

Strategies for cellular labeling

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Strategies for cellular labeling

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graph TD; A([Strategies for cellular labeling]) --> B[“IN VIVO”]; A --> C[“IN VITRO”]; B --- D[Labelling of targeted cells and visualization of cellular processes in living organisms.]; C --- E[Monitoring of migration and location of transplanted cells labeled “in vitro” with contrast agents.]
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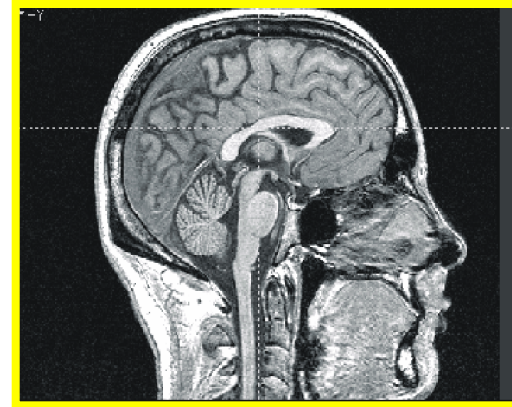
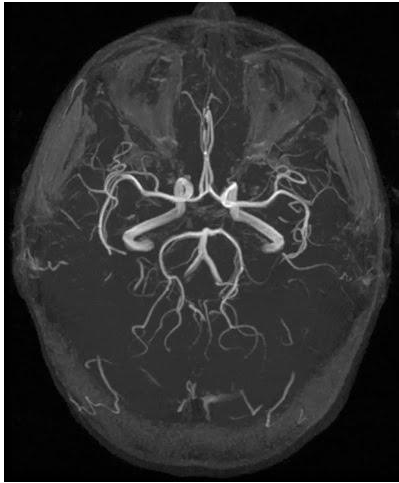
“IN VIVO”

Labelling of targeted cells and visualization of cellular processes in living organisms.

“IN VITRO”

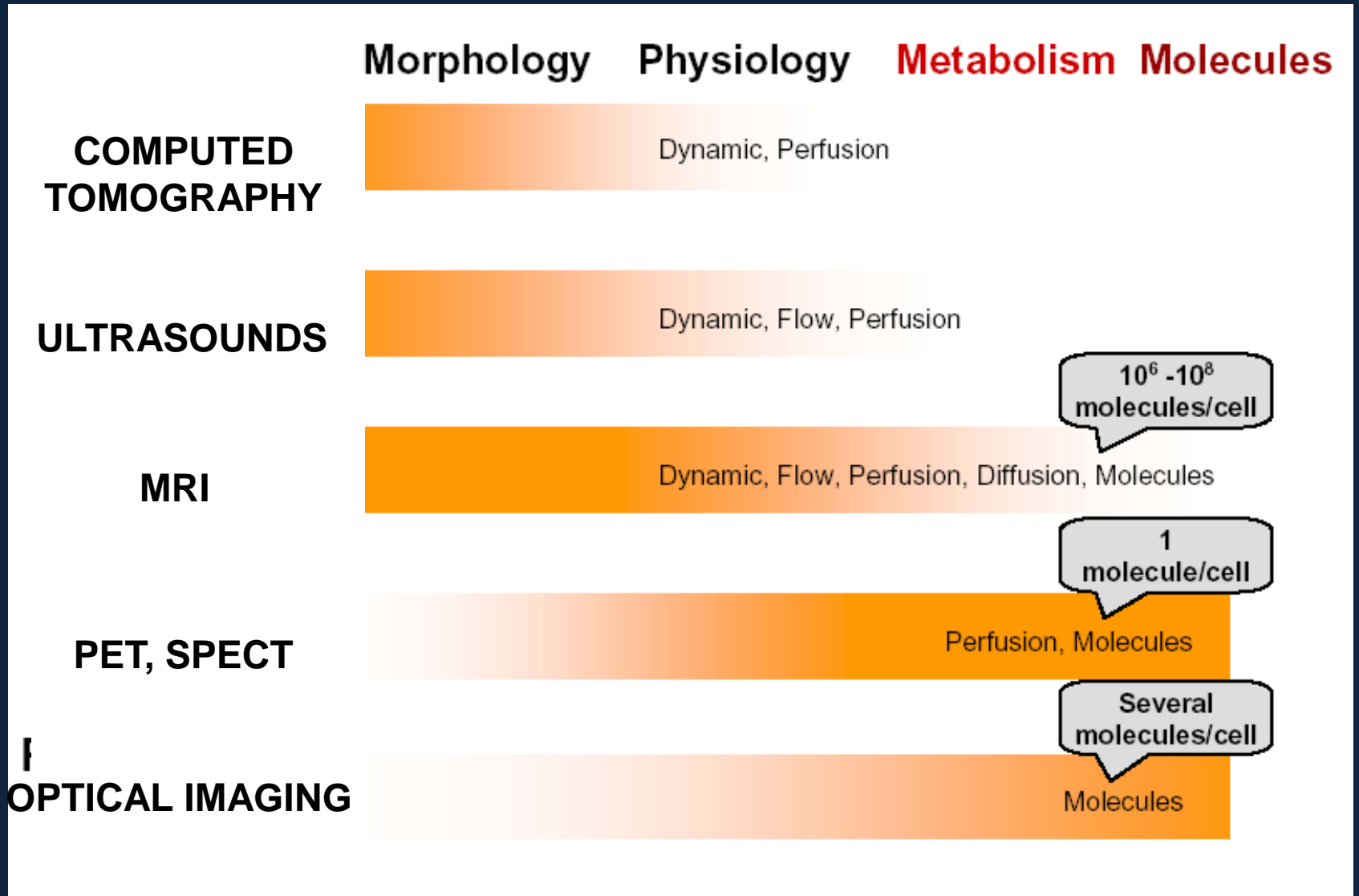
Monitoring of migration and location of transplanted cells labeled “in vitro” with contrast agents.

Magnetic Resonance Imaging



- Non invasive and repetitive imaging
- High resolution
- Absence of radiation
- Low sensitivity

Imaging Modalities: range of detection



MRI Contrast Agents



paramagnetic species



Complexes of
paramagnetic metal ions
(Gd^{3+} , Mn^{2+})

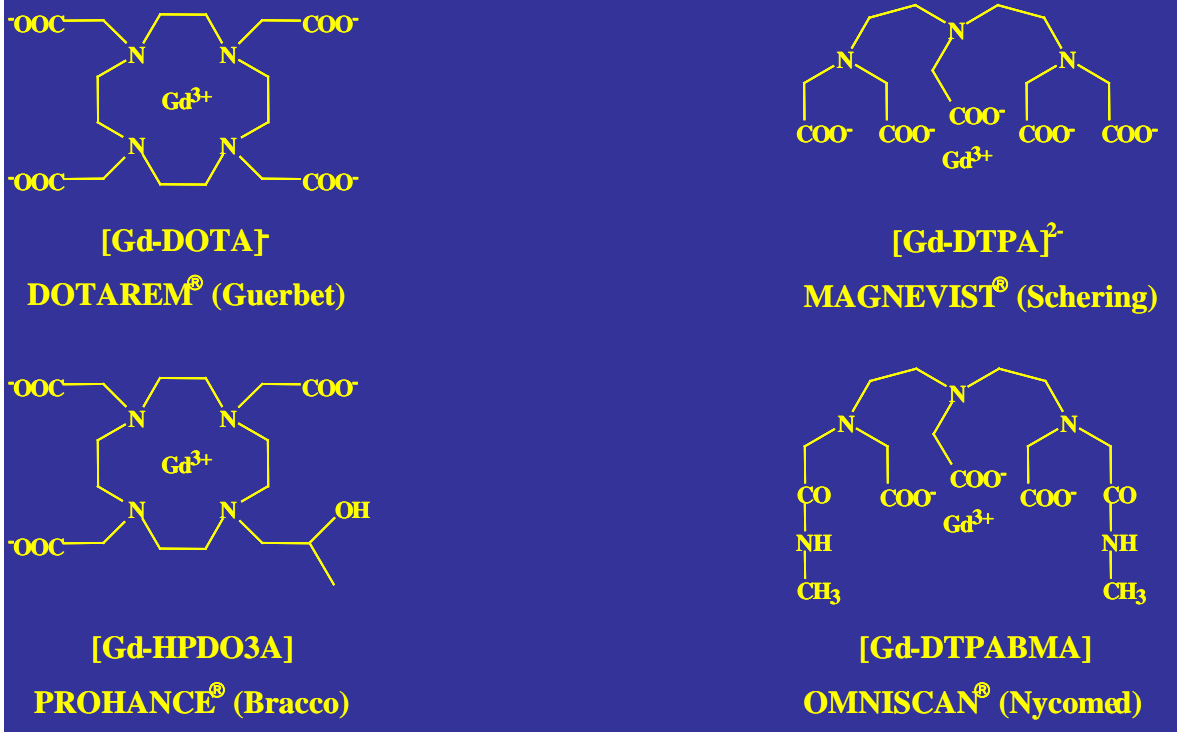
-Positive Contrast

Superparamagnetic
particles (SPIOs, USPIOs)

-Negative Contrast

Clinical MRI Contrast Agents

Clinical dose 0.1 mmol/Kg



Concentration needed to obtain sufficient contrast

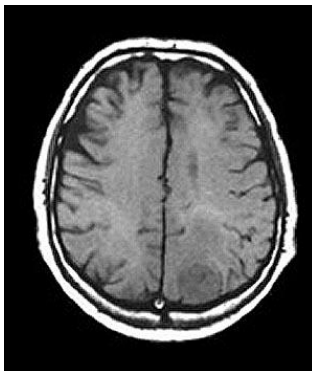


10^{-4} - 10^{-5} M

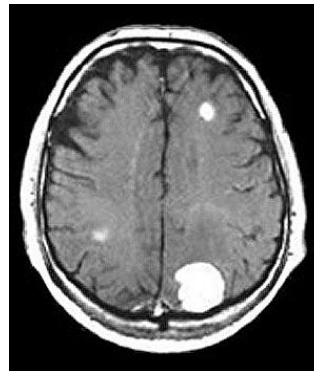
Concentration of target receptors



10^{-6} - 10^{-10} M



Without CA



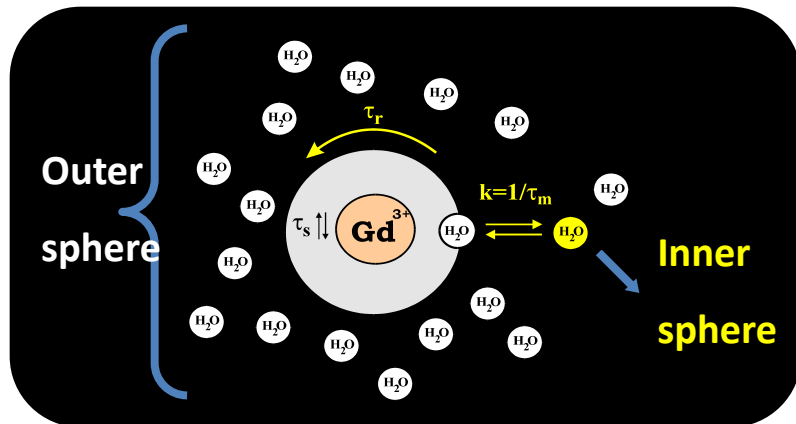
With Gd(III)-based CA

Can Gd^{3+} based probes be considered good candidates for MR Molecular Imaging applications?

Designing Highly Sensitive Agents ; optimization of parameters that influence relaxivity

$R_{1obs} \propto R_1(\text{inner sphere}) + R_1(\text{second sphere})$

(τ_r, τ_M, q)



Increasing the MR-contrast agent concentration at the target site

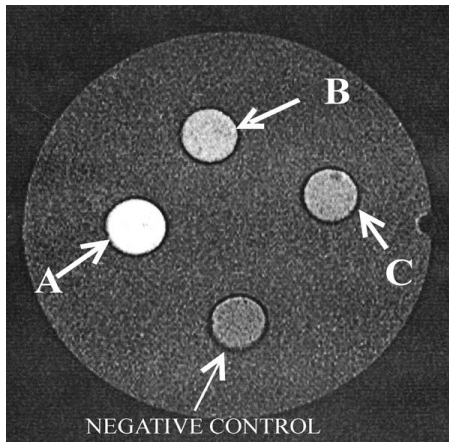
-nanosized systems
-high capacity receptors

The minimum amount of Gd-probe necessary to visualize target Cells depends on its relaxivity

Empirical Equation:

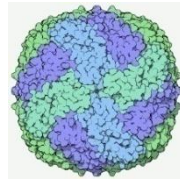
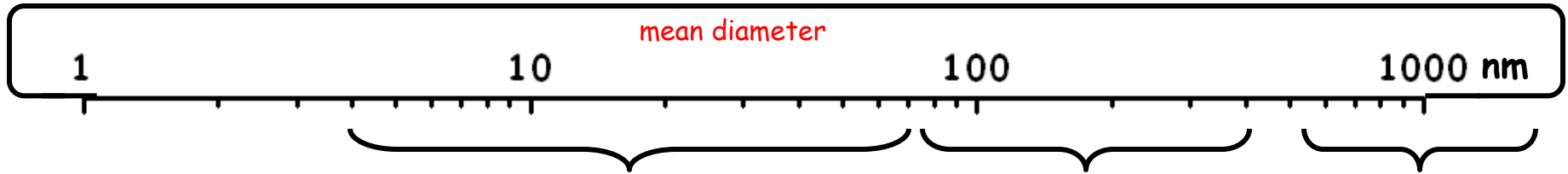


$$r_{1p} \times N^{\circ} \text{ Gd-probe/cell} \geq 10^9$$

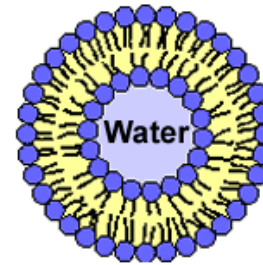


T1-weighted MR image of a phantom formed by three pellets of cells hepatocytes incubated (six hours) in media containing increasing amounts of Gd-DTPAIOPsp, and a negative control. The brightness in the image increases as the amount of internalized Gd(III) complex increases.

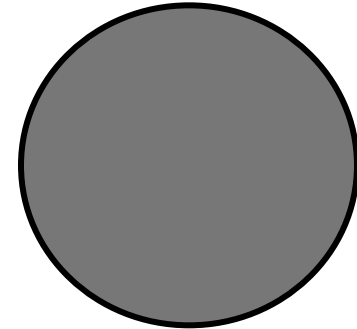
Nanosized carriers for MRI probes



Proteins
(Apoferritin
Virus capsides, ...)

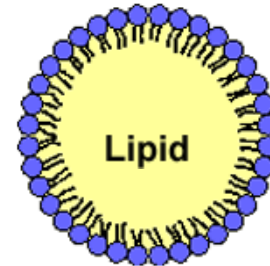
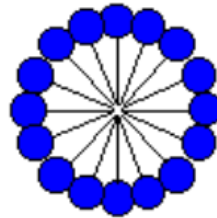


Liposomes

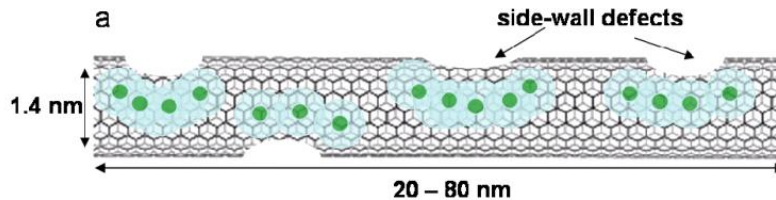


**Micronized
solid particles**

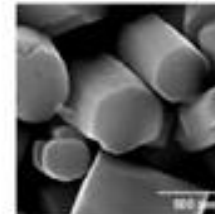
Micelles



**Nanoemulsions
(PFCarbon)**

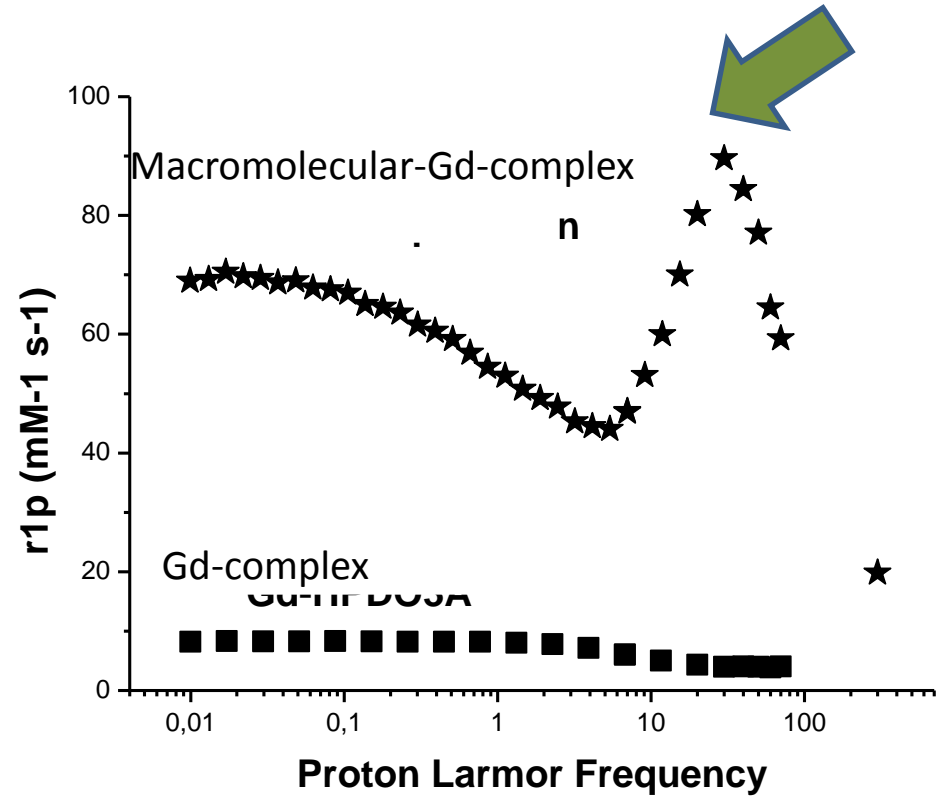
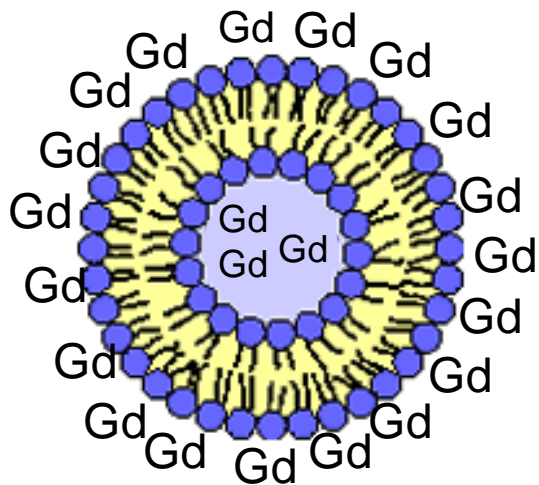


