*, 1480

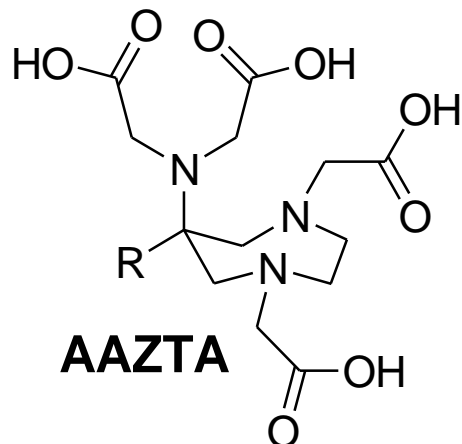
The Gd complexes must have high thermodynamic stability and kinetic inertness

How to increase r_{1p} at high fields?

Multimeric systems



How to increase r_{1p} at high fields?



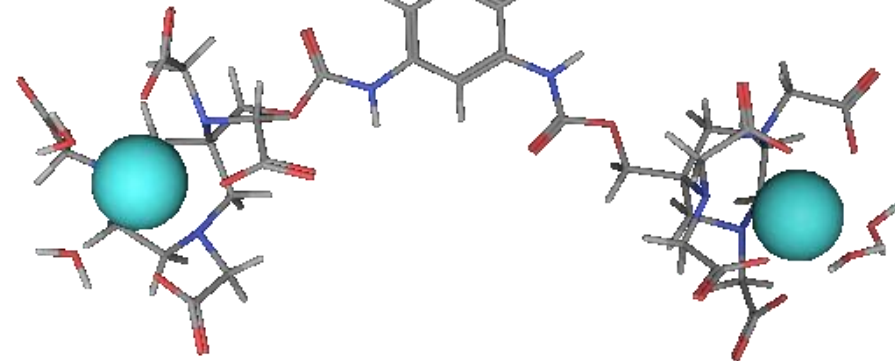
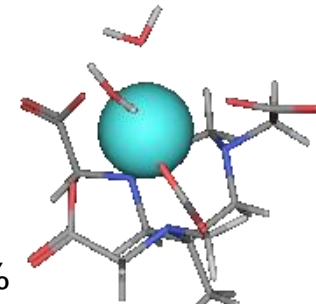
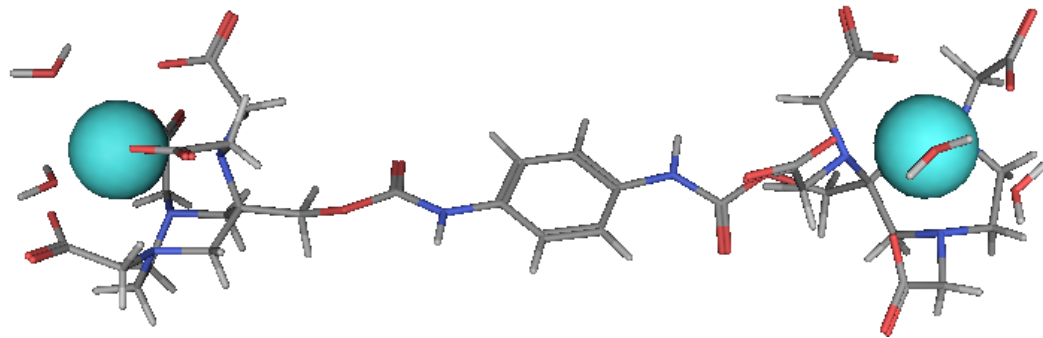
Easily synthesized heptadentate ligand
In 4 steps from cheap starting materials

Giovenzana et al. *Inorg. Chem.* **2004**, 43, 7588.