SW Carbon nanotubes



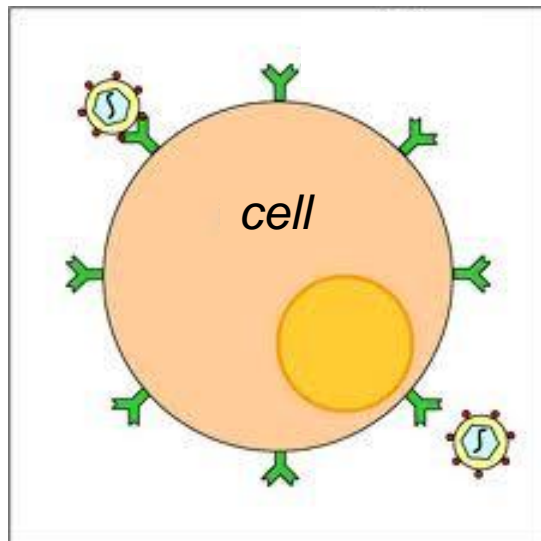
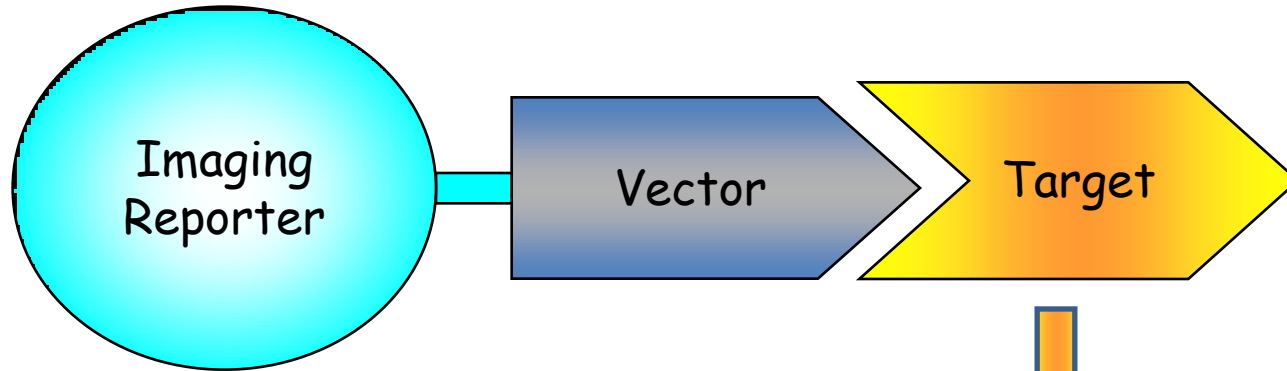
Gd-loaded Zeolites

NMRD profiles of Macromolecular systems



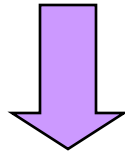
Higher relaxivity at low fields ($0.5 < \text{Tesla} < 1.5$)

"In vivo" molecular imaging



- Overexpressed or downregulated specific cell receptors
- High capacity transporters
- Molecular processes (enzymes)

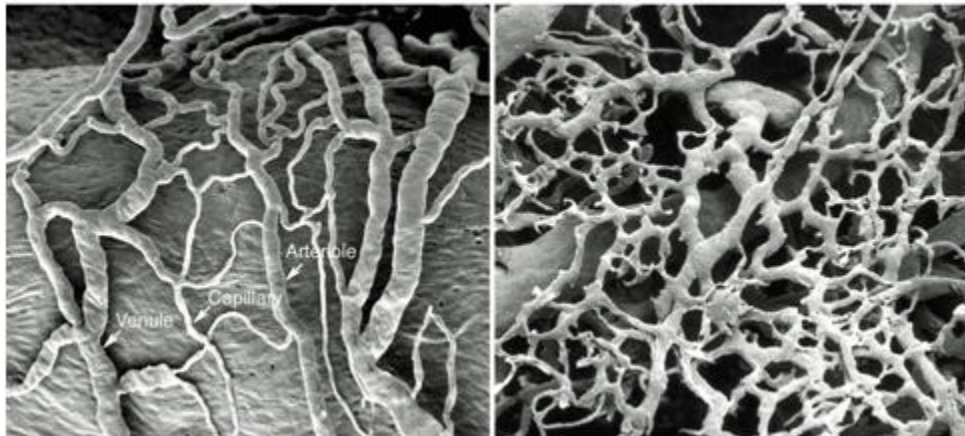
Biological Targets



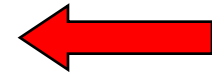
Imaging of Tumor Angiogenesis

- The identification of vessels in tumors is extremely important in oncological research.
- Imaging of angiogenesis have an additional utility in assessing the efficacy of angiogenic inhibitors used in the treatment of cancer.

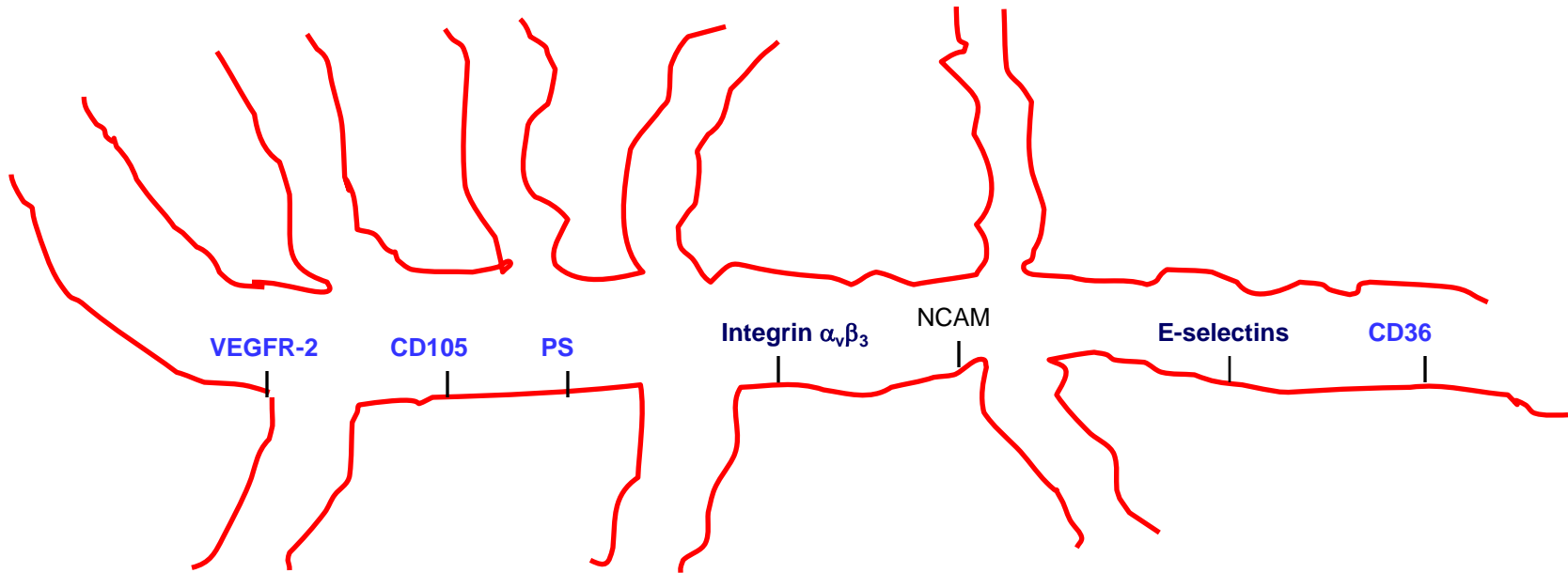
Normal
Microvasculature



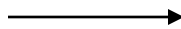
Tumor
Microvasculature



Molecular Biological Markers of Neoangiogenesis



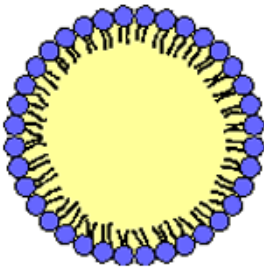
High accessibility of these markers



Large molecular size (nanosize) of Contrast Agents

Targeting of $\alpha_v\beta_3$ Integrins

Theranostics for tumor and plaque angiogenesis with perfluorocarbon nanoemulsions

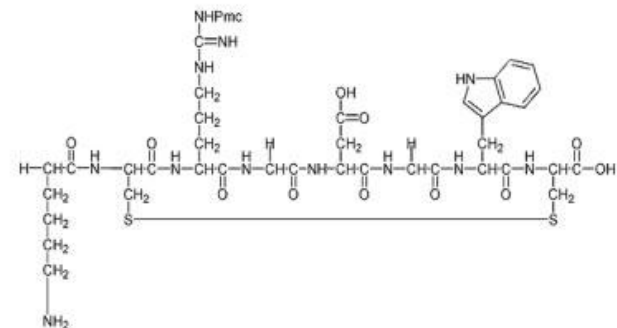


**Nanoemulsions
(PFCarbon)**

Particle size (nm) 280
20% (v/v) perfluorooctylbromide
30 % Gd-DOTA-PE/particle (100000)
 $r1 (s \cdot mM)^{-1} [Gd] 12.5$
 $r1 (s \cdot mM)^{-1} [Particle] 1,6 \times 10^6$
Gd-perfluorocarbon nanoparticles
Covalently coupled to the 3-integrin
peptidomimetic antagonist

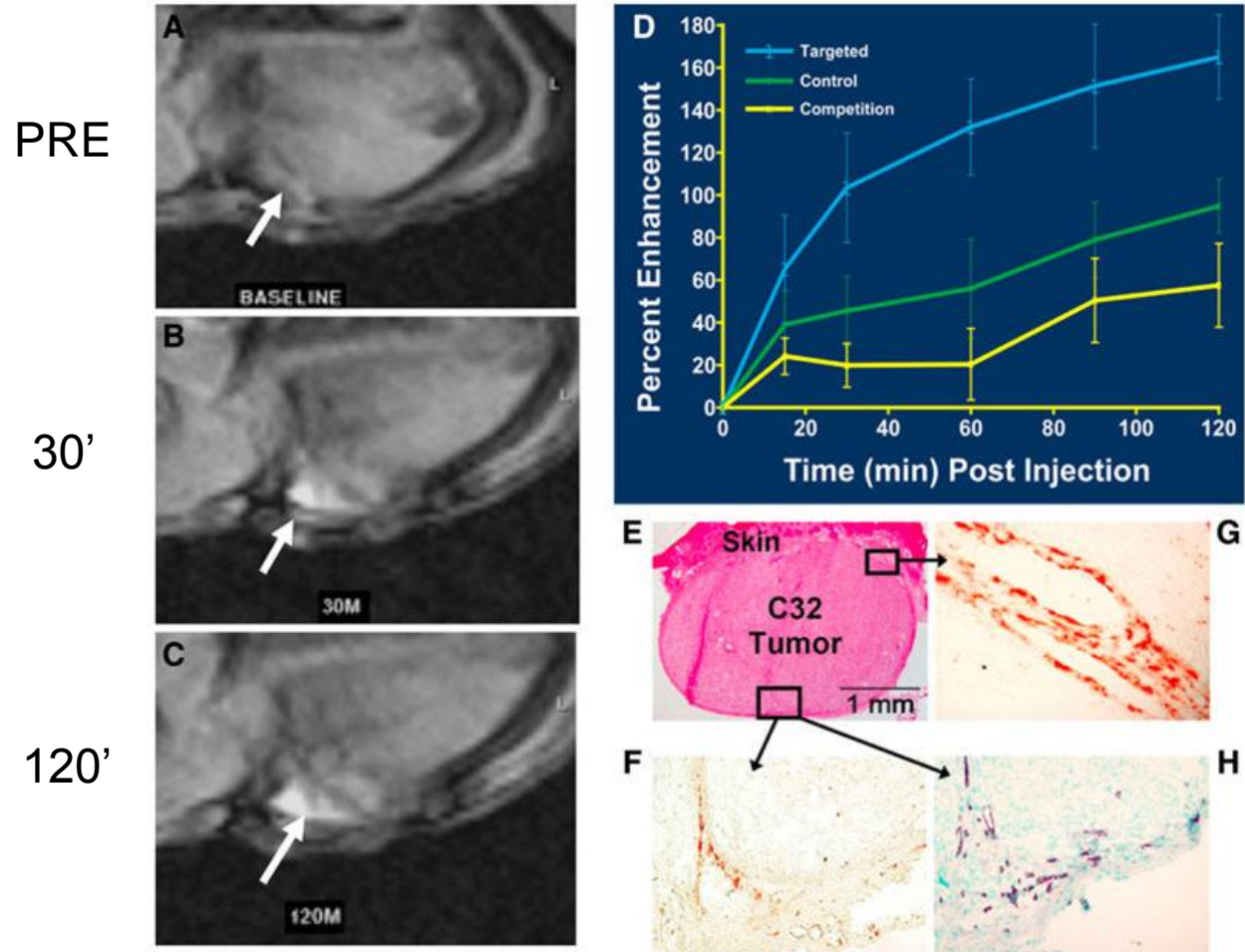
PFC NPs are constrained within the vasculature during the targeting phase, which makes them ideal candidates for specific homing to intravascular biosignatures, such as integrins, selectins, or adhesion molecules.

A $\alpha_5\beta_1$ (RGD) peptide sequence (CRGDGWC)

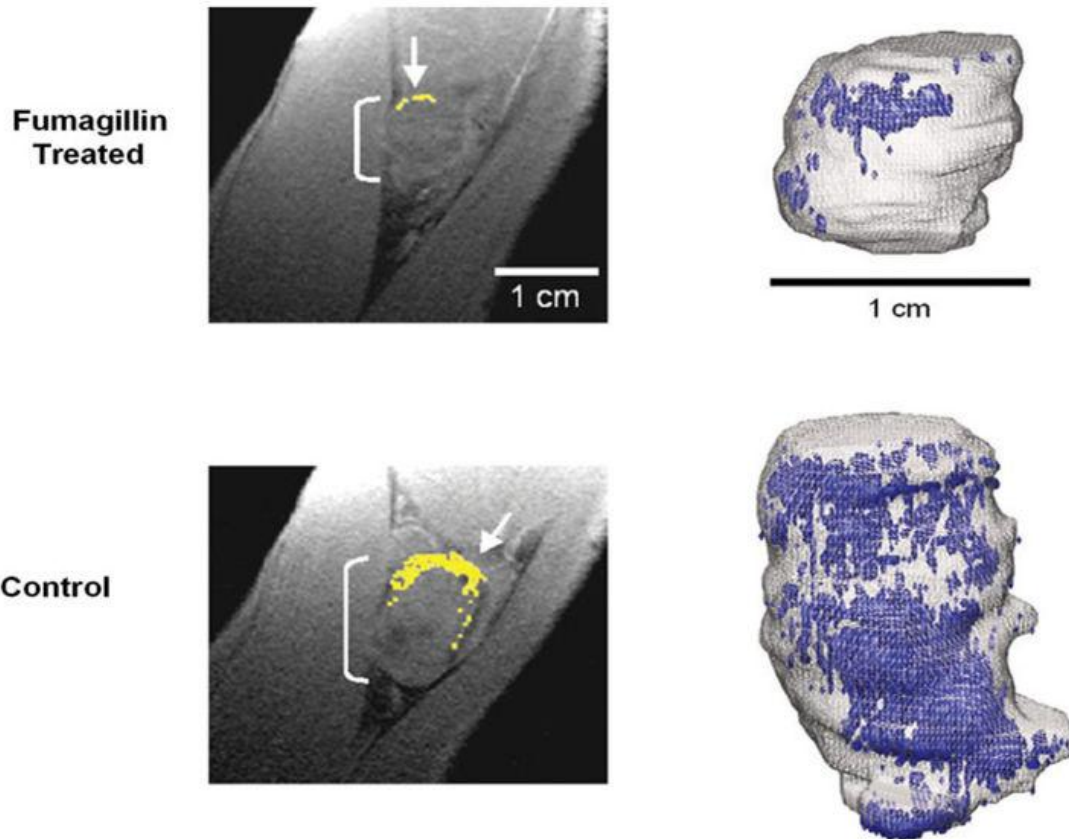


-Lanza et al. Angiogenesis. 2010

T1-weighted MR image (axial view, 1.5 T) of a nude mouse before injection of $\alpha\beta3$ -targeted paramagnetic nanoparticles.



Diminished $\alpha\beta3$ integrin contrast enhancement in T1-weighted, fat-suppressed, 3D gradient echo MR single slice images in rabbits administered $\alpha\beta3$ -targeted fumagillin nanoparticles (top) versus those given $\alpha\beta3$ -targeted nanoparticles without drug (bottom).

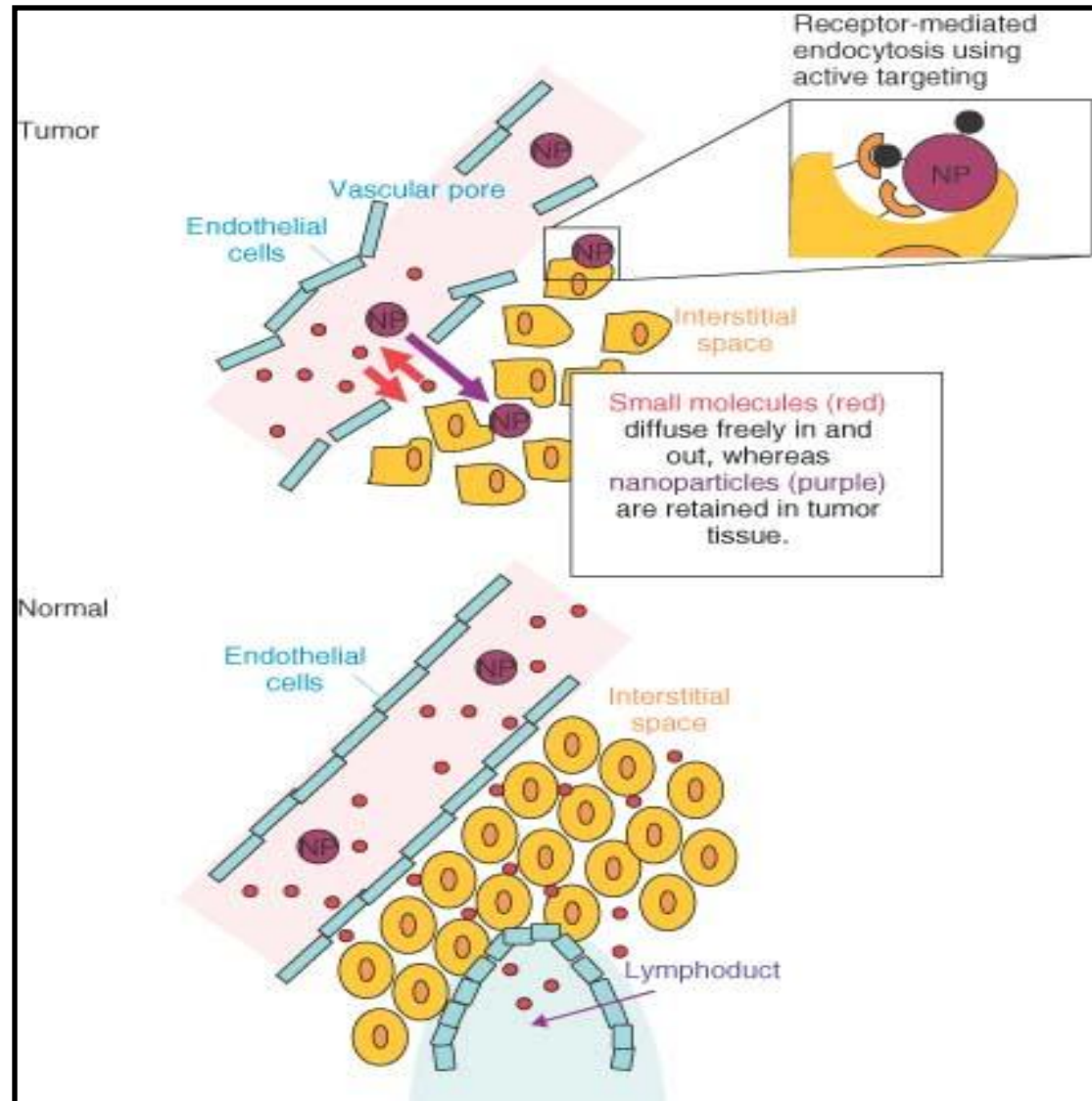


2) Imaging of Tumor Cells

-The visualization of different cell surface targets in solid tumors needs the extravasation of the MR contrast agents.

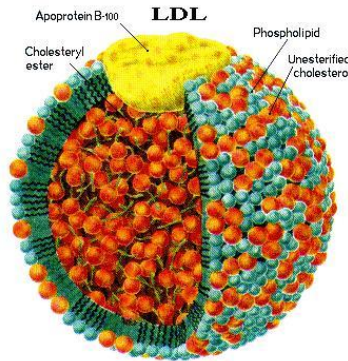
-In solid tumors the vessels formed by the process of angiogenesis show an increase permeability due to large fenestrae (up to 400 nm)

-Normal vasculature endothelium consists of a continuous lining of endothelial cells tightly connected with each other.



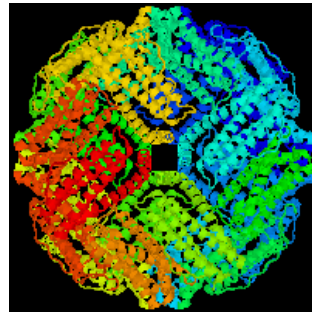
Naturally occurring biological nanocarriers loaded with paramagnetic ions

-Lipoproteins



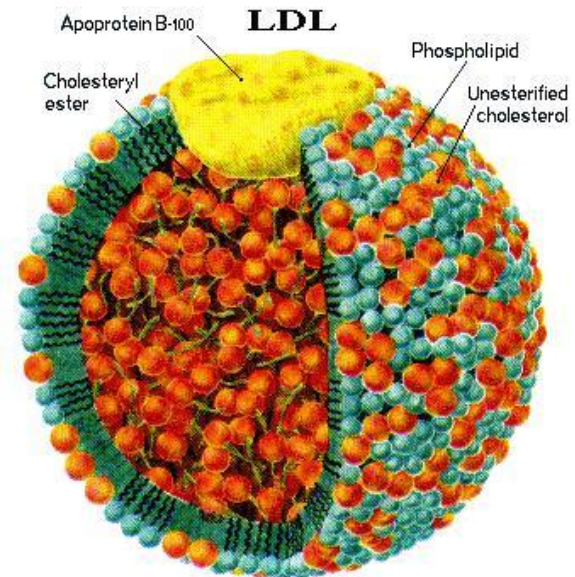
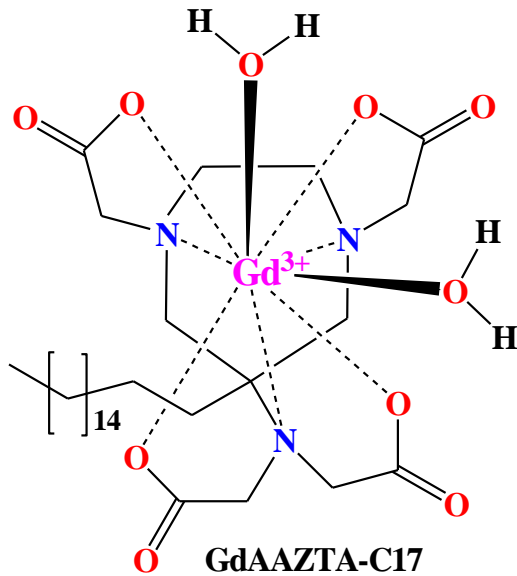
22 nm

-Apoferritin



12 nm

The macromolecular imaging probe: Low Density Lipoproteins (LDL)



$r_{1p} = 10.5 \text{ mM}^{-1}\text{s}^{-1}$ (20 MHz, 298 K)
(free complex $< 0.1 \text{ mM}$)

$q = 2$

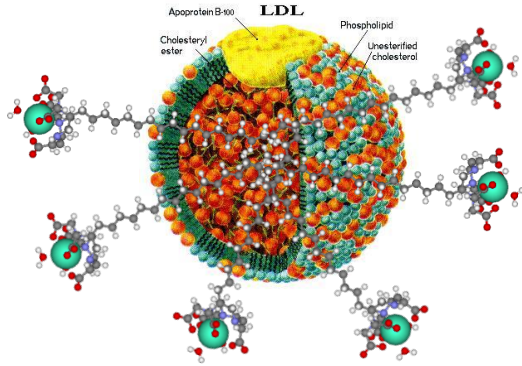
$\tau_M = 70 \text{ ns}$

$\log \beta_{Gd-L} = 19.3$

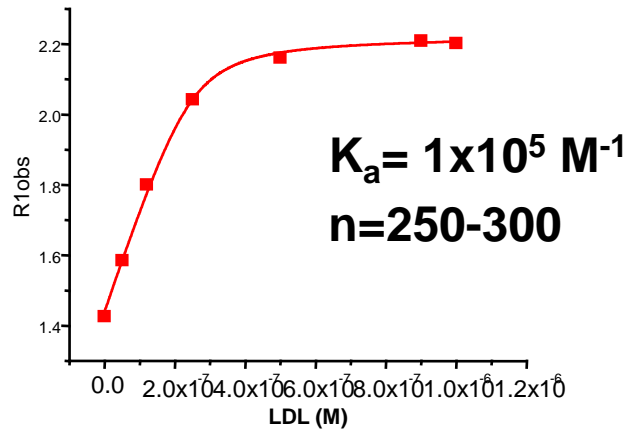
size = 20 nm

S. Aime, et al.; *Inorganic Chemistry* 2004, 43, 7588

S. Baranyai et al.; *Chemistry* 2009 15, 1696

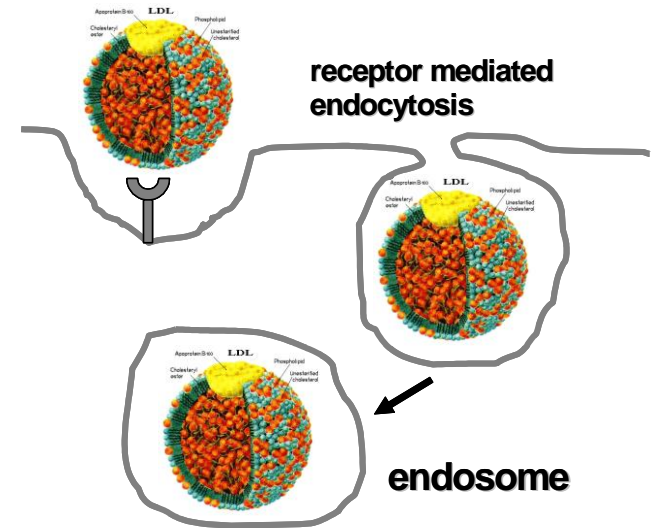


**Gd-AAZTA-C17/LDL
macromolecular adduct**

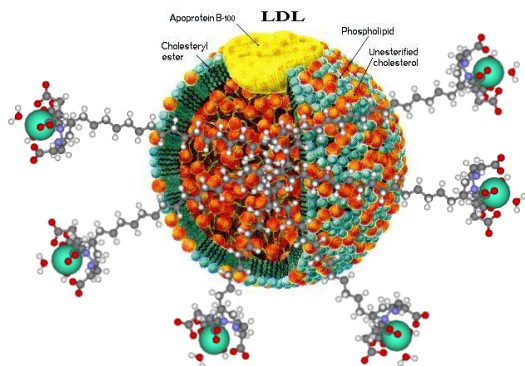


Size:
Native LDL = 21 2 nm
Gd-AAZTA-C17/LDL = 22 2 nm

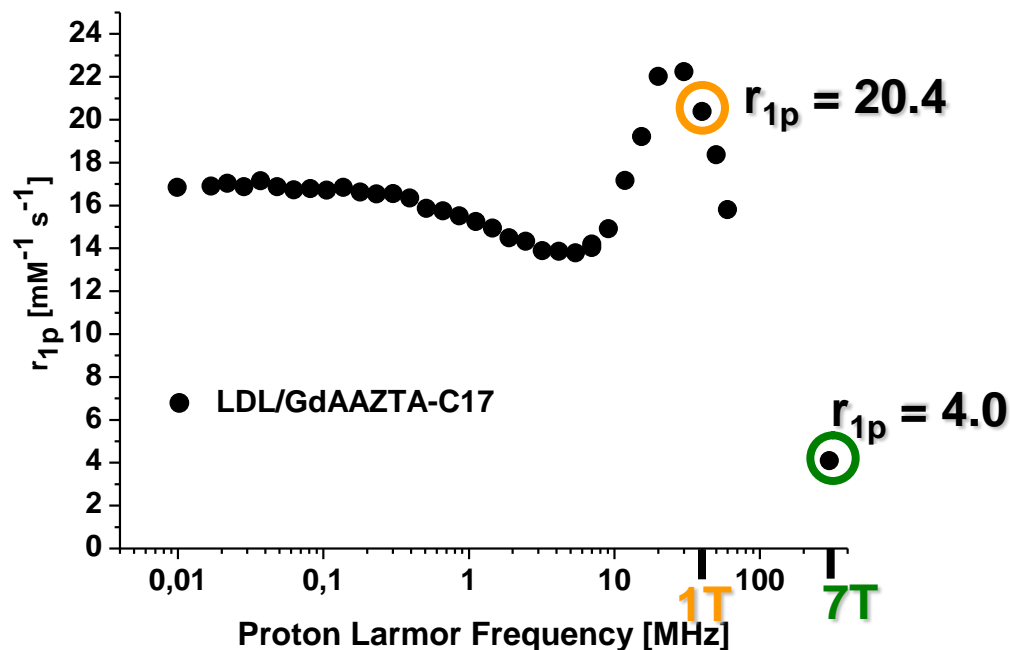
altered LDL Receptors levels are found in tumor cells over-expressing LDL-R to supply the high cholesterol demand



NMRD PROFILE OF Gd-AAZTA-C17/LDL



**Gd-AAZTA-C17/LDL
macromolecular adduct**

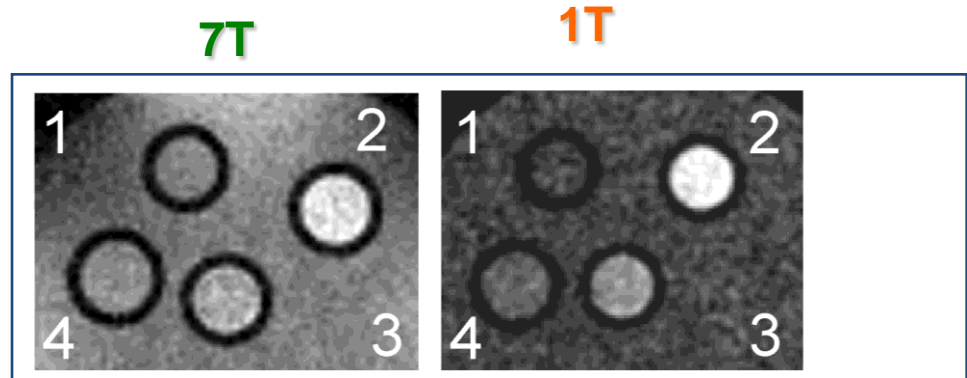
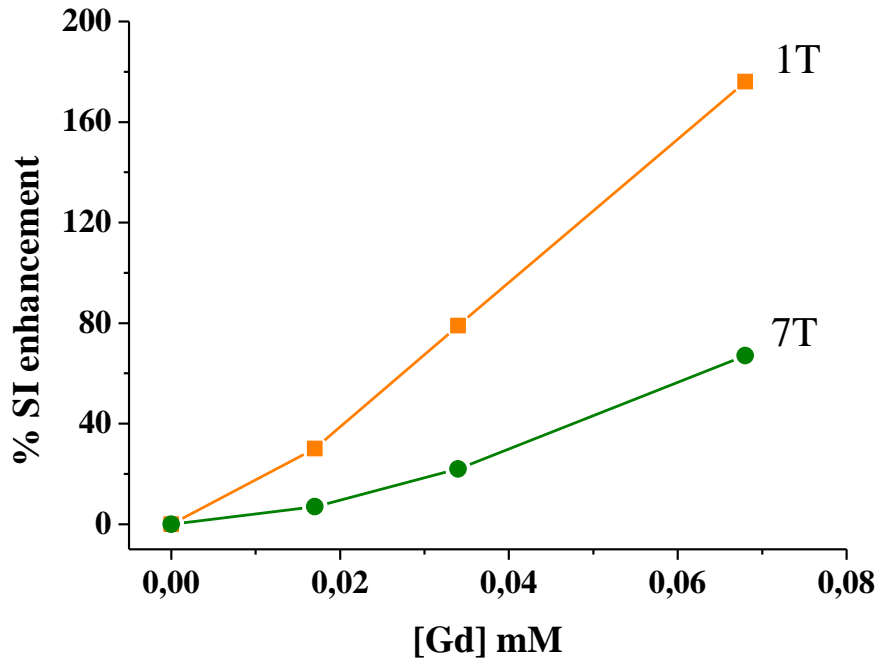


r_{1p} of the **Gd-AAZTA-C17/LDL** macromolecular adduct
 $6150 \text{ mM}^{-1} \text{ s}^{-1}$ at 1T and $1200 \text{ mM}^{-1} \text{ s}^{-1}$ at 7T

IN VITRO RESULTS ON B16-F10 melanoma

cell culture – uptake

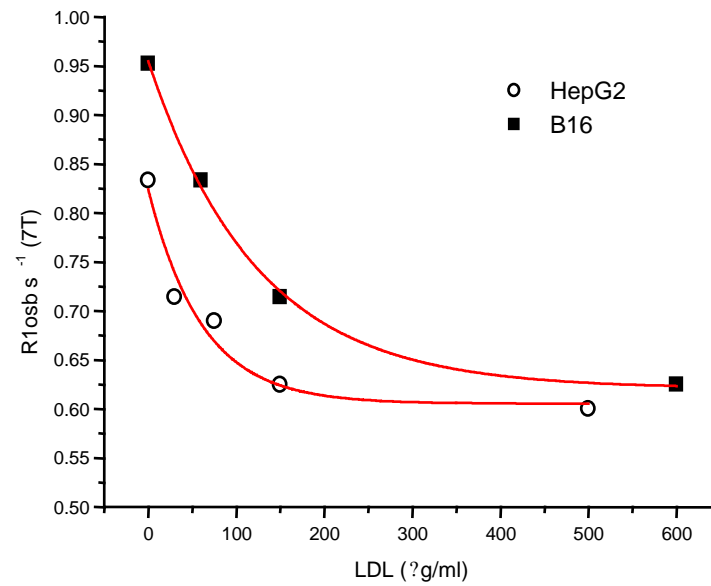
- incubation at 37 °C – 24h with 10% LPDS
- incubation with 60 µg/ml adduct for 16h
- washed 3x PBS, detached trypsin/EDTA
 - transferred into glass capillaries
 - Gd content determined by ICP-MS
 - cell viability assessed by colorimetric assay (WST-1)