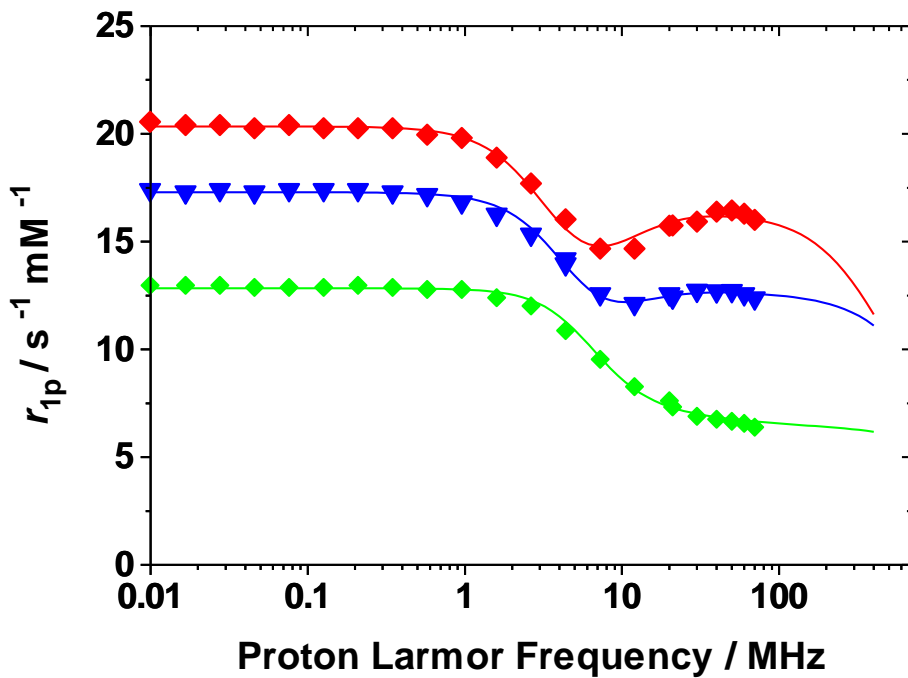
- **Good thermodynamic stability**
(Log K_f = 20.24)
- **Unaffected by mono and bidentate anions**
- **Kinetically inert towards transmetallation**

Baranyai, Z.; Uggeri, F.; Giovenzana, G. B.; Benyei, A.;
Brucher, E.; Aime S. *Chem. Eur. J.* **2009**, 15, 1696.

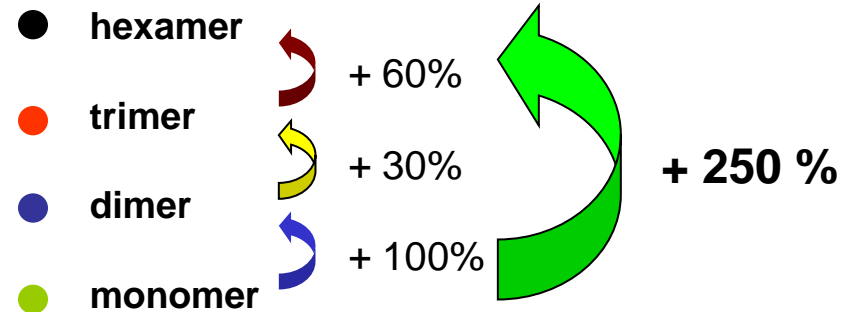
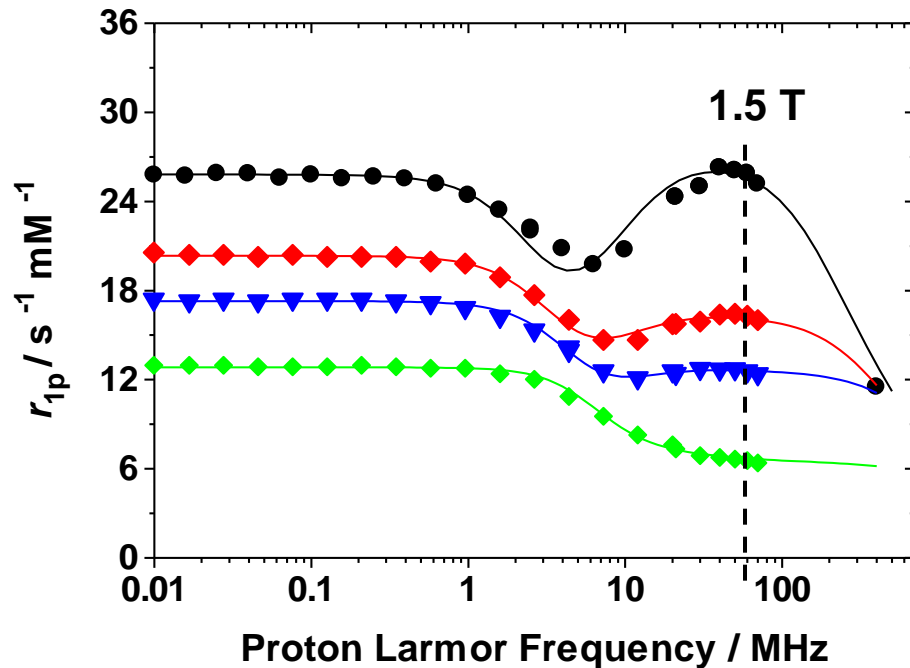
from monomer to trimer