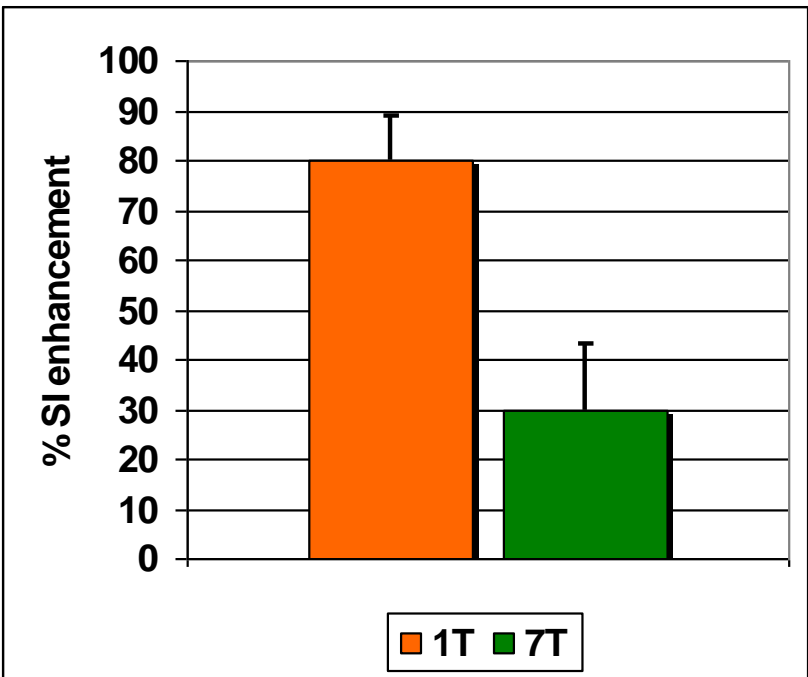
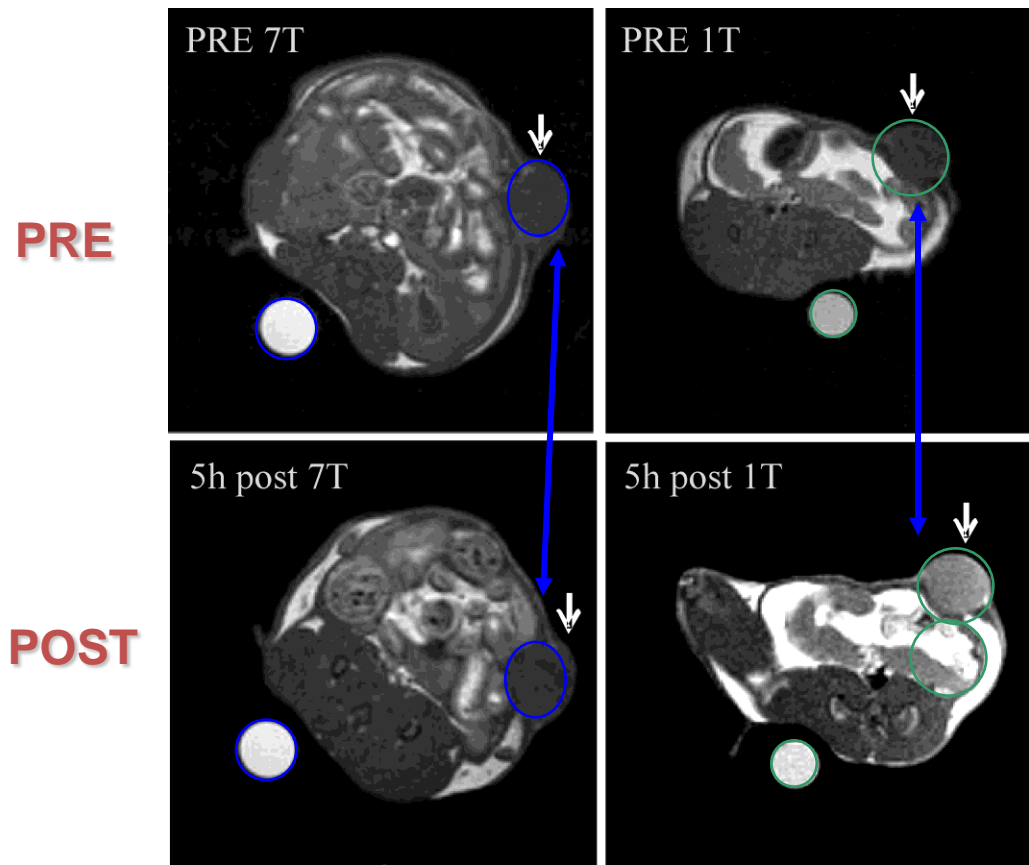
T_1 -weighted spin echo image, recorded at 1 and 7T, of an agar phantom containing B16 cells labeled with Gd-AAZTAC17/LDL adduct

| | cells/µl | [Gd] mM | Measured % SI (1T) | Measured % SI (7T) |
|---|----------|---------|--------------------|--------------------|
| 2 | 20000 | 0.068 | 176 | 67 |
| 3 | 10000 | 0.034 | 79 | 22 |
| 4 | 5000 | 0.017 | 30 | 7 |
| 1 | ctrl | 0 | 0 | 0 |

Competition study with unlabeled LDL



T₁-weighted multislice spin echo MR images of C57BL/6 mice grafted subcutaneously with B16 melanoma cells



n=3

PRE

POST

7T Bruker

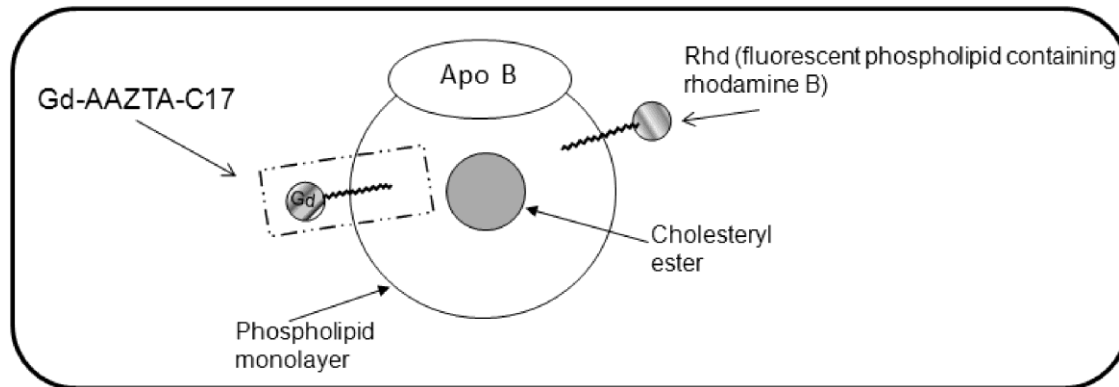


1T, Aspect

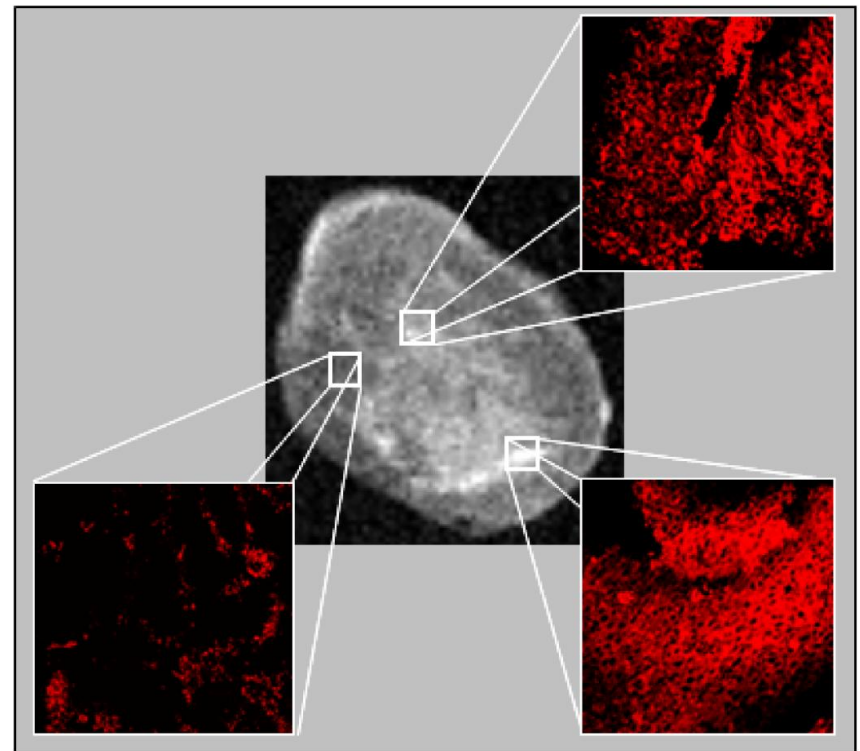


Images were obtained before (PRE) and 5 hours after (POST) the administration of Gd-AAZTAC17/LDL 0.06mmol/Kg
 7T: TR/TE = 250/4 ms; in plane resolution: 78 μm
 1T: TR/TE= 250/7 ms; in plane resolution: 78 μm

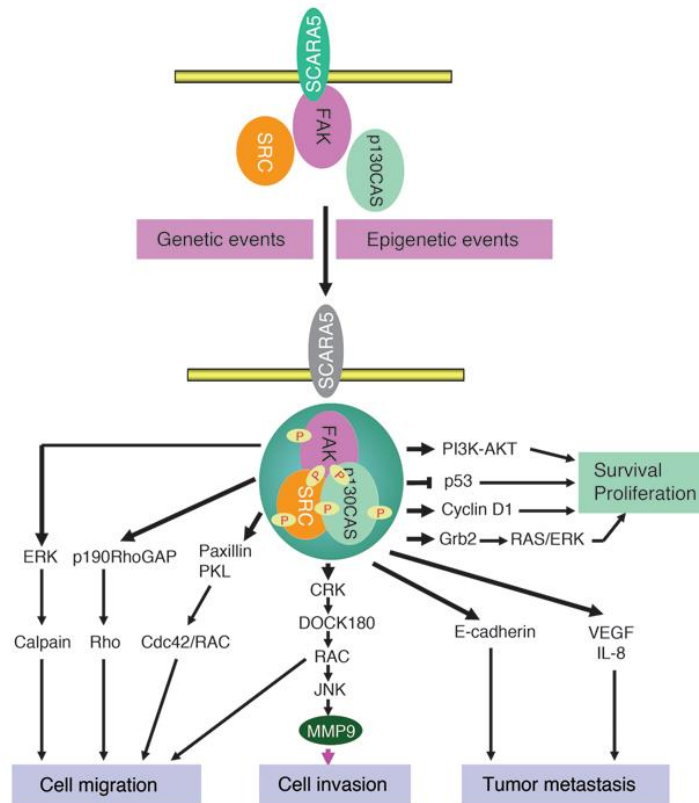
Ex vivo MRI and confocal analysis after administration of Gd-AAZTAC17/Rhd/LDL



The hyperintense areas detected in the MR images correspond to areas of greater Rhd fluorescence whereas in the other regions the Rhd signal is weak or completely absent.



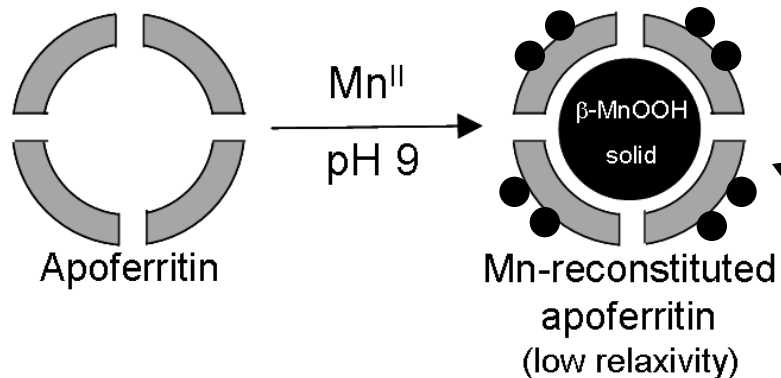
Genetic and Epigenetic silencing of Scara5 may contribute to human Hepatocellular carcinoma by activating FAK signaling



Scara5 Is a Ferritin Receptor Mediating Non-Transferrin Iron Delivery

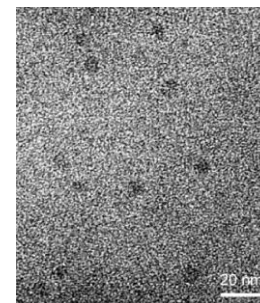
Can apoferritin loaded with paramagnetic ions act as MRI gene expression reporter ?

Reconstitution of Manganese Oxide Cores in Horse Spleen Apoferritin

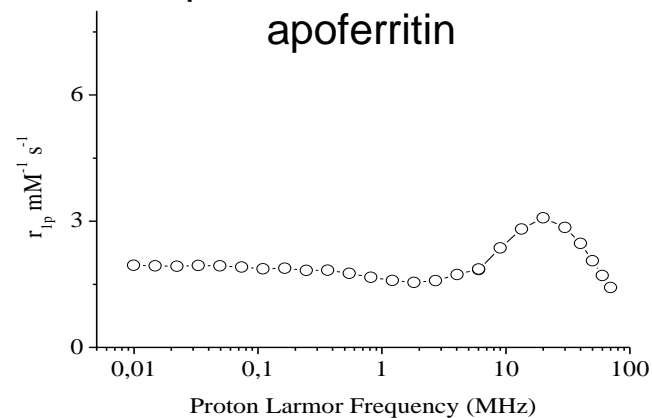


- Mn(II) 5 unpaired electrons
- $r_{1p}(20\text{MHz})=8.0 \text{ mM}^{-1}\text{s}^{-1}$
- $\text{LD}_{50} = 0.22 \text{ mmol/Kg}$
- Essential Ion

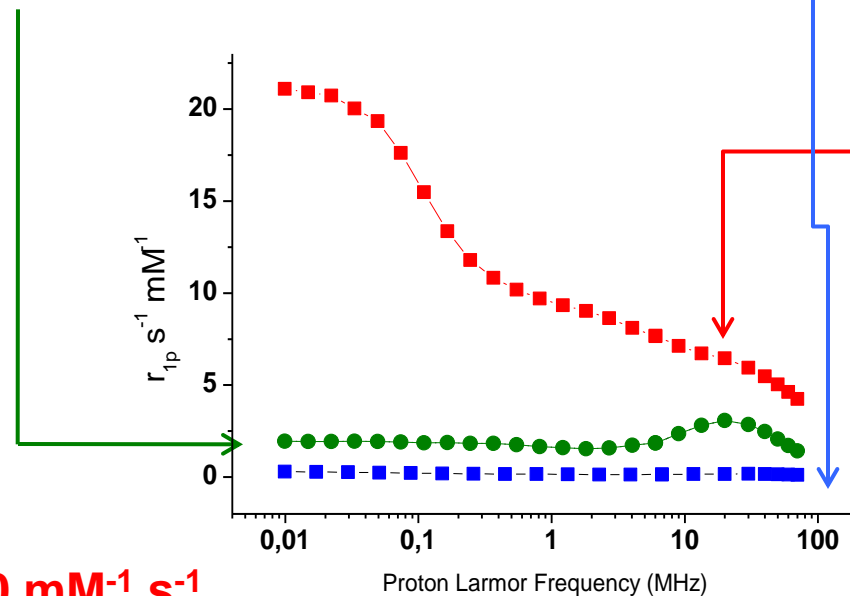
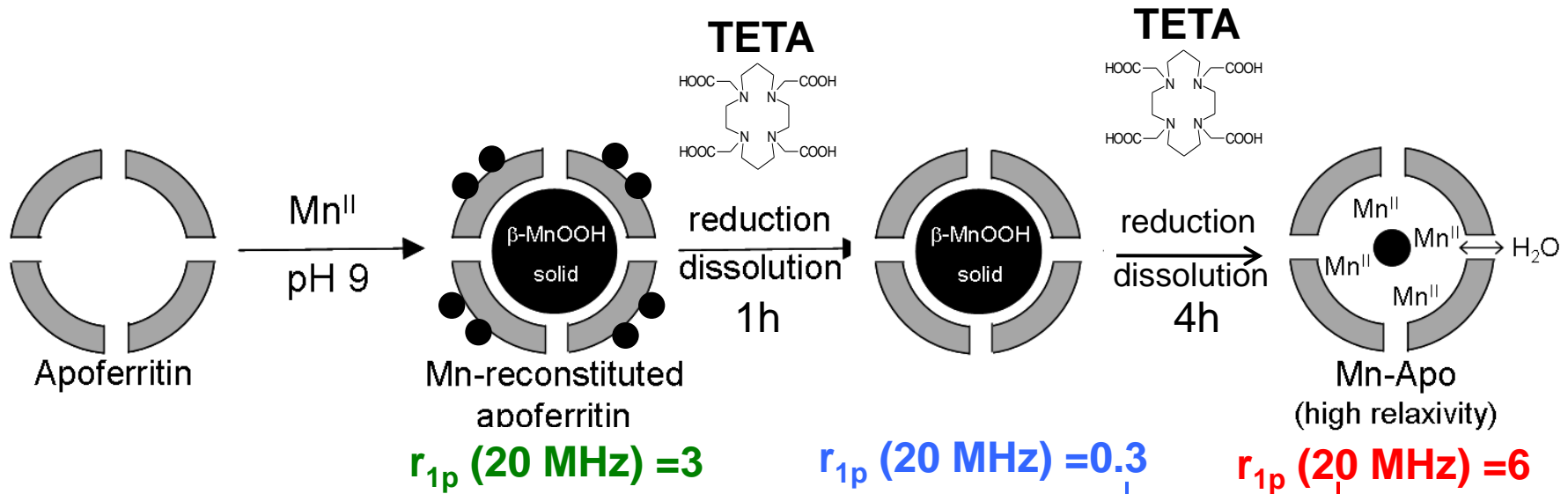
TEM image of Mn reconstituted apoferritin



NMRD profile of Mn reconstituted apoferritin

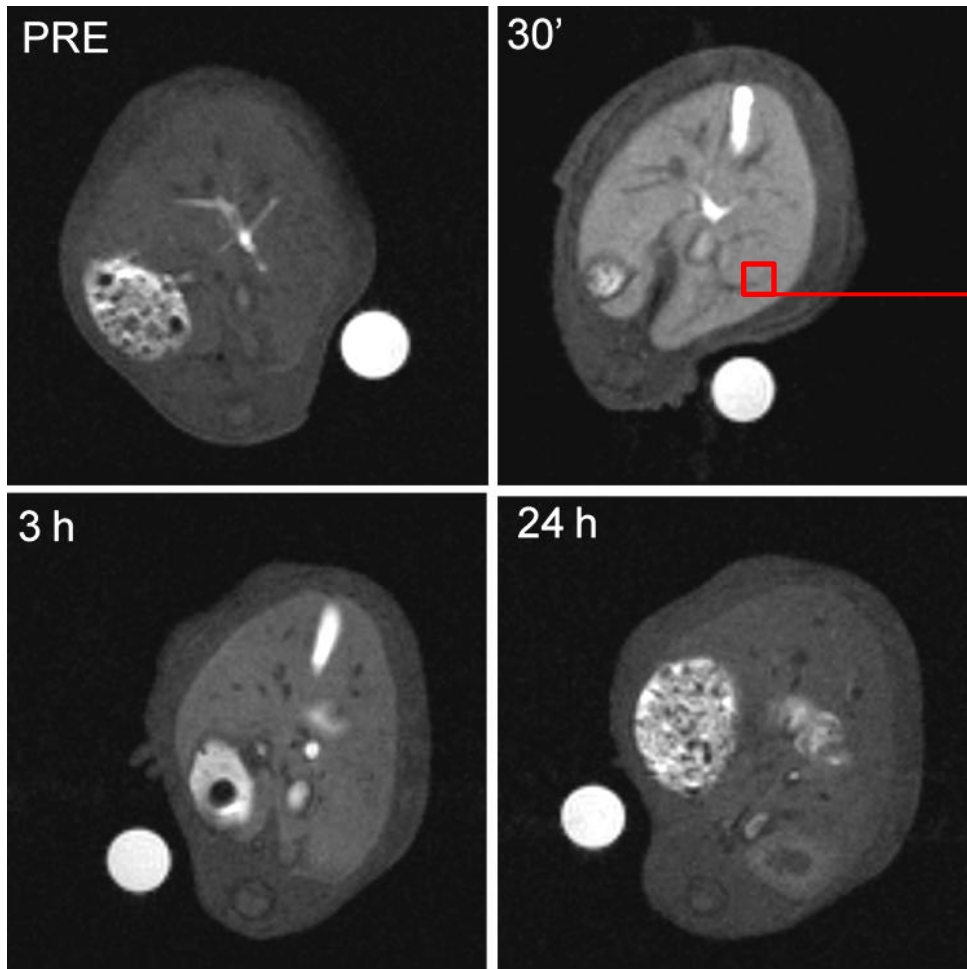


Mn-loaded Apoferritin as a MR Molecular Imaging probe

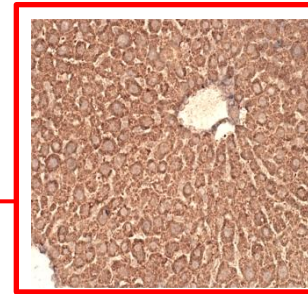


$r_{1p} \text{Mn-APO} = 6500 \pm 500 \text{ mM}^{-1} \text{ s}^{-1}$

T₁-weighted MR images of C57BL/6 mice (liver region) acquired before and 30', 3 and 24h after the administration of Mn-Apo at a Mn dose of 0.01 mmol/kg.

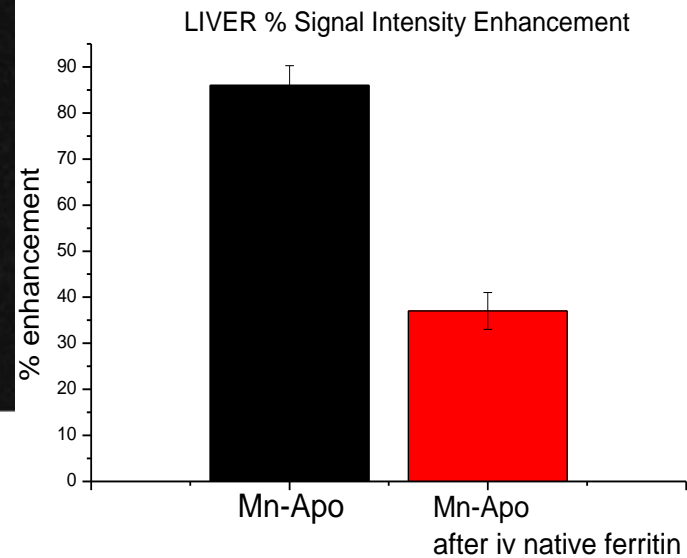


Immunohistochemistry;



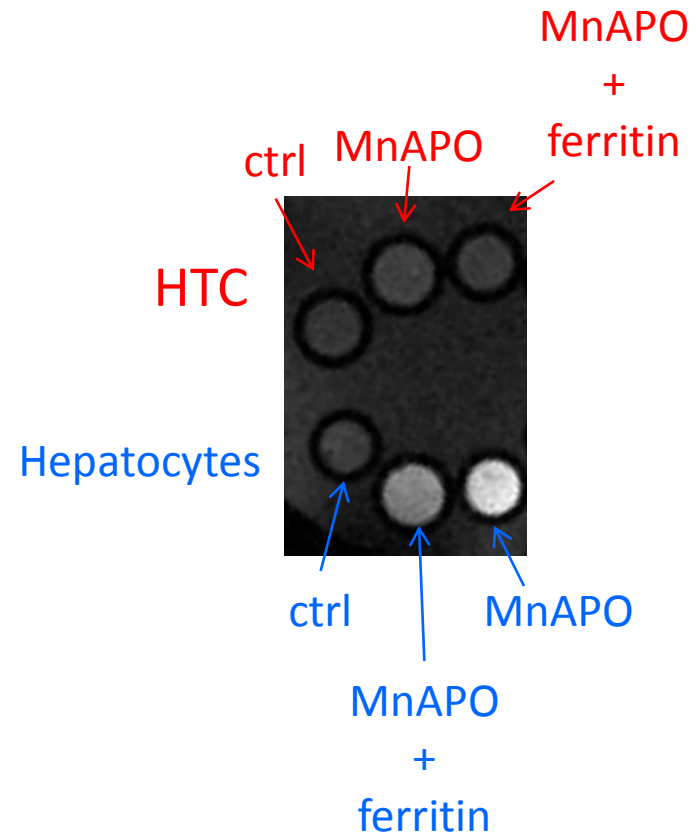
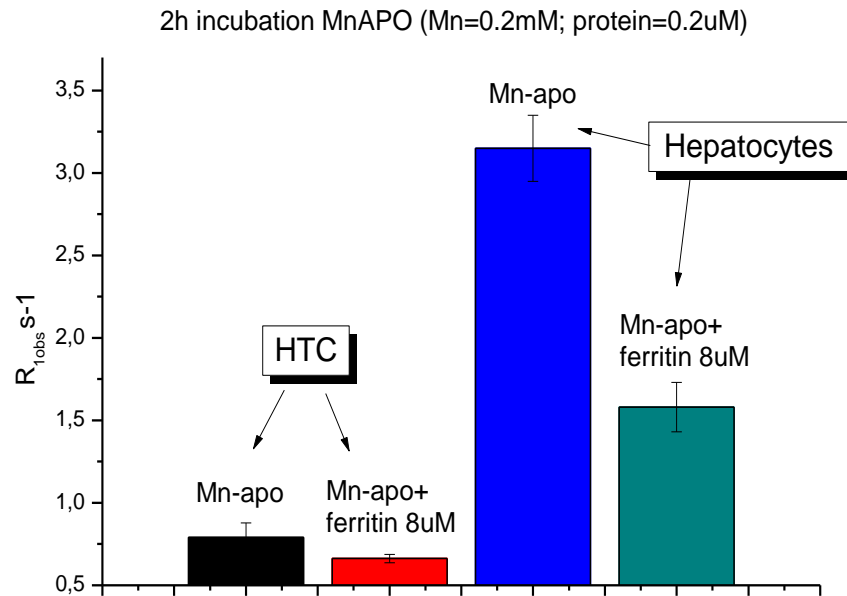
Plasma-membrane positive reaction for SCARA5 in wt mouse liver

Competition with native ferritin



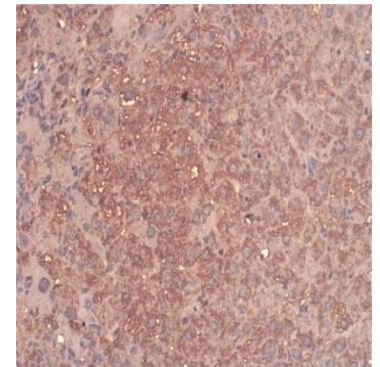
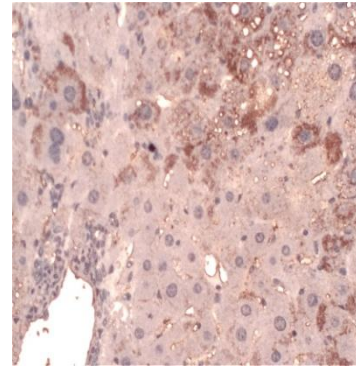
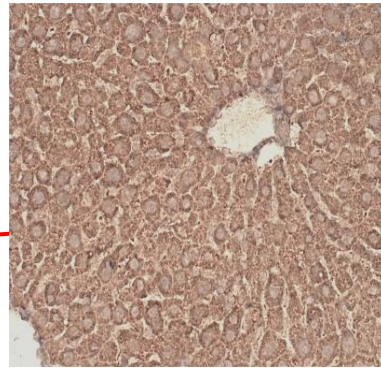
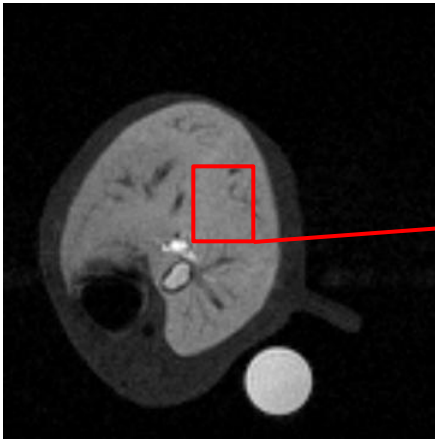
Uptake of Mn-APOFERRITIN by rat Hepatoma (HTC) and healthy rat hepatocytes

(2h incubation 0.2mM Mn, 0.2 uM protein)

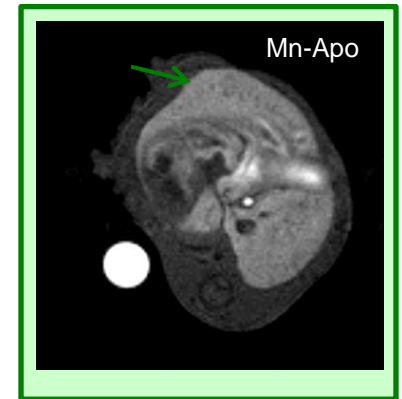
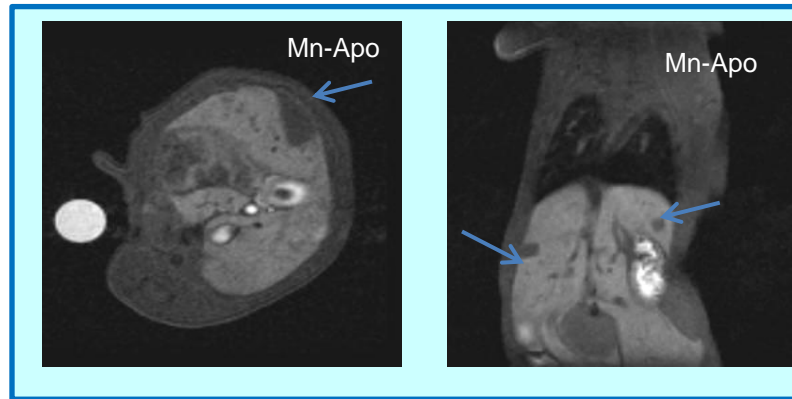


T₁-weighted MR images (liver region) of C57BL/6 w.t HBV transgenic mice at a Mn dose of 0.01 mmol/kg.

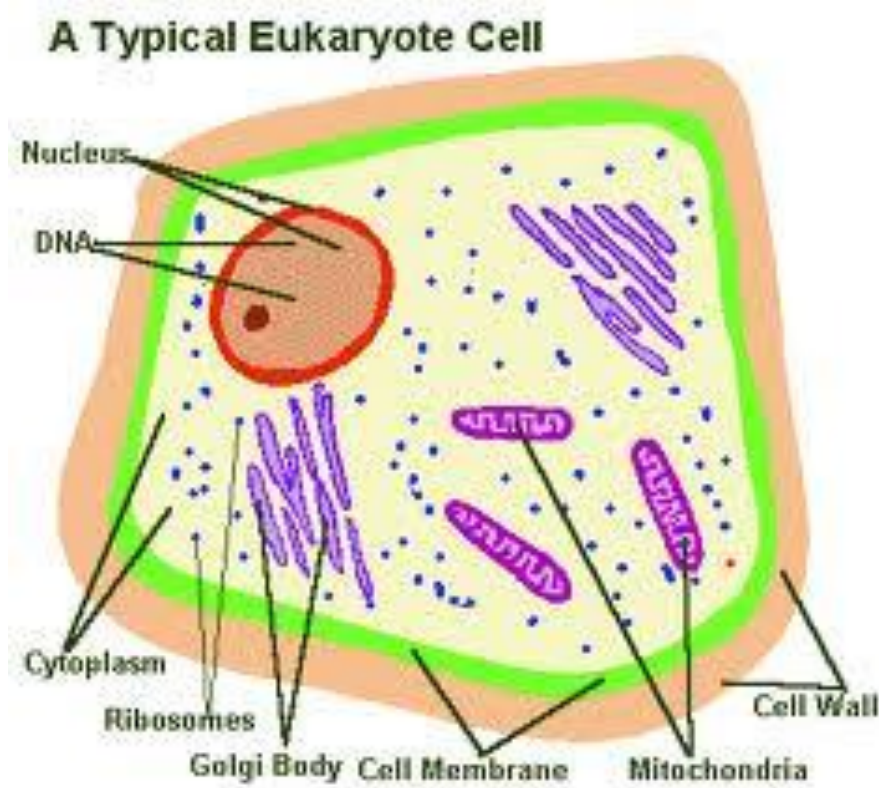
WT



HBV-Tg-mice
13 months old



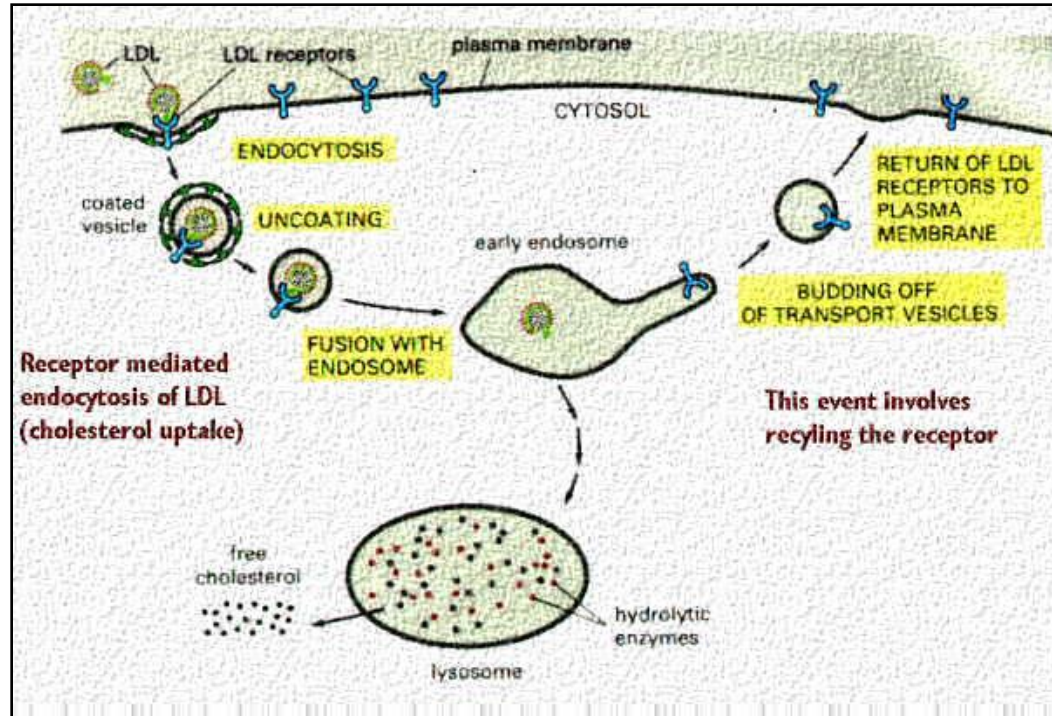
Localization of MRI Imaging Probes



Extracellular → cell membrane

Intracellular → cytoplasm or endosomes/lysosomes

Receptor Mediated Endocytoses



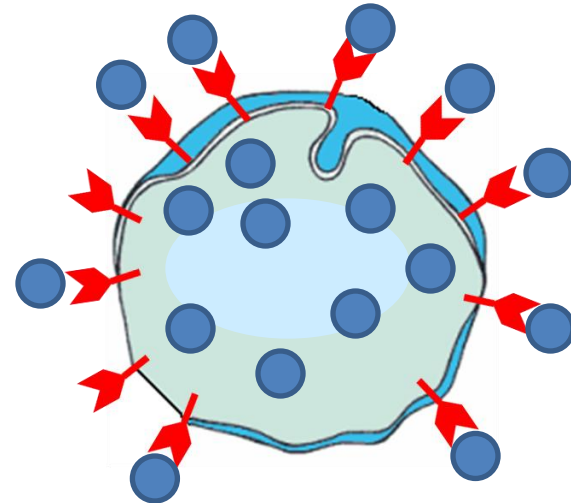
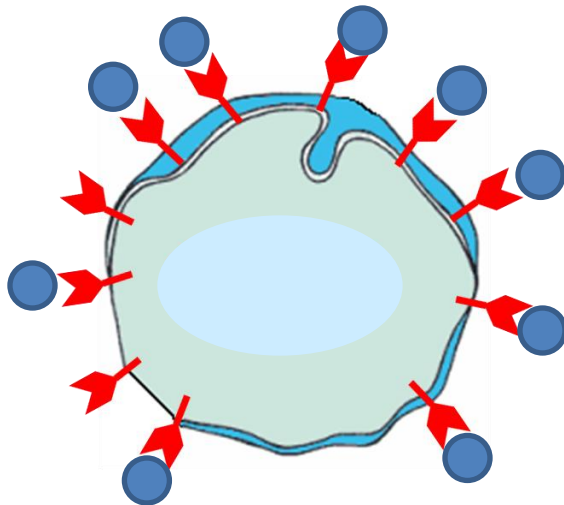
Receptor recycling time → 10' LDL receptor
12 h Folate receptor

Number of Gd units internalized



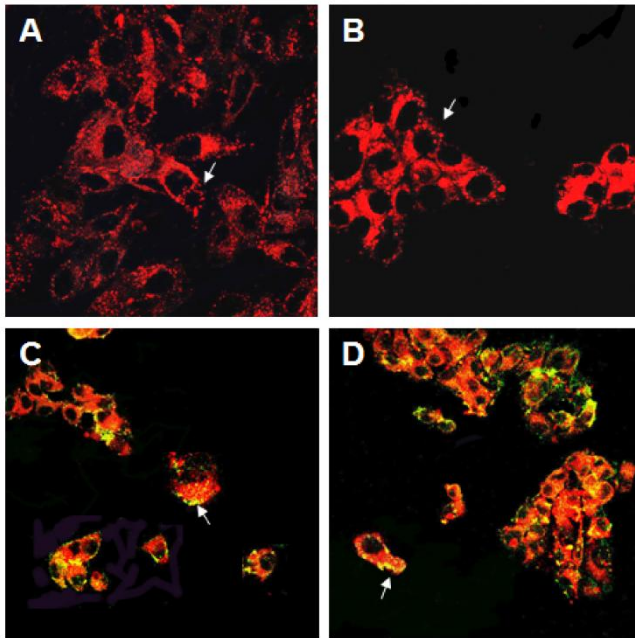
number of receptors x receptor
recycling number

● = Imaging
probe



How can you detect Imaging Probe localization?