- trimer + 30%
- dimer
- monomer + 100%



How to increase r_{1p} at high fields?



The hexamer presents high relaxivity **per Gd** over a broad range of imaging fields (0.5-3.0 T).

r_{1p} **per molecule** ranges from **138** $\text{mM}^{-1}\text{s}^{-1}$ at 3.0 T to **156** at 1.5 T (**145** at 0.5 T)

Development

Functionalization of multimers
with targeting vectors

References

- (1) Lauffer, R.B. *Chem. Rev.* **1987**, *87*, 901-927.
- (2) Aime S.; Botta M.; Fasano, M.; Terreno, E. *Chem. Soc. Rev.* **1998**, *27*, 19-29.
- (3) Caravan, P.; Ellison, J.J., McMurry, T.J.; Lauffer, R.B. *Chem Rev.* **1999**, *99*, 2293-2352
- (4) Merbach, A.E.; Tóth, E. *The Chemistry of Contrast Agents in Medical Magnetic Resonance Imaging*; John Wiley & Sons: Chichester, **2001**.
- (5) Aime S.; Botta M.; Terreno, E. *Adv. Inorg. Chem.* **2005**, *57*, 173-237.
- (6) Banci, L.; Bertini, I.; Luchinat, C. *Nuclear and Electron Relaxation*; VCH: Weinheim, 1991
- (7) Aime S.; Botta M.; Fasano, M.; Terreno, E. *Acc. Chem. Res.* **1999**, *32*, 941-949. (*water and prototropic exchange*)
- (8) Koenig, S.H.; Brown III, R.D. *Prog. NMR Spectrosc.* **1990**, *22*, 487-567. (*field-cycling relaxometry*)
- (9) Peters, J. A.; Huskens, J.; Raber, D. J. *Prog. NMR Spectrosc.* **1996**, *28*, 283-350. (*Ln-induced shift and relaxation*)
- (10) Botta, M. *Eur. J. Inorg. Chem.* **2000**, 399-407. (*second sphere relaxivity*)
- (11) Helm, L.; Tóth, E.; Merbach, A.E. in *Metal Ions in Biological Systems*; Sigel, A.; Sigel, H. Eds.; Marcel Dekker Inc.: New York, 2003; Chap. 15.
- (12) Tóth, E.; Helm, L.; Merbach, A.E. *Top. Curr. Chem.* **2002**, *221*, 61-101
- (13) Caravan, P. *Chem. Soc. Rev.* **2006**, *35*, 512-523.
- (14) S. Aime, S. Geninatti Crich, E. Gianolio, G.B. Giovenzana, L. Tei, E. Terreno, *Coord. Chem. Rev.*, **2006**, *250*, 1562–1579
- (15) E.J. Werner, A. Datta, C.J. Jocher, K.N. Raymond, *Angew. Chem. Int. Ed.* **2008**, *47*, 8568 – 8580
- (16) C.F.G.C. Geraldes, S. Laurent, *Contrast Media Mol. Imaging*, **2009**, *4*, 1–23