-Confocal Microscopy after labeling with a fluorescent dye

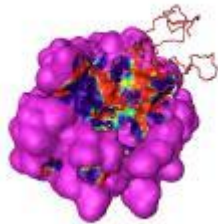


Confocal microscopy indicates that fluorescently labeled nLDL particles are taken into the cell by LDL receptors and are found together in the cell's lysosomes,

Cells detaching methods



Trypsin/EDTA



- Intracellular Imaging Probe

Scraper



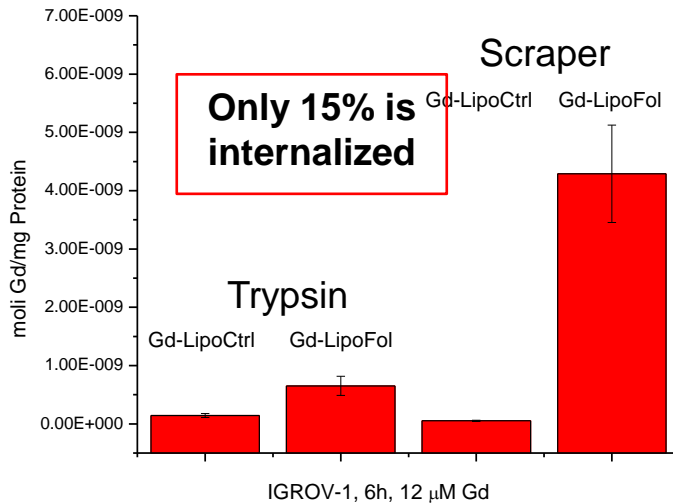
Total cell associated Imaging Probe
Extra- + Intra- cellular

Extra- + Intra- cellular

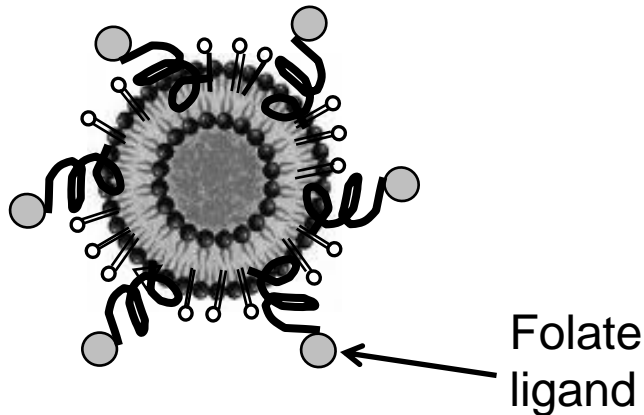
MRI + ICP-MS (Gd determination)

ICP-MS (Gd determination)

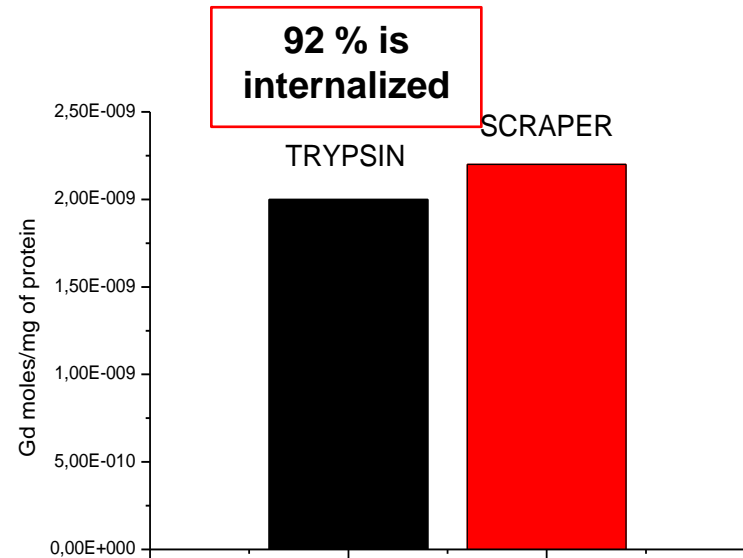
IGROV-1 cells overexpressing Folate Receptors



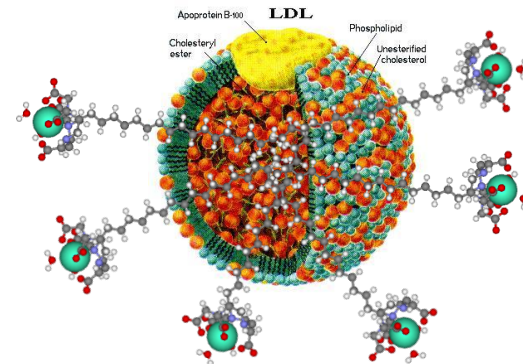
Imaging Probe:
FR targeted Gd-Loaded Liposome



B16-F10 cells overexpressing LDL Receptors



Imaging Probe:
Gd-loaded/LDL



Relaxivity dependence on intracellular localization of the Gd-based imaging probe.

Pinocytoses

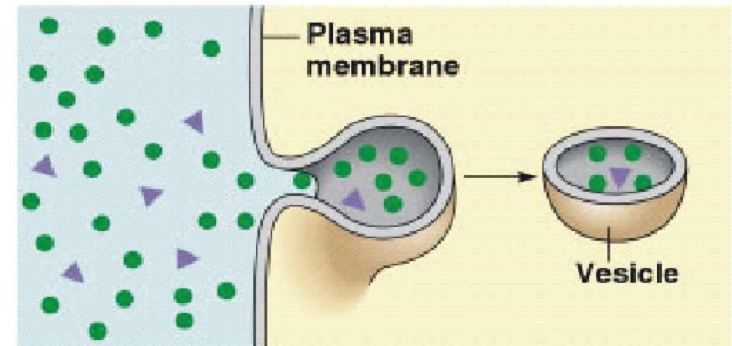
Electroporation

Pinocytosis

Labelling of Cells in culture
(stem cells, leucocytes, etc...)



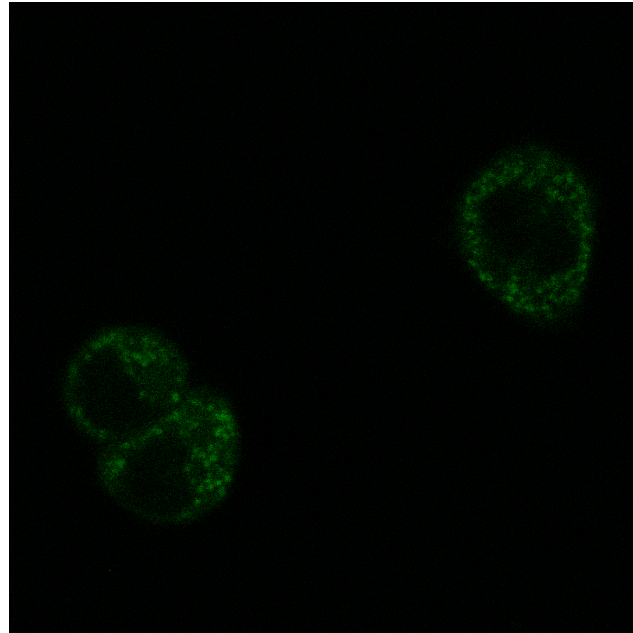
Incubation at high concentration
of well-tolerated Gd-chelates



(b) Pinocytosis

Fluorescence Microscopy of EPCs labelled with **Eu-HPDO3A**

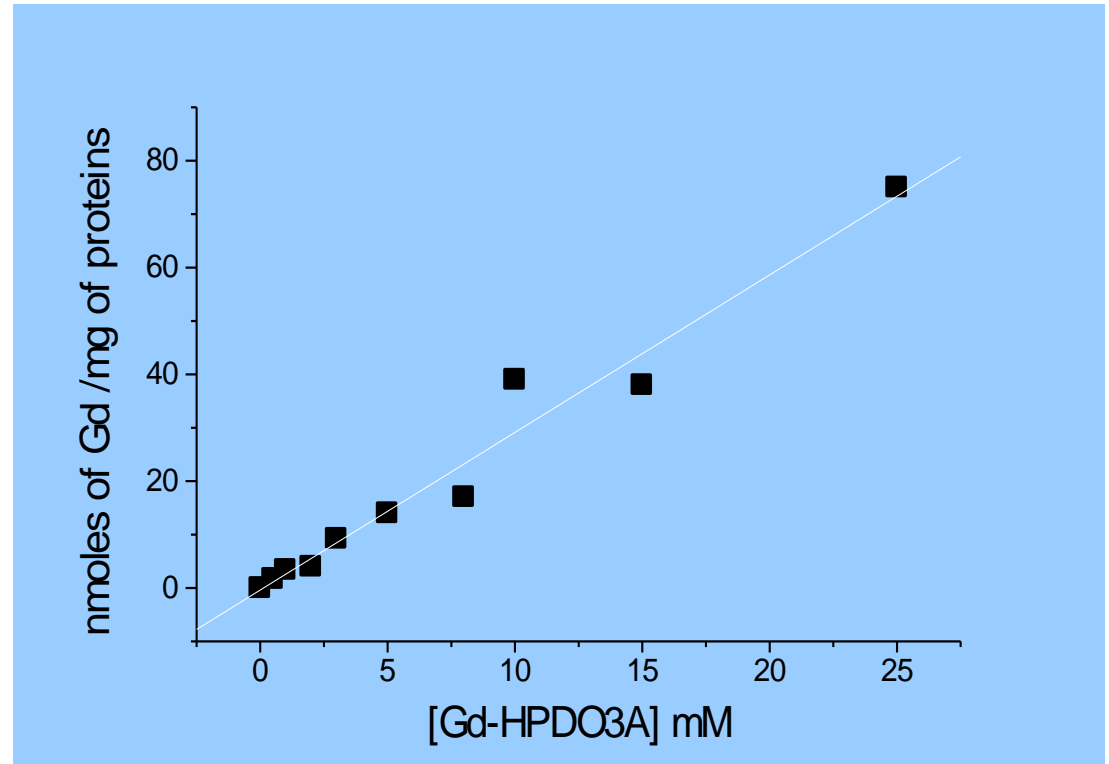
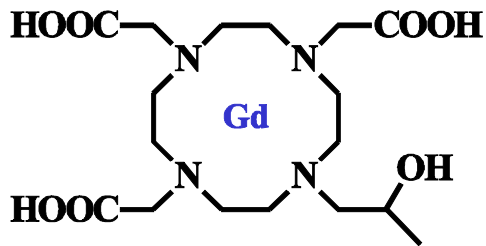
Eu-HPDO3A and Gd-HPDO3A have almost identical physico-chemical properties and therefore the same biodistribution



Incubation time: 16h at 37°C (in the presence of 50 mM of Eu-HPDO3A)

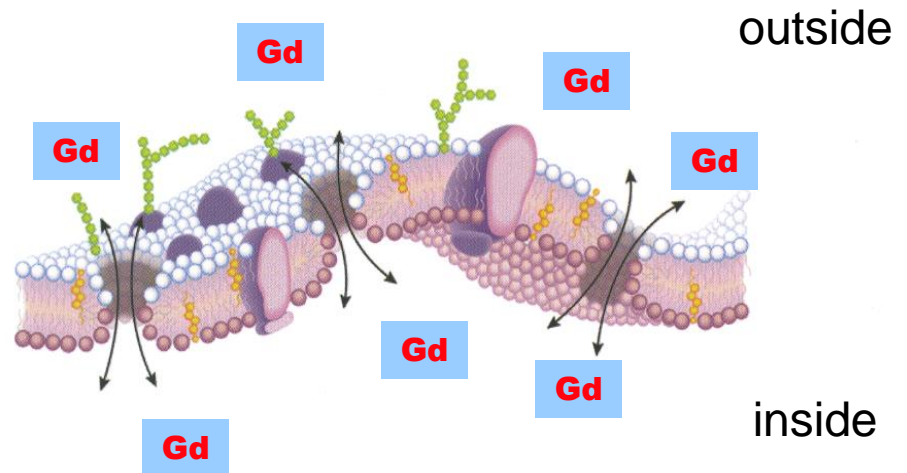
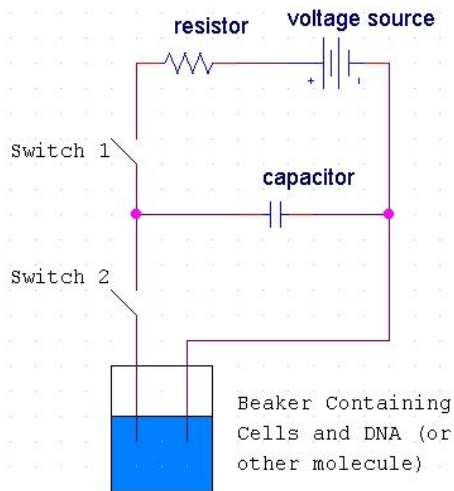
**FLUORESCENT Eu-HPDO3A ACCUMULATES
IN ENDOSOMES AROUND NUCLEOUS**

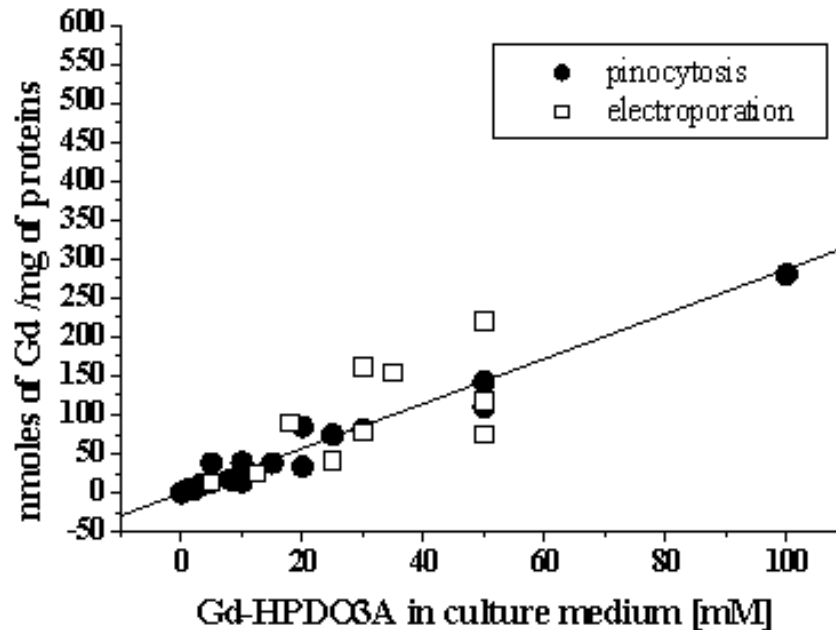
Labelling of HTC cells with **Gd-HPDO3A** via endosomic entrapment



Electroporation

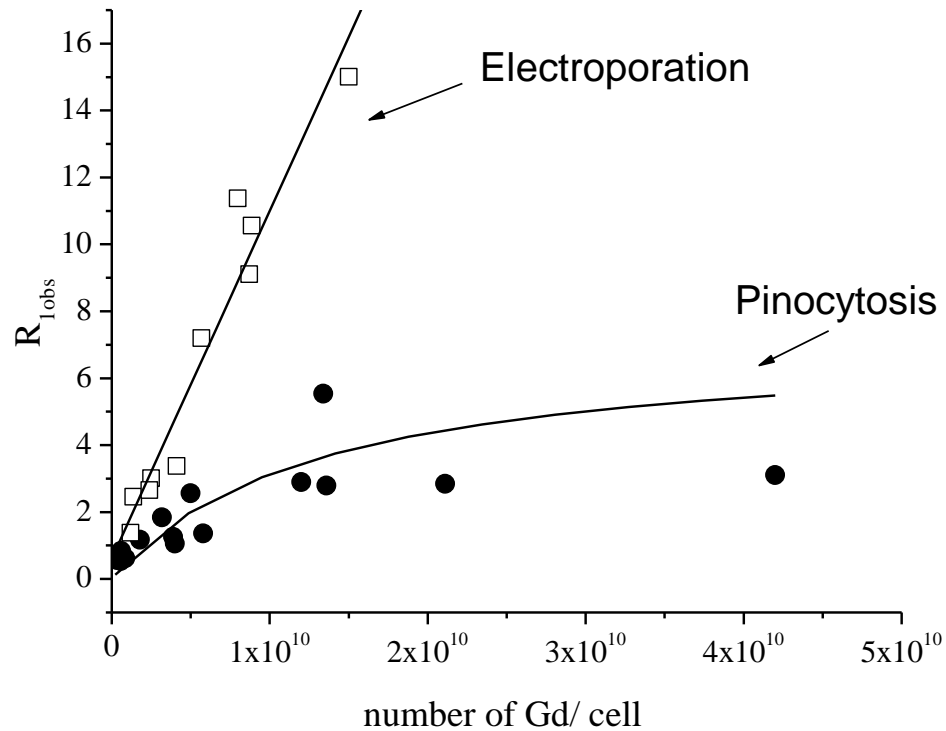
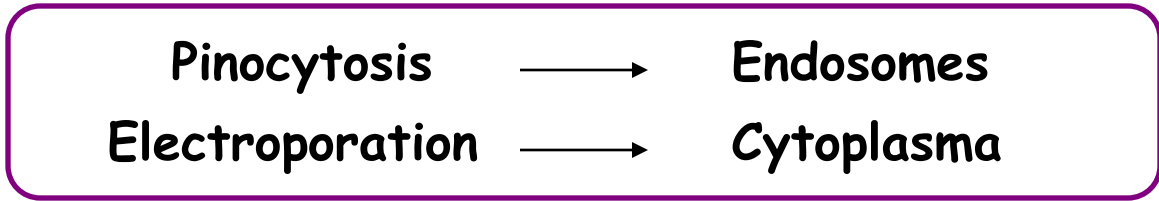
The imaging probe enters the cell through transient hydrophilic pores formed upon application of a suitable electric pulse





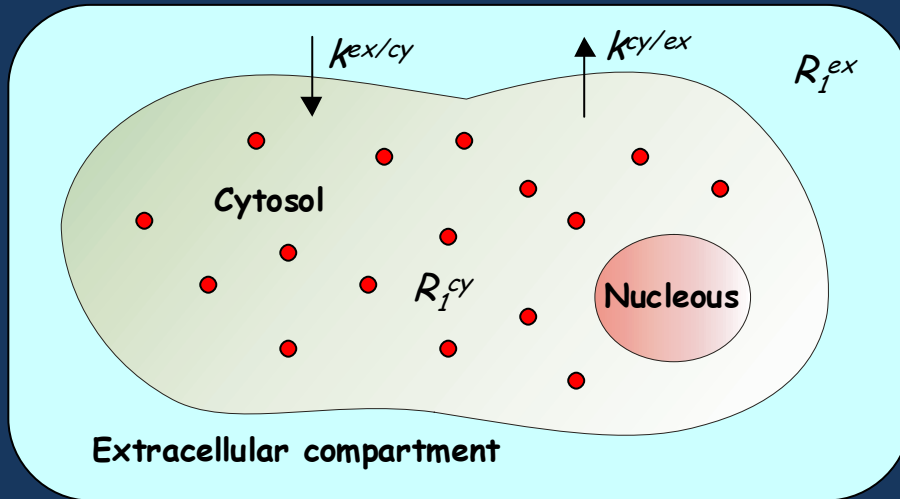
The internalization of Gd-HPDO3A into HTC cells. About 5-6 million cells were (●) incubated at 37 C for 16 hours (pinocytosis) or (□) electroporated applying a pulse of 0.2 kV. Both electroporation and pinocytosis were performed in the presence of increasing concentrations of contrast agent (Gd-HPDO3A).

How the intracellular localisation affects the attainable relaxivity



How the intracellular localisation affects the attainable relaxivity ?

● Gd(III)complex

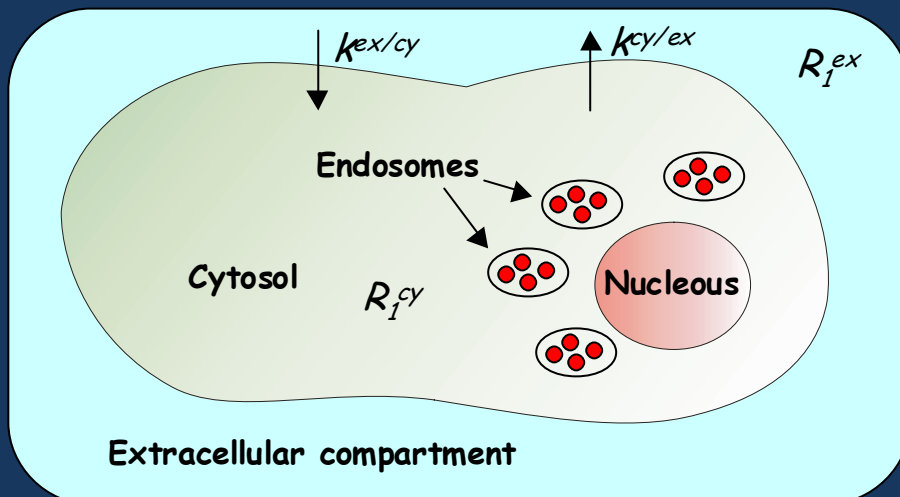


Electroporation

Two compartments (cytosol/extracellular)

$$|k^{cy/ex} + k^{ex/cy}| > |R_1^{cy} - R_1^{ex}|$$

no r_1 quenching !



Pinocytosis

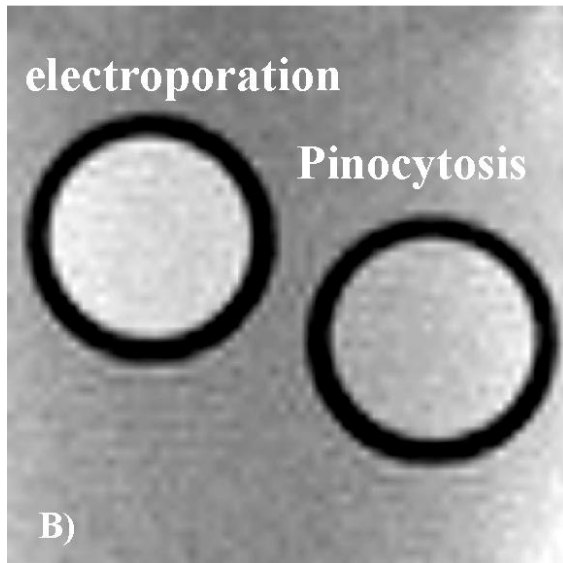
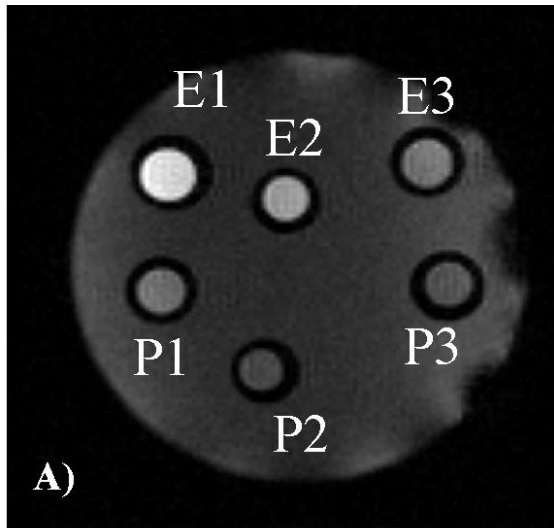
Three compartments

(endosomes/cytosol/extracellular)

$$|k^{cy/end} + k^{end/cy}| < |R_1^{end} - R_1^{cy}|$$

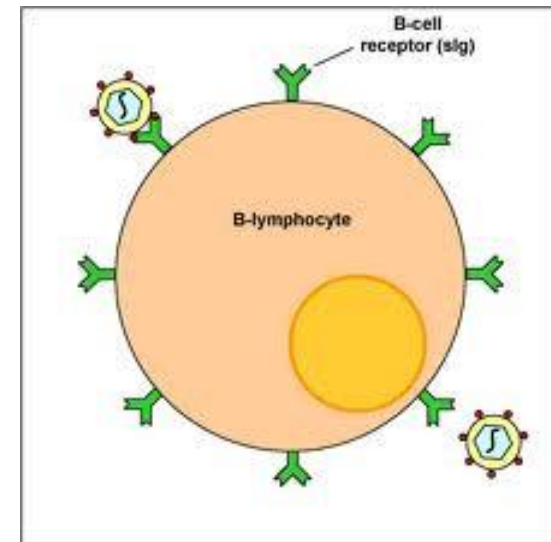
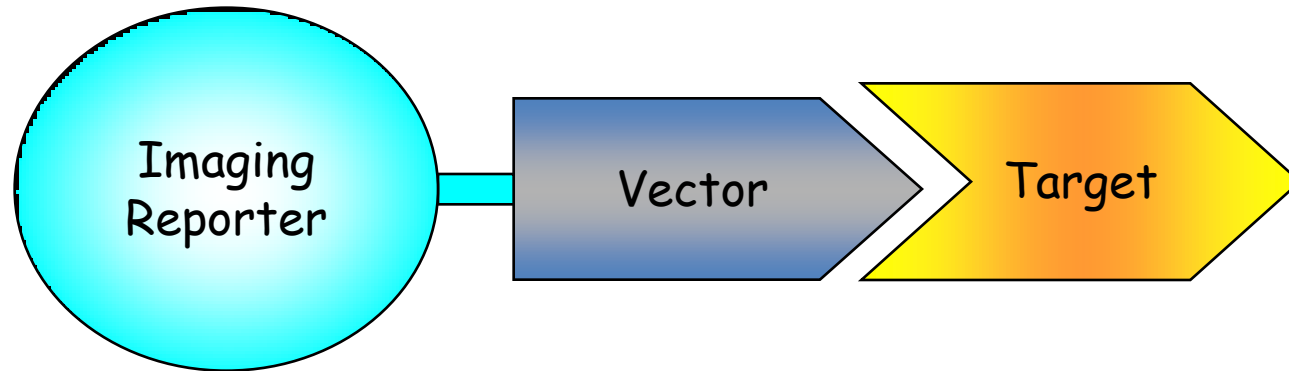
$$|k^{cy/ex} + k^{ex/cy}| > |R_1^{cy} - R_1^{ex}|$$

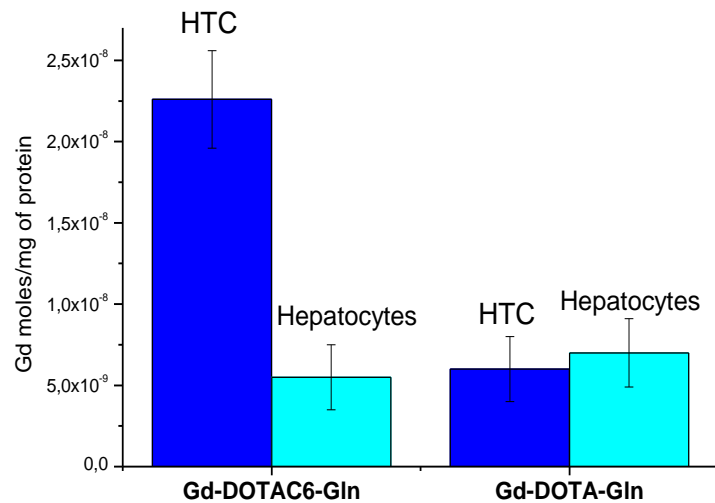
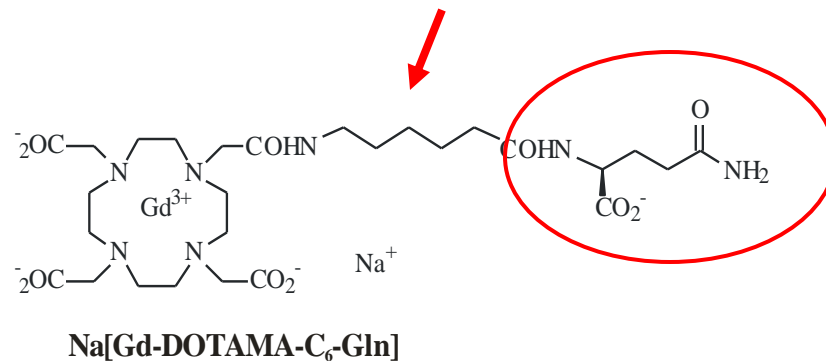
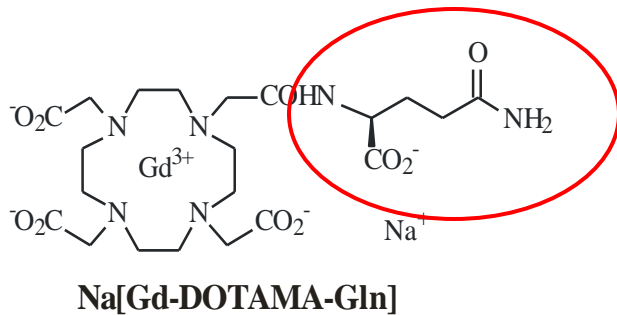
r_1 quenching !



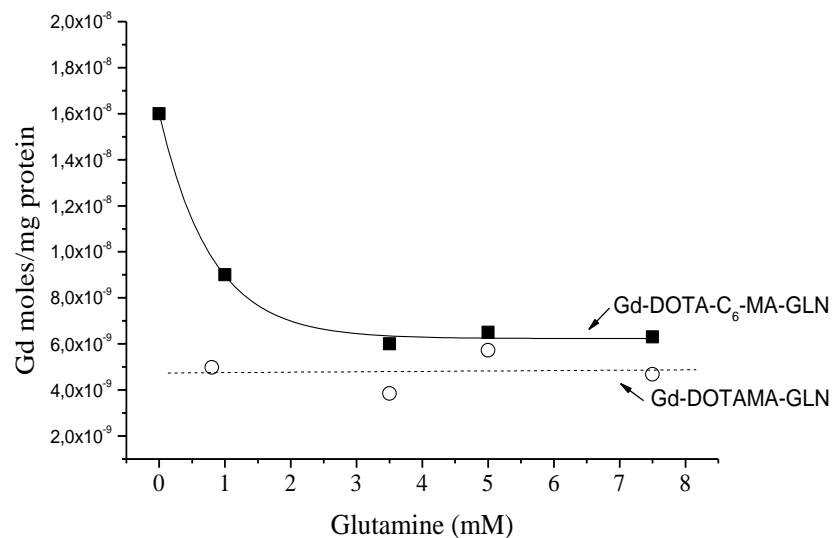
Minimum number of cells detectable $\times \mu\text{l} =$ 5000 Pinocytosis
500 Electroporation

Affinity of the Imaging Probe for the target receptor





Competition with glutamine in HTC cells



Strategies for cellular labeling

```
graph TD; A([Strategies for cellular labeling]) --> B[“IN VIVO”]; A --> C[“IN VITRO”]; B --- D[Labelling of targeted cells and visualization of cellular processes in living organisms.]; C --- E[Monitoring of migration and location of transplanted cells labeled “in vitro” with contrast agents.]
```

“IN VIVO”

Labelling of targeted cells and visualization of cellular processes in living organisms.

“IN VITRO”

Monitoring of migration and location of transplanted cells labeled “in vitro” with contrast agents.

“In vivo” Cell Tracking

- The use of stem and progenitor cells in human clinical Studies will require a technique that can monitor their tissue biodistribution noninvasively
- Whole body imaging will be useful to study the biodistribution of injected cells and help reveal undesired seeding in nontarget organs.

Characteristics of the Ideal Imaging Agent

Real-time visualization of injected cells

Cell quantification

Long term, serial traceability

Single cell sensitivity in any location

Less false-positive imaging

Minimal or no transfer of contrast agent to other cells

MRI Contrast Agents



paramagnetic species



Complexes of
paramagnetic metal ions
(Gd^{3+} , Mn^{2+})

-Positive Contrast

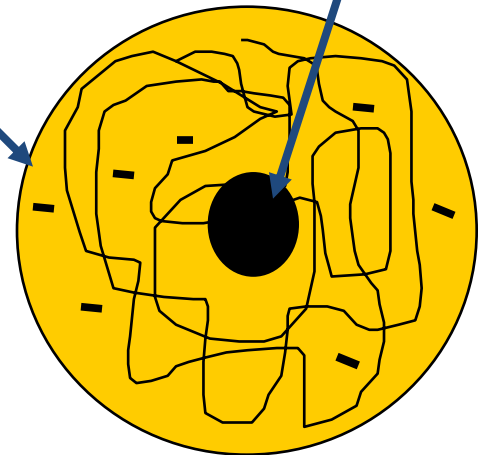
Superparamagnetic
particles (SPIOs, USPIOs)

-Negative Contrast

Iron Oxide Nanoparticles

Schematic view

- **Iron Oxide cristal core**
 - **Superparamagnetic properties**
- **Hydrophilic Coating + charge**
 - **Biological behavior**



**Monitoring of implanted stem cell migration in vivo:
A highly resolved in vivo magnetic resonance imaging
investigation of experimental stroke in rat**

Mathias Hoehn et Al. *PNAS* December 10, 2002, 99 no. 25 16267–16272

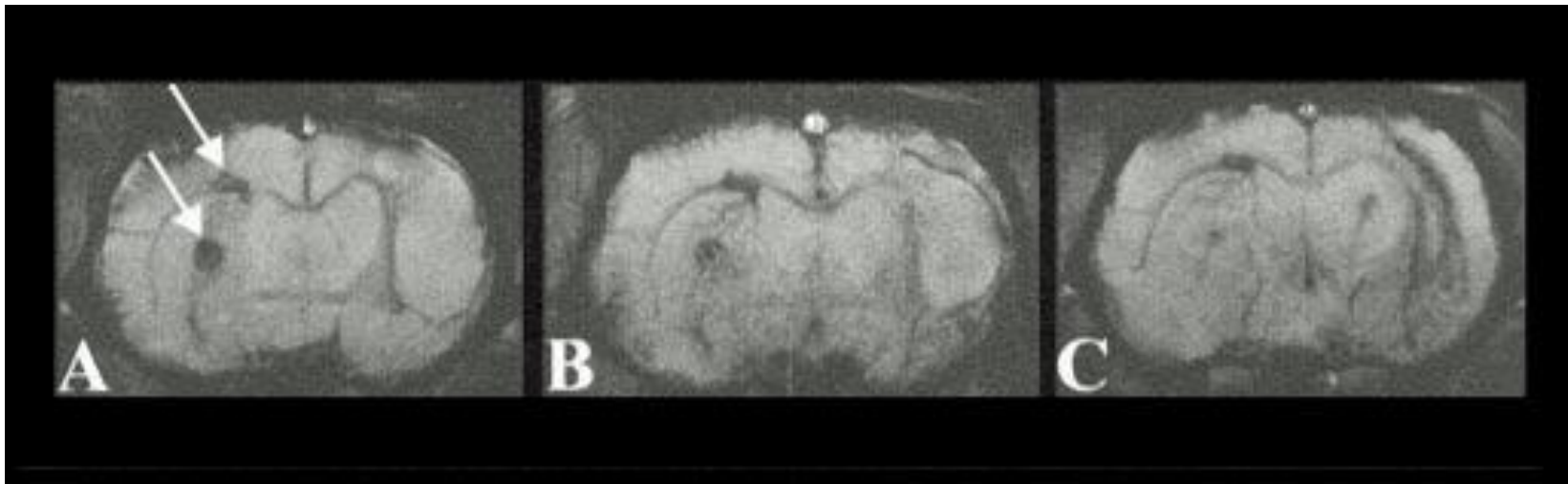
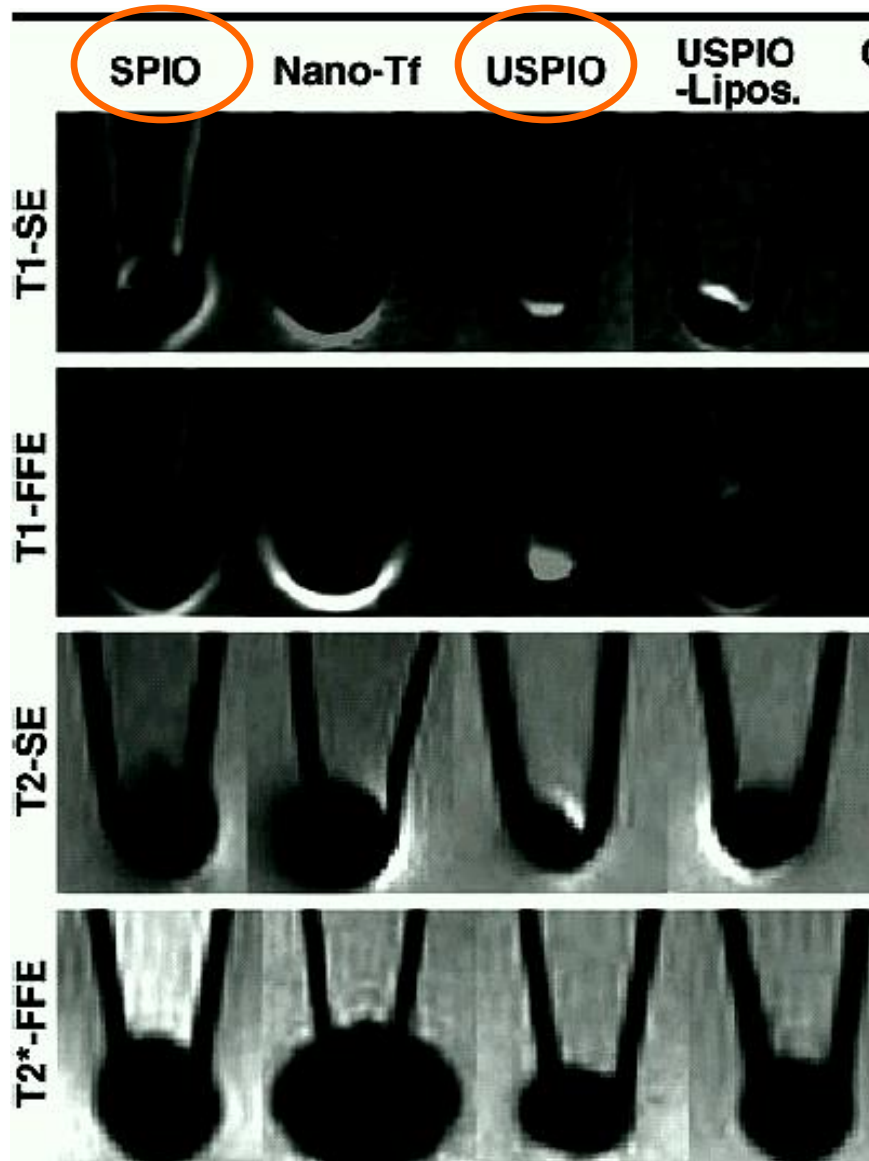


Fig. 2. Coronal sections from a 2D multislice MRI experiment through a rat brain at the day of stem cell implantation (A) and 8 days (B) and 16 days (C) after implantation. Transient focal cerebral ischemia of the right hemisphere (60 min) had been induced 14 days before implantation.

MR signal with SPIO-USPIO



Cells pellet in test tubes
Incubation with SPIO-USPIO

T1, T2 and T2* images

T2*

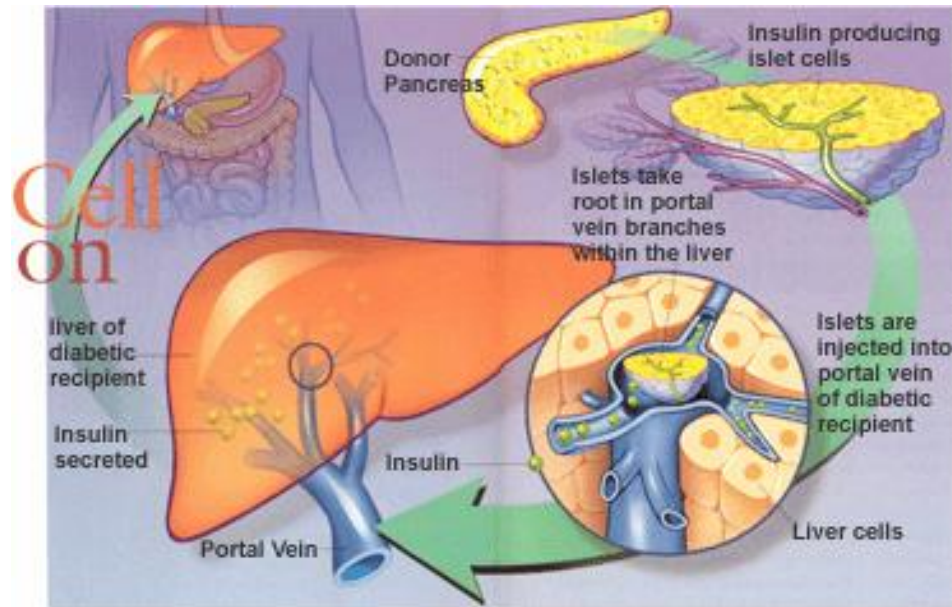
**Good for sensitivity,
Macrophages uptake
Bad for spatial resolution
High field is needed**

Susceptibility artifacts

HE Daldrup et al. *Radiology*, 2003

Bulte JWM Krathchman *NMR in BIOMEDICINE* 2004

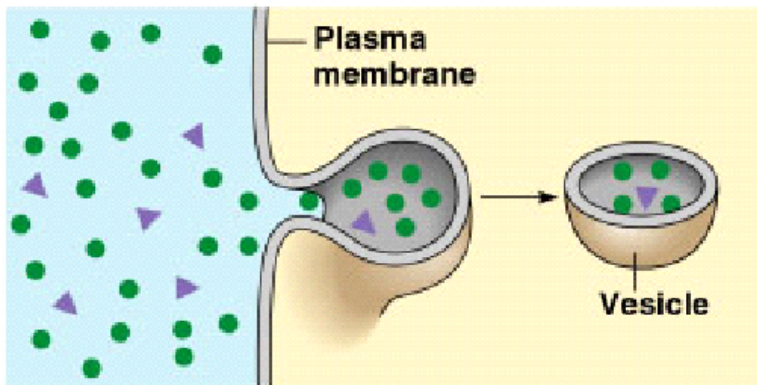
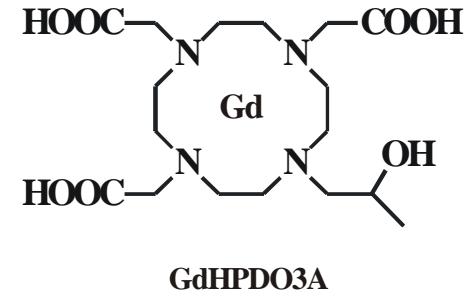
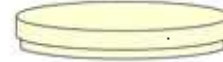
- **Islet transplantation** is now considered as a viable routine option for treatment of insulin-dependent diabetes mellitus but.....



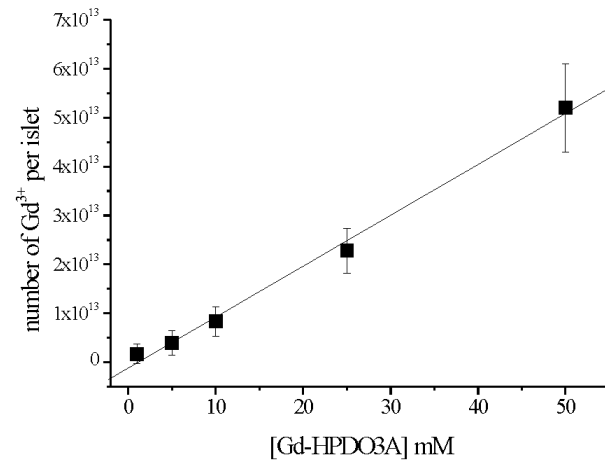
.....the first phases of an islet transplant are critical and the possibility of initial monitoring of the engraftment only possible throughout indirect functional data, representing a severe limitation for research and clinical transplantation.

LABELLING PROCEDURE

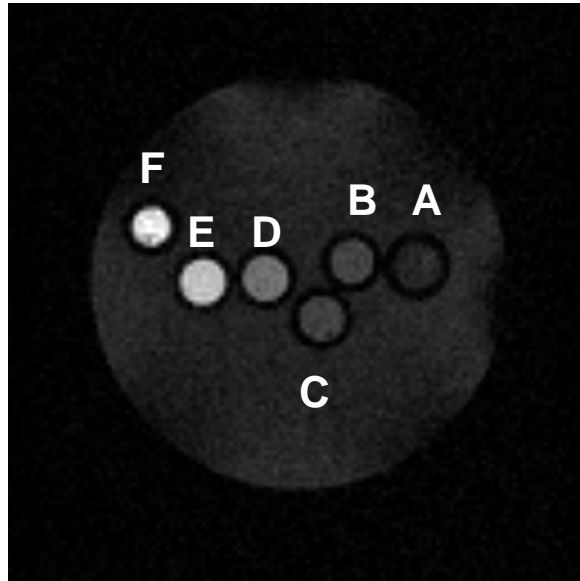
About 1000 islets were incubated for 16 hours with increasing concentrations of the complex (1-50mM)



(b) Pinocytosis

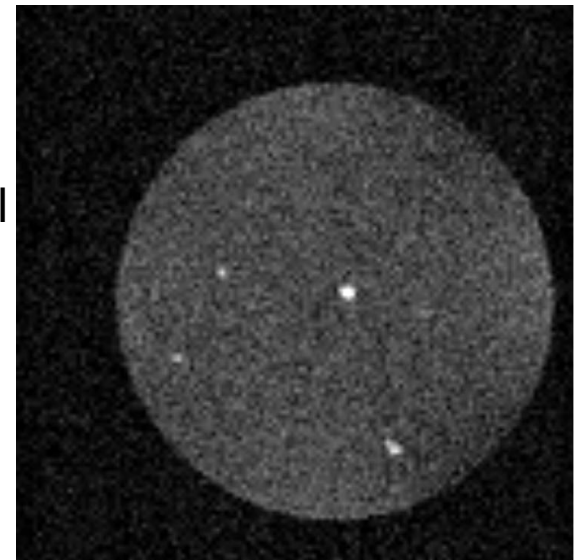
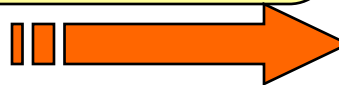


T_1 weighted spin echo image of glass capillaries placed in a agar phantom containing:

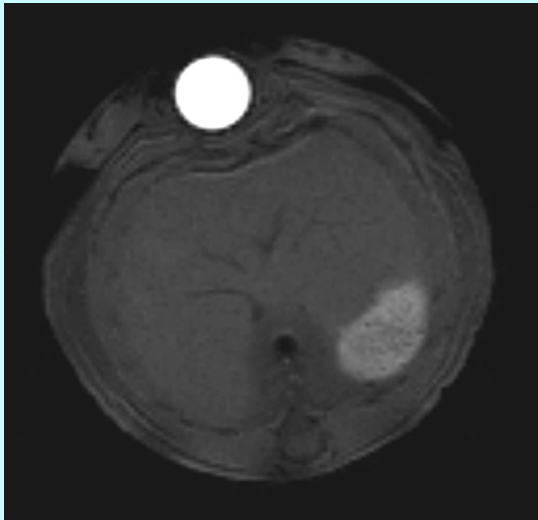


A) human islets incubated with GdHPDO3A at increasing concentrations. Each pellet (ca. 1000 islets) was incubated with Gd-HPDO3A: 0 (A), 1 (B), 5 (C), 10 (D), 25 (E) 50 (F) mM respectively.

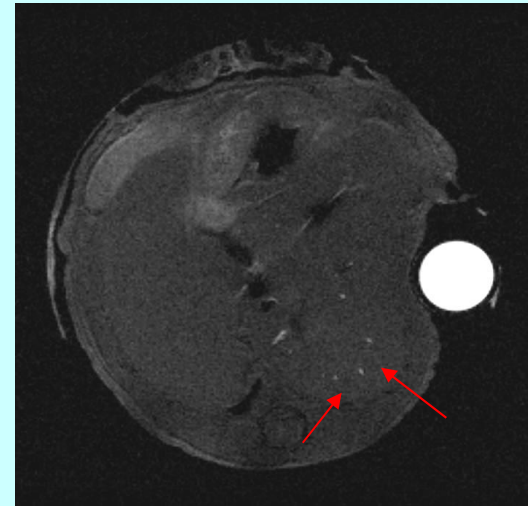
200 IEQ of human islets dispersed in about 200 μ l of AGAR (each hyperintense spot corresponds to the signal deriving from one islet).



T1 weighted spin echo images of SCID mice transplanted into the liver via portal vein with human islets and analyzed one day after transplantation.



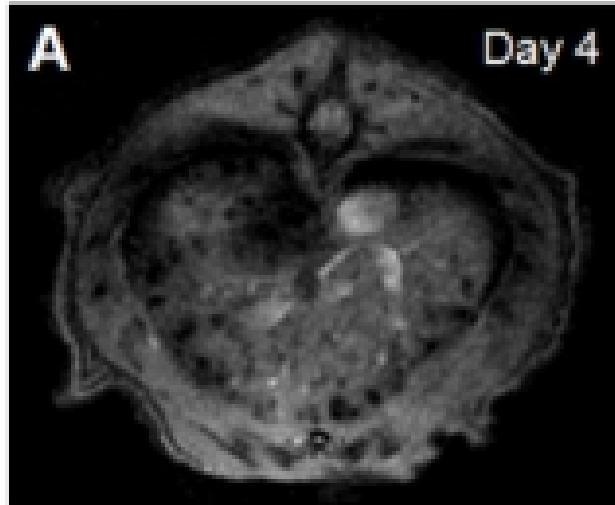
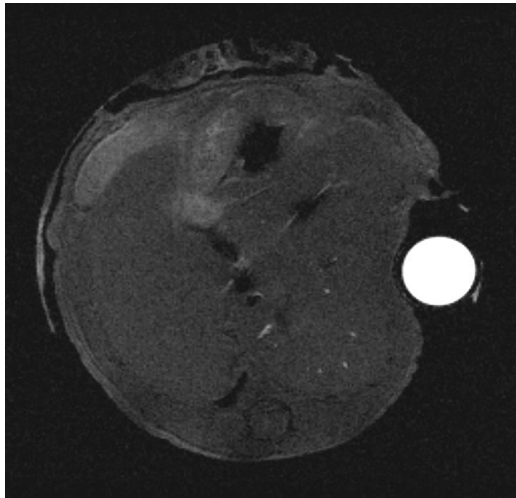
Mouse liver with unlabeled islets



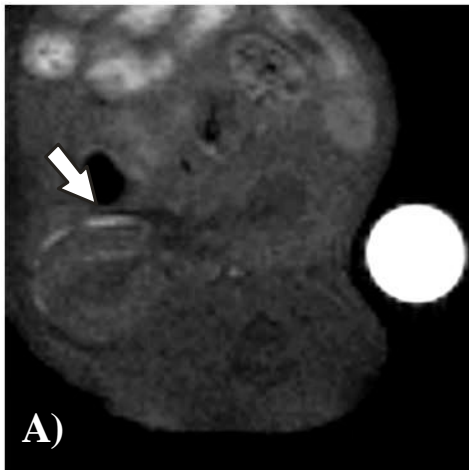
Mouse liver with islets labeled with 50 mM Gd-HPDO3A for 16 hrs in culture prior to infusion

Spin-echo T1-weighted images: TR/TE/NEX 260/4.4/64

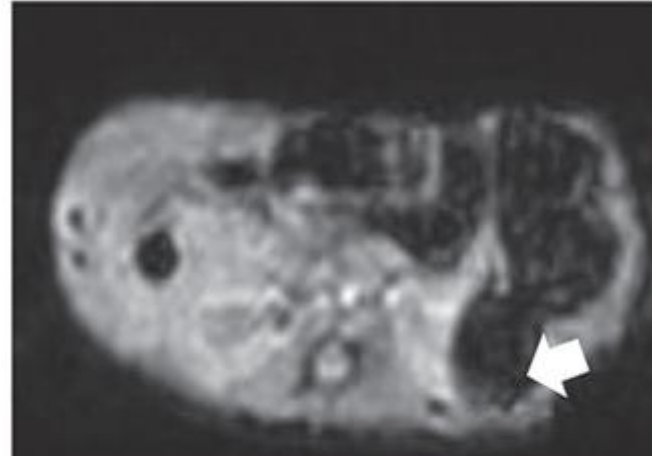
Comparison between Gd and iron-oxide labeled islets



LIVER



Gd



KIDNEYS

Fe

Introduction to the practical session.

Cellular Labelling by receptor mediated Endocytosis

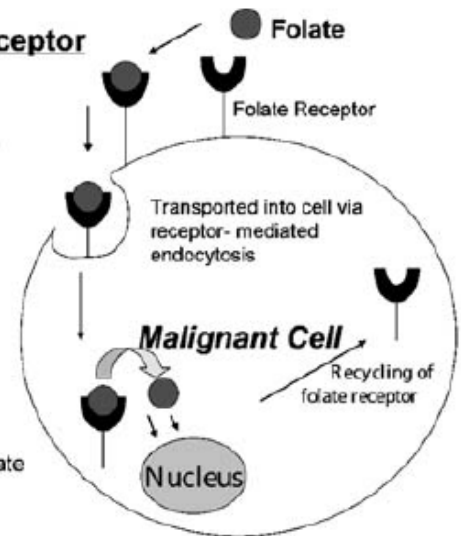
❖ Target: Cells over expressing Folate receptors (IGROV Human ovarian cancer Cells)

❖ Vector: Folic acid

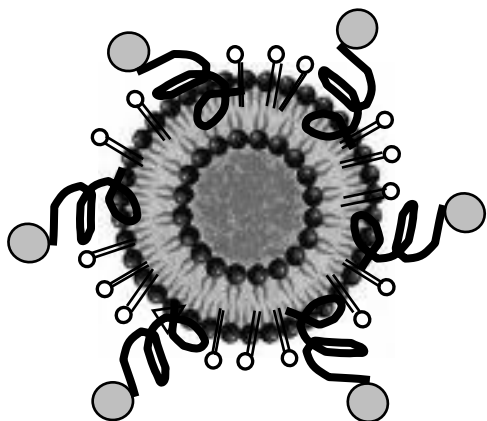
❖ MRI Probe Gd-loaded Liposomes

Membrane Folate Receptor

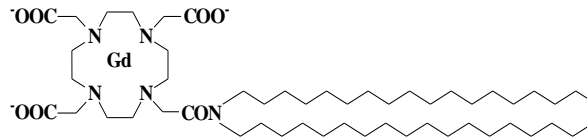
- over-expressed in certain malignancies (ovary, breast, prostate, mesothelioma, osteosarcoma).
- high affinity for folate
- low affinity for reduced folates
- low affinity for anti-folates (chemotherapeutic agents)
- can transport folate and folate linked tracers and drugs



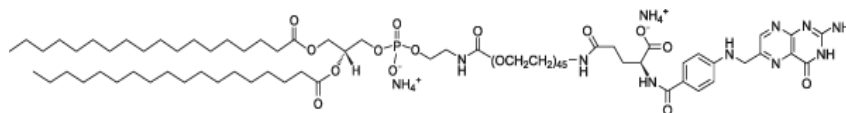
Folic acid Targeted Liposomes



 = Amphiphilic Gd-complex



 = Folate Fospholipid



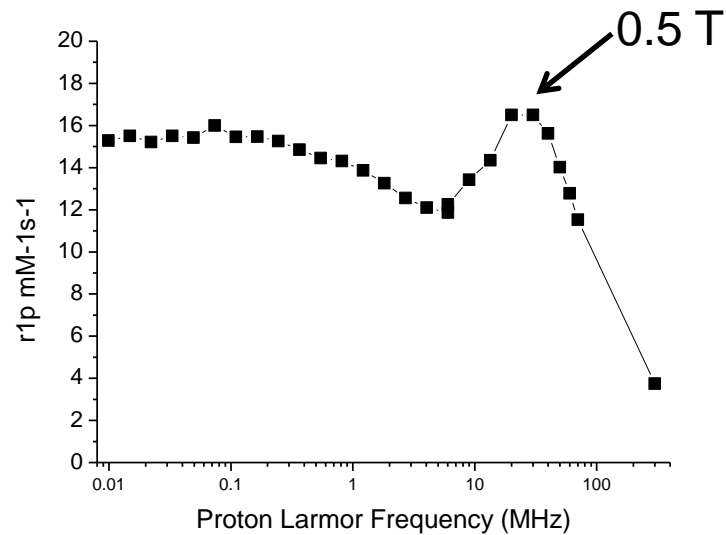
Liposome Formulation

| | |
|---------------------|-----|
| POPC | 59% |
| CHOLESTEROL | 23% |
| Gd-COMPLEX | 15% |
| DSPE-PEG2000 | 2% |
| DSPE-PEG2000-FOLATE | 1% |

r_{1p} ($\text{mM}^{-1}\text{sec}^{-1}$): 16.5

Size (nm): ≈ 150

NMRD Profile



Uptake experiment:

- Cells were incubated in the presence of FA- targeted and non-targeted liposomes 24h before at 37°C, 5% CO₂ at the same concentration.
- Washing with PBS
- Dethaching with trypsin/EDTA
- Re-suspension of cell pellets within 100 µl of PBS and transferring in glass capillaries for MRI measurements